

Annex 3

Publications

Review Article

Possible Involvement of TLRs and Hemichannels in Stress-Induced CNS Dysfunction via Mastocytes, and Glia Activation

Adam Aguirre,^{1,2} Carola J. Maturana,^{1,2} Paloma A. Harcha,^{1,2} and Juan C. Sáez^{1,2}

¹ Departamento de Fisiología, Pontificia Universidad Católica de Chile, Santiago, Chile

² Instituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Valparaíso, Chile

Correspondence should be addressed to Adam Aguirre; aaguirred@uc.cl

Received 22 February 2013; Revised 16 May 2013; Accepted 11 June 2013

Academic Editor: Dennis Daniel Taub

Copyright © 2013 Adam Aguirre et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

In the central nervous system (CNS), mastocytes and glial cells (microglia, astrocytes and oligodendrocytes) function as sensors of neuroinflammatory conditions, responding to stress triggers or becoming sensitized to subsequent proinflammatory challenges. The corticotropin-releasing hormone and glucocorticoids are critical players in stress-induced mastocyte degranulation and potentiation of glial inflammatory responses, respectively. Mastocytes and glial cells express different toll-like receptor (TLR) family members, and their activation via proinflammatory molecules can increase the expression of connexin hemichannels and pannexin channels in glial cells. These membrane pores are oligohexamers of the corresponding protein subunits located in the cell surface. They allow ATP release and Ca^{2+} influx, which are two important elements of inflammation. Consequently, activated microglia and astrocytes release ATP and glutamate, affecting myelination, neuronal development, and survival. Binding of ligands to TLRs induces a cascade of intracellular events leading to activation of several transcription factors that regulate the expression of many genes involved in inflammation. During pregnancy, the previous responses promoted by viral infections and other proinflammatory conditions are common and might predispose the offspring to develop psychiatric disorders and neurological diseases. Such disorders could eventually be potentiated by stress and might be part of the etiopathogenesis of CNS dysfunctions including autism spectrum disorders and schizophrenia.

1. Introduction

Signaling between nervous and immune systems is in part due to the fact that these two systems share ligands and receptors. The cellular components involved in these interactions within the central nervous system (CNS) are mainly mastocytes, also called mast cells, and glia. In human brain, mastocytes are very scarce and are preferentially located in perivascular territories. By contrast, glial cells comprise about 90% of the total cell content in the CNS and are classified as microglia and macroglia (astrocytes, oligodendrocytes, and ependymal cells) [1]. Representative of the immune system in the CNS are mastocytes and microglia, two cell types derived from hematopoietic cells of the bone marrow that migrate to the brain before closure of the blood brain barrier (BBB) [2, 3].

The CNS challenged by different aggressions frequently elicits immune and inflammatory responses [4, 5]. Mastocytes and microglia are efficient sensors of adverse endogenous or exogenous conditions of the CNS [2, 6]. Moreover, stress conditions induce rapid mastocyte degranulation via the hypothalamic peptide corticotropin-releasing hormone (CRH) [7] and exogenous danger molecules like polyinosinic-polycytidylic acid (poly (I:C)), bacterial lipopolysaccharide (LPS), and peptidoglycan (PGN), which are detected by mastocytes and microglia via toll-like receptors (TLRs) [8, 9]. Also, glucocorticoids (GCs) play a relevant role in stress-induced potentiation of neuroinflammatory responses by sensitizing microglia to proinflammatory stimuli [10]. As part of these responses, glial TLRs, connexin hemichannels (Cx HCs), pannexin (Panx) channels might be key players in acute and chronic neurodegenerative diseases

FasL-Triggered Death of Jurkat Cells Requires Caspase 8-Induced, ATP-Dependent Cross-Talk Between Fas and the Purinergic Receptor P2X₇

ADAM AGUIRRE,¹ KENJI F. SHOJI,³ JUAN C. SÁEZ,^{3,4} MAURICIO HENRÍQUEZ,^{1,2*}
AND ANDREW FG QUEST^{1*}

¹Facultad de Medicina, Centro de Estudios Moleculares de la Célula, Universidad de Chile, Santiago, Chile

²Facultad de Medicina, Programa Fisiología y Biofísica (ICBM), Universidad de Chile, Santiago, Chile

³Departamento de Fisiología, Pontificia Universidad Católica de Chile, Santiago, Chile

⁴Instituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Valparaíso, Chile

Fas ligation via the ligand FasL activates the caspase-8/caspase-3-dependent extrinsic death pathway. In so-called type II cells, an additional mechanism involving tBid-mediated caspase-9 activation is required to efficiently trigger cell death. Other pathways linking FasL–Fas interaction to activation of the intrinsic cell death pathway remain unknown. However, ATP release and subsequent activation of purinergic P2X₇ receptors (P2X₇Rs) favors cell death in some cells. Here, we evaluated the possibility that ATP release downstream of caspase-8 via pannexin I hemichannels (Panx1 HCs) and subsequent activation of P2X₇Rs participate in FasL-stimulated cell death. Indeed, upon FasL stimulation, ATP was released from Jurkat cells in a time- and caspase-8-dependent manner. Fas and Panx1 HCs colocalized and inhibition of the latter, but not connexin hemichannels, reduced FasL-induced ATP release. Extracellular apyrase, which hydrolyzes ATP, reduced FasL-induced death. Also, oxidized-ATP or Brilliant Blue G, two P2X₇R blockers, reduced FasL-induced caspase-9 activation and cell death. These results represent the first evidence indicating that the two death receptors, Fas and P2X₇R connect functionally via caspase-8 and Panx1 HC-mediated ATP release to promote caspase-9/caspase-3-dependent cell death in lymphoid cells. Thus, a hitherto unsuspected route was uncovered connecting the extrinsic to the intrinsic pathway to amplify death signals emanating from the Fas receptor in type II cells.

J. Cell. Physiol. 228: 485–493, 2013. © 2012 Wiley Periodicals, Inc.

Apoptosis is an active process that depends on a defined sequence of signaling events and can occur via two different routes, referred to as the extrinsic (death receptor mediated) and the intrinsic (endogenous) pathways. The extrinsic pathway is activated in a manner dependent on ligand binding to death receptors. Fas ligand (FasL) binding to the Fas/CD95 receptor promotes formation of the death-induced signaling complex (DISC) by recruiting Fas-associating death domain (DD) [FADD-containing protein] through its DD domain, which then recruits death-effector domain (DED)-containing initiator caspases, such as caspase-8 and -10. Proximity of pro-caspase molecules is required for autocleavage and activation of initiator caspases, which then go on to proteolytically cleave and activate executioner caspases, including caspase-3, -6, -7 (Scaffidi et al., 1998; Peter and Krammer, 2003).

In some cells, referred to as Type I, this sequence of events is sufficient to trigger apoptosis. Alternatively in Type II cells, caspase-8 activation is weak and signaling downstream of the DISC needs to be amplified via a loop involving cleavage of Bid to generate tBid and activation of the mitochondrial pathway, which generally responds to intracellular stress signals. Changes in mitochondrial membrane potential and mitochondrial membrane permeability induced downstream of tBid formation culminate in release of mitochondrial molecules to the cytosol. Mitochondrial cytochrome c in conjunction with cytosolic apoptosis protease activating factor 1 (Apaf-1) and adenosine triphosphate (ATP) assemble into a multi-protein complex, called the apoptosome, which catalyzes proteolytic auto-activation of the initiator caspase-9 and activation downstream

of the executioner caspases (Hengartner, 2000; Samraj et al., 2006).

The purinergic receptor P2X₇ (P2X₇R), whose ligand is ATP, can trigger either necrosis and/or apoptosis in various cell types, including myeloid cells, dendritic cells, thymocytes, macrophages and lymphocytes (Di Virgilio et al., 1998).

The authors declare no conflict of interest.

Additional supporting information may be found in the online version of this article.

Contract grant sponsor: FONDECYT-FONDAP;

Contract grant number: 15010006.

Contract grant sponsor: FONDECYT;

Contract grant number: 1090071.

Contract grant sponsor: FONDECYT-Postdoctoral Fellowship;

Contract grant number: 3070045.

Contract grant sponsor: ANILLO;

Contract grant number: ACT-71.

*Correspondence to: Mauricio Henríquez and Andrew FG Quest, Facultad de Medicina, Universidad de Chile, Independencia 1027 Santiago, Chile. E-mail: mhenriqu@med.uchile.cl; aquest@med.uchile.cl

Manuscript Received: 24 April 2012

Manuscript Accepted: 3 July 2012

Accepted manuscript online in Wiley Online Library (wileyonlinelibrary.com): 17 July 2012.

DOI: 10.1002/jcp.24159

**Sección 3.2. – Nuevos
conocimientos biomédicos y
fisiológicos de la práctica
deportiva (con énfasis en canales
de iones en músculo esquelético)**

Marcus Vinicius Almeida Campos¹, Carlos Costá², Carlos González León³, Ramón Latorre³, Verónica Mileti⁴ y Gonzalo Ferreira².

1. Depto de Educação Física, Faculdades
Euclides da Cunha-FEUC, Faculdade de
Filosofia Ciências e Letras de São José do
Rio Pardo, Brasil. Programa ProInBio.
UdelaR-REMH, Uruguay

2. Laboratorio de Canales Iónicos,
Departamento de Biofísica, Facultad de
Medicina, Universidad de la República,
Montevideo, Uruguay.

3. Centro de Neurociencias de Valparaíso,
Universidad de Valparaíso, Chile.

4. Departamento de Fisiopatología, Facultad
de Ciencias Exactas, Universidad Nacional
de La Plata, Argentina.

El movimiento y la vida están interconectados desde el punto de vista biológico, yendo desde fundamentos de fenomenología y de la observación, hasta descripciones que nos han permitido en años recientes un entendimiento cada vez mayor de sus bases moleculares y fisiológicas. Se describen distintos niveles de movimiento que van desde actividad física a deporte. En esta sección procuraremos definir estos conceptos para luego centrarnos en las modificaciones que se

producen a nivel de canales iónicos en el músculo esquelético durante el ejercicio.

3.2.1. Actividad física, ejercicio y deporte, precisiones del lenguaje e importancia a nivel molecular y de la vida y envejecimiento.

Puede ocurrir movimiento en diferentes niveles, que conocemos como actividad física, ejercicio físico o deporte, de acuerdo con sus variaciones de forma, intensidad y frecuencia. Estos tres niveles de movimiento llevan a modificaciones moleculares, celulares y fisiológicas de acuerdo con la forma como son realizados. La comprensión del comportamiento de estas variables ha mejorado mucho con el correr de los años, debido al avance tecnológico. Así entonces, el movimiento o actividad física en general en el campo biológico puede manifestarse a varios niveles entrelazados que conocemos como actividad física, ejercicio o deporte, de acuerdo a sus variantes en formas, intensidad, competitividad y periodicidad de prácticas al respecto. En sí pueden considerarse distintos escalones de conjuntos inclusivos (ver Fig. 1). La **actividad física** puede ser definida como cualquier movimiento corporal producido por la musculatura esquelética, que resulta en un gasto energético por encima de los niveles de reposo. Se ha determinado que la misma es central a la hora de definir los

Emerging Model Organisms

Octodon degus (Molina 1782): A Model in Comparative Biology and Biomedicine

Alvaro O. Ardiles,¹ John Ewer,¹ Monica L. Acosta,² Alfredo Kirkwood,³ Agustin D. Martinez,¹ Luis A. Ebensperger,⁴ Francisco Bozinovic,⁴ Theresa M. Lee,⁵ and Adrian G. Palacios^{1,6}

¹Centro Interdisciplinario de Neurociencia de Valparaíso, Facultad de Ciencias, Universidad de Valparaíso, 2360102 Valparaíso, Chile; ²Department of Optometry and Vision Science, University of Auckland, Auckland 1142, New Zealand; ³Mind/Brain Institute and Department of Neurosciences, Johns Hopkins University, Baltimore, Maryland 21218; ⁴Departamento de Ecología, Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Santiago 6513677, Chile; ⁵Department of Psychology and Neuroscience Program, University of Michigan, Ann Arbor, Michigan 48109

One major goal of integrative and comparative biology is to understand and explain the interaction between the performance and behavior of animals in their natural environment. The Caviomorph, *Octodon degu*, is a native rodent species from Chile, and represents a unique model to study physiological and behavioral traits, including cognitive and sensory abilities. Degus live in colonies and have a well-structured social organization, with a mostly diurnal–crepuscular circadian activity pattern. More notable is the fact that in captivity, they reproduce and live between 5 and 7 yr and show hallmarks of neurodegenerative diseases (including Alzheimer's disease), diabetes, and cancer.

BACKGROUND

The Octodontidae family (Rodentia) is endemic to South America and shows a wide range of diversity, from its genes to its communities. Octodontidae fossil records go back to the last Eocene, around 40 million years ago. Currently, they are dispersed in north and central Chile from sea level to ~3500 m altitude, with microhabitats in matorral (bush) with evergreen sclerophyllous shrubs over a seasonal herbaceous layer. This environment forms a characteristic South American scrub ecosystem with diverse flora and fauna. The Octodontidae includes nine species (Spotorno et al. 1995), with a unique morphological gradient ranging from cursorial/generalist to subterranean. The number of chromosomes varies among Octodontidae species from 38 (2N) to 102 and the phylogenetic relationship of Octodontidae has been established by cladistics analysis (Honeycutt et al. 2003). The monophyletic *Octodon* genus has three species: The common *O. degu*, *O. lunatus*, and *O. bridgesi*. All are endemic to Chile and related to the Chinchilloidea and Caviioidea families (Opazo 2005). *Octodon degu* is a small, diurnal, herbivorous rodent that lives in social groups (see Fig. 1).

SOURCES AND HUSBANDRY

The accompanying protocol, **Husbandry and Breeding in the *Octodon degu* (Molina 1782)** (Palacios and Lee 2013), describes successful husbandry and breeding practices based on the experience of the University of Michigan *degu* colony.

⁶Correspondence: adrian.palacios@uv.cl

© 2013 Cold Spring Harbor Laboratory Press

Cite this article as *Cold Spring Harb Protoc*; 2013; doi:10.1101/pdb.emo071357

Properties, Projections, and Tuning of Teleost Olfactory Receptor Neurons

Alejandra Bazáes · Jesús Olivares · Oliver Schmachtenberg

Received: 11 September 2012 / Revised: 17 December 2012 / Accepted: 1 February 2013
© Springer Science+Business Media New York 2013

Abstract In many fishes, the olfactory sense participates in such vital processes as feeding, reproduction, orientation, and predator avoidance. In teleosts, these tasks are fulfilled by a single type of olfactory organ for odorant and pheromone detection, containing ciliated and microvillus receptor neurons, and olfactory crypt cells. Recently, progress was made in understanding crypt cell function with the discovery of a V1R-like odorant receptor expressed in this neuron, an analysis of crypt cell odorant tuning properties, and the dissection of crypt cell connectivity within the telecephalon. Here, we review recent findings on the molecular properties, functions, and associated neural pathways of the three types of teleost olfactory receptor neurons with special emphasis on the crypt cell, and evaluate their roles in the detection of food, social and sexual odorants.

Keywords Olfaction · Smell · Pheromone · Crypt cell · Odorant receptor · Fish

Introduction

About 50 % of all vertebrates on this planet are teleost fishes. As other vertebrates, teleosts have an olfactory system that allows them to detect and discriminate between different kinds of olfactory stimuli, such as amino acids, nucleotides, bile salts, gonadal steroids, and prostaglandins (Sorensen and Caprio, 1998). These distinct odorant classes are related to different behaviors, such as feeding, orientation, and reproduction in fishes. Due to their aqueous habitat, compounds

that elicit chemosensory responses in fishes are mostly hydrophilic, and may stimulate either the olfactory or the taste system or both. Traditionally, substances effective at sub-micromolar concentrations have been considered as odorants, and those with higher functional thresholds as tastants, but this distinction is invalid in fishes, whose taste system is as or more sensitive to certain substances as their olfactory system (Ogawa and Caprio, 2010; Hara, 2011). Notwithstanding this caveat, a long series of behavioral and electrophysiological experiments in different teleost species has provided us with a large body of data on fish olfaction, revealing sometimes astonishing sensitivity and functional sophistication. This review provides an overview of the current knowledge regarding the cellular and molecular properties, odorant tuning, and neural connections of the three types of olfactory receptor neurons (ORNs) in teleosts, with an emphasis on recent findings on the physiology of the olfactory crypt cell (CC).

Teleost ORNs

As opposed to the majority of vertebrates, teleosts lack both a vomeronasal organ and an accessory olfactory bulb. Instead, they have a single pseudostratified olfactory epithelium on each side of the head (Fig. 1a, b), generally forming a multilamellar olfactory rosette, although single plain epithelial sheets also can be found in some species (Hansen and Zielinski, 2005). Three morphologically distinct types of ORN are present in the olfactory epithelium: Ciliated and microvillus bipolar neurons (cORNs and mORNs), and the CCs, which are thought to be absent from any other vertebrate group (Hansen and Finger, 2000; Laberge and Hara, 2001) (Fig. 1c–f). In addition, undifferentiated basal cells in the olfactory epithelium allow the regeneration of sensory neurons and accessory cells, and the growth of the organ with age (Evans et al., 1982) (Fig. 1g).

A. Bazáes · J. Olivares · O. Schmachtenberg (✉)
Centro Interdisciplinario de Neurociencia de Valparaíso CINV,
Facultad de Ciencias, Universidad de Valparaíso,
Avda. Gran Bretaña 1111, Playa Ancha,
2360102 Valparaíso, Chile
e-mail: oliver.schmachtenberg@uv.cl

Quantitative Analysis of Cell Migration Using Optical Flow

Katica Boric¹, Patricio Orio¹, Thierry Viéville², Kathleen Whitlock^{1*}

1 Centro Interdisciplinario de Neurociencia, Facultad de Ciencias, Universidad de Valparaíso, Valparaíso, Chile, **2** Inria, Mnémosyne, Sophia Antipolis, France

Abstract

Neural crest cells exhibit dramatic migration behaviors as they populate their distant targets. Using a line of zebrafish expressing green fluorescent protein (*sox10:EGFP*) in neural crest cells we developed an assay to analyze and quantify cell migration as a *population*, and use it here to characterize in detail the subtle defects in cell migration caused by ethanol exposure during early development. The challenge was to quantify changes in the *in vivo* migration of all *Sox10:EGFP* expressing cells in the visual field of time-lapse movies. To perform this analysis we used an Optical Flow algorithm for motion detection and combined the analysis with a fit to an affine transformation. Through this analysis we detected and quantified significant differences in the cell migrations of *Sox10:EGFP* positive cranial neural crest populations in ethanol treated versus untreated embryos. Specifically, treatment affected migration by increasing the left-right asymmetry of the migrating cells and by altering the direction of cell movements. Thus, by applying this novel computational analysis, we were able to quantify the movements of populations of cells, allowing us to detect subtle changes in cell behaviors. Because cranial neural crest cells contribute to the formation of the frontal mass these subtle differences may underlie commonly observed facial asymmetries in normal human populations.

Citation: Boric K, Orio P, Viéville T, Whitlock K (2013) Quantitative Analysis of Cell Migration Using Optical Flow. PLoS ONE 8(7): e69574. doi:10.1371/journal.pone.0069574

Editor: Brian Key, School of Biomedical Sciences, The University of Queensland, Australia

Received: April 3, 2013; **Accepted:** June 13, 2013; **Published:** July 31, 2013

Copyright: © 2013 Boric et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by NIHR01HD50820 (KEW), ICM Proyecto P09-022-F (PO, KEW, KB), FONDECYT 1111046 (KEW), ANR-47 CONICYT (PO, TV), FONDECYT 1130862 (PO), ACT-1113 (PO), and CONICYT Doctoral Fellowship (KB). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: kathleen.whitlock@uv.cl

Introduction

The cranial neural crest cells (CNCC) are a population of progenitors that give rise to both craniofacial structures and components of the peripheral nervous system (PNS). These cells migrate from the dorsal neural tube to populate the face. After leaving the neuroepithelium, CNCC split into discrete streams separated by CNCC-free regions [1]. Three main streams are formed according to their relative pre-migratory position along the cranial neural tube. The most anterior stream of CNCC migrates dorsal to the eye and populates the frontal mass (or anterior neurocranium) [2], ganglia of cranial nerves and melanocytes among others. More posterior streams of CNCC populate the branchial arches, where they form the cartilage and bones of the jaw [3]. Much is known about the CNCC migration and differentiation as it relates to the branchial arch derivatives including the jaw elements [4,5]. By contrast, the migration of CNCC that pass dorsal to the eye and contribute to the facial structures has not been extensively described with the exception of the migration of neural crest cells that contribute to specific parts of the neurocranium [2,6,7]. We (this study) and others [8] have found that the migration of the dorsal anterior CNCC appears to be sensitive to environmental factors such as alcohol, smoking, and herbicides. Yet, the cell movements that occur during the migration of this population of CNCC are complex and difficult to describe and characterize quantitatively. For this reason we developed an analytical method to characterize cell migration, and

use it here to describe the defects of dorsal anterior CNCC migration caused by alcohol exposure.

There is a large body of evidence demonstrating that ethanol (EtOH) affects CNCC development. The primary defects observed are cell death in both *in vitro* and *in vivo* systems; some studies also report defects in cell migration. *In vivo* EtOH-induced cell death has been reported in regions populated by CNCC in the developing chick embryo [9] and while the number of CNCC is reduced, the migration patterns were unaffected [10]. Additional studies in chick have reported cell death in neural crest cells [11]. *In vitro* analysis of CNCC has shown that EtOH exposure produces permanent changes in cell shape, surface morphology, migration, and cell death [12,13]. Interestingly no analysis of migration defects have been reported *in vivo*, although histological studies have suggested the occurrence of EtOH-induced migration defects, based on the pattern of immunostained neural crest in treated chick embryos [14]. Thus, whereas EtOH induced cell death has consistently been reported in a variety of model systems, the effects on CNCC migration have not been well documented. An *in vivo* real time analysis of the CNCC population contributing to the frontal mass would improve our understanding of CNCC migration in normal and EtOH exposed animals.

Many studies of cell migration use techniques that track single cells and base their analysis on the average migration behavior of single cells. In zebrafish and *Xenopus* embryos the cell-cell interactions of individual CNCC have been analyzed to demonstrate contact inhibition of locomotion in which cells cease migrating after contact with another cell [15]. Because the CNCC



Measurement of autophagy flux in the nervous system *in vivo*

K Castillo^{1,2}, V Valenzuela^{1,2}, S Matus³, M Nassif^{1,2}, M Oñate^{1,2}, Y Fuentealba^{1,2}, G Encina^{1,2}, T Irrazabal^{1,2}, G Parsons⁴, FA Court^{3,5}, BL Schneider⁶, D Armentano⁴ and C Hetz^{*,1,2,3,7}

Accurate methods to measure autophagic activity *in vivo* in neurons are not available, and most of the studies are based on correlative and static measurements of autophagy markers, leading to conflicting interpretations. Autophagy is an essential homeostatic process involved in the degradation of diverse cellular components including organelles and protein aggregates. Autophagy impairment is emerging as a relevant factor driving neurodegeneration in many diseases. Moreover, strategies to modulate autophagy have been shown to provide protection against neurodegeneration. Here we describe a novel and simple strategy to express an autophagy flux reporter in the nervous system of adult animals by the intraventricular delivery of adeno-associated viruses (AAV) into newborn mice. Using this approach we efficiently expressed a monomeric tandem mCherry-GFP-LC3 construct in neurons of the peripheral and central nervous system, allowing the measurement of autophagy activity in pharmacological and disease settings.

Cell Death and Disease (2013) 4, e917; doi:10.1038/cddis.2013.421; published online 14 November 2013

Subject Category: Neuroscience

Macroautophagy, here referred to as autophagy, is the major catabolic pathway involved in the degradation of damaged or superfluous organelles, abnormal protein aggregates and other cytosolic components.¹ An explosion of literature in the last few years highlights the contribution of autophagy to diverse physiological processes including metabolic control, lipid homeostasis, immunity and myopathies.² Autophagy is initiated by the formation of autophagosomes, double membrane vesicles that engulf cytosolic components and fuse with endosomes to form hybrid organelles called amphisomes that later fuse with lysosomes where cargoes are degraded. The mechanisms underlying the regulation of different steps of autophagy are highly complex and dynamic and are reviewed in detail elsewhere.^{3,4} In brief, two ubiquitin-like conjugation pathways control autophagy. One of them involves the covalent binding of Atg12 to Atg5, which occurs at the phagophore level, and dissociation after autophagosome consolidation. One of the key steps in the regulation of autophagy is the conjugation of microtubule-associated protein 1 light chain 3 (LC3) to phosphatidylethanolamine to form microtubule-associated protein 1 light chain 3 lipidated form (LC3-II). Lipidated LC3 binds to the expanding phagophore and remains associated with autophagosomes even

after fusion with lysosomes. Then, LC3-II can be either delipidated and recycled or degraded by hydrolytic enzymes at the lysosome. Monitoring LC3-II conversion, LC3 distribution or its flux through the autophagy pathway are the gold standards for measuring autophagy activity and are widely used in the field.⁴

The generation of conditional knockout mice for essential autophagy regulatory components in the nervous system revealed a key role of the pathway in the basal maintenance of protein homeostasis in neurons.^{5,6} Most prevalent neurodegenerative diseases, including Alzheimer's disease, amyotrophic lateral sclerosis, Parkinson's disease and Huntington's disease are associated with the misfolding and aggregation of specific proteins.⁷ Many studies indicate that autophagy operates as an efficient mechanism for the degradation of aggregation-prone proteins linked to neurodegeneration and suggest that pharmacological activation of autophagy offers a promising therapeutic avenue.⁸ Accumulating evidence also suggests that autophagy impairment may underlie the etiology of several neurodegenerative diseases.^{8,9}

One of the main limitations in the field is the lack of reliable tools to monitor autophagy activity in the nervous system,^{4,10} a challenging issue due to barriers for drug penetrance into

¹Institute of Biomedical Sciences, Faculty of Medicine, University of Chile, Santiago, Chile; ²Center for Molecular Studies of the Cell, Program of Cellular and Molecular Biology, Institute of Biomedical Sciences, University of Chile, Santiago, Chile; ³Neurounion Biomedical Foundation, Santiago, Chile; ⁴Department of Molecular Biology, Genzyme Corporation, 49 New York Avenue, Framingham, MA, USA; ⁵Millennium Nucleus for Regenerative Biology, Faculty of Biology, Pontificia Universidad Católica de Chile, Santiago, Chile; ⁶Neurodegenerative Studies Laboratory, Brain Mind Institute, École Polytechnique Fédérale de Lausanne, Lausanne, Switzerland and ⁷Department of Immunology and Infectious diseases, Harvard School of Public Health, Boston, MA, USA

*Corresponding author: C Hetz, Institute of Biomedical Sciences, Faculty of Medicine, University of Chile, Independencia 1027, FXB Rm. 205, 651 Huntington Avenue, PO BOX 70086, Santiago, Chile. Tel: + 56 2 29786506; Fax: + 56 2 29786871; E-mail: chetz@hsph.harvard.edu or chetz@med.uchile.cl

Keywords: Autophagy; adeno-associated vector (AAV); microtubule-associated protein 1 light chain 3 (LC3); nervous system; autophagy flux

Abbreviations: AAV, adeno-associated vectors; LC3, microtubule-associated protein 1 light chain 3; LC3-II, microtubule-associated protein 1 light chain 3 lipidated form; DRP, DNase-resistant particles; ICV, intracerebroventricular; ChAT, choline acetyltransferase; EBSS, Earle's balanced salt solution; IP, intraperitoneal; SCI, spinal cord injury; DRG, dorsal root ganglia; CNS, central nervous system; BBB, blood-brain barrier; DMSO, dimethyl sulfoxide; PBS, phosphate-buffered saline solution; DAPI, 4',6-diamidino-2-phenylindole

Received 19.2.13; revised 28.8.13; accepted 29.8.13; Edited by A Verkhratsky

De novo expression of connexin hemichannels in denervated fast skeletal muscles leads to atrophy

Luis A. Cea^{a,b,1}, Bruno A. Cisterna^a, Carlos Puebla^{a,b}, Marina Frank^c, Xavier F. Figueroa^a, Christopher Cardozo^{d,e}, Klaus Willecke^c, Ramón Latorre^{b,1}, and Juan C. Sáez^{a,b,1}

^aDepartamento de Fisiología, Pontificia Universidad Católica de Chile, Santiago 6513677, Chile; ^bCentro Interdisciplinario de Neurociencias de Valparaíso, Universidad de Valparaíso, Valparaíso 2366103, Chile; ^cLife and Medical Sciences Institute, Division of Molecular Genetics, University of Bonn, 53115 Bonn, Germany; ^dCenter of Excellence for the Medical Consequences of Spinal Cord Injury, James J. Peters Veterans Affairs Hospitals Medical Center, Bronx, NY 10468; and ^eDepartment of Medicine, Mount Sinai School of Medicine, New York, NY 10468

Contributed by Ramón Latorre, August 14, 2013 (sent for review June 25, 2013)

Denervation of skeletal muscles induces atrophy, preceded by changes in sarcolemma permeability of causes not yet completely understood. Here, we show that denervation-induced Evans blue dye uptake in vivo of fast, but not slow, myofibers was acutely inhibited by connexin (Cx) hemichannel/pannexin1 (Panx1) channel and purinergic ionotropic P2X₇ receptor (P2X₇R) blockers. Denervated myofibers showed up-regulation of Panx1 and de novo expression of Cx39, Cx43, and Cx45 hemichannels as well as P2X₇Rs and transient receptor potential subfamily V, member 2, channels, all of which are permeable to small molecules. The sarcolemma of freshly isolated WT myofibers from denervated muscles also showed high hemichannel-mediated permeability that was slightly reduced by blockade of Panx1 channels or the lack of Panx1 expression, but was completely inhibited by Cx hemichannel or P2X₇R blockers, as well as by degradation of extracellular ATP. However, inhibition of transient receptor potential subfamily V, member 2, channels had no significant effect on membrane permeability. Moreover, activation of the transcription factor NF- κ B and higher mRNA levels of proinflammatory cytokines (TNF- α and IL-1 β) were found in denervated WT but not Cx43/Cx45-deficient muscles. The atrophy observed after 7 d of denervation was drastically reduced in Cx43/Cx45-deficient but not Panx1-deficient muscles. Therefore, expression of Cx hemichannels and P2X₇R promotes a feed-forward mechanism activated by extracellular ATP, most likely released through hemichannels, that activates the inflammasome. Consequently, Cx hemichannels are potential targets for new therapeutic agents to prevent or reduce muscle atrophy induced by denervation of diverse etiologies.

connexons | membrane leakage | purinergic receptors | phosphorylated p65 | inflammation

Denervated skeletal muscles undergo a change in membrane permeability along with a progressive array of metabolic, structural, and functional changes that lead to atrophy (1). For example, at approximately 7 d after denervation, rodent skeletal muscles show a decrease in intracellular K⁺ concentration (2) and an increase in intracellular Na⁺ concentration (1) and total calcium content (1). In addition, contraction of denervated skeletal muscle depends on extracellular Ca²⁺ as early as 6 d after denervation (3). A possible explanation for this latter result is that denervation induces the expression of the cardiac Ca²⁺-permeable dihydropyridine receptor isoform (4). However, this protein is only expressed from day 25 of denervation. Therefore, the Ca²⁺-dependency of denervated muscles for a single contraction remains unexplained (4). The increase in dihydropyridine receptors Cav1.1 and ryanodine receptor complex has also been proposed to contribute to the increase in free Ca²⁺ concentration (5), but the denervation-induced reduction in membrane potential (V_m) is not sufficient to activate these channels (6). The changes in intracellular Na⁺ and K⁺ concentrations and V_m reduction of denervated myofibers might be explained by a deficiency in Na⁺/K⁺-dependent ATPase pump activity, but ouabain

still induces a ~10% reduction in V_m in denervated myofibers (7). Consequently, the transmembrane electrochemical changes induced by denervation are, at present, not fully explained. An alternative mechanism could be the de novo expression of non-selective cation channels, which, to our knowledge, has not been reported. Investigation of this possibility was the main goal of the present work.

To date, treatments with several compounds have only partially reduced the development of atrophy (8). However, substantial reduction of myofiber atrophy has been obtained upon muscle-specific inhibition of NF- κ B through expression of I κ B- α super-repressor (1) or genetic deletion of either of two muscle-specific E3 ligases, atrogin-1 or muscle ring finger-1 (MurF1) (1). However, the sequence of events that initiates muscle atrophy and the relevance of most changes induced by denervation remain uncertain.

Here, we demonstrated that denervated fast skeletal muscles express de novo the monovalent cation and Ca²⁺-permeable channels connexins (Cxs) 39, 43, and 45, purinergic ionotropic P2X₇ receptors (P2X₇Rs), as well as transient receptor potential, subfamily V, member 2 (TRPV2), channels, and show an up-regulation of pannexin1 (Panx1). The relevance of functional Cx hemichannels and P2X₇Rs in denervation-induced permeabilization of the sarcolemma was also demonstrated. In addition, denervation was found to induce an inflammatory state of myofibers associated with muscular atrophy, and both responses were greatly

Significance

In this paper two biological findings are described and explain several muscle changes induced by denervation: (i) the sarcolemma of fast myofibers are permeabilized to small molecules such as Evans blue via connexin (Cx) hemichannels and (ii) the absence of Cx43/Cx45 hemichannels greatly attenuates the inflammasome activation and muscle atrophy. The first finding explains the activation of proteolysis in denervated muscles. The second demonstrates that muscle inflammation can occur without inflammatory cell infiltration, offering an explanation how denervated muscles can alter other tissues. These findings unveil therapeutic targets to reduce atrophy in diverse clinical conditions. Because Cx hemichannels are permeable to Evans blue, the use of this dye as tracer of cell damage should be reevaluated in different systems.

Author contributions: L.A.C., X.F.F., C.C., K.W., R.L., and J.C.S. designed research; L.A.C., B.A.C., C.P., and M.F. performed research; L.A.C. and M.F. contributed new reagents/analytic tools; L.A.C., B.A.C., and C.P. analyzed data; and L.A.C., C.P., X.F.F., C.C., K.W., R.L., and J.C.S. wrote the paper.

The authors declare no conflict of interest.

¹To whom correspondence may be addressed. E-mail: lacea@uc.cl, ramon.latorre@uv.cl, or jsaez@bio.puc.cl.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1312331110/-DCSupplemental.

Chapter 1

Spike Train Statistics From Empirical Facts To Theory: The Case Of The Retina.

Bruno Cessac and Adrian Palacios

Abstract This chapter focuses on methods from statistical physics and probability theory allowing the analysis of spike trains in neural networks. Taking as an example the retina we present recent works attempting to understand how retina ganglion cells encode the information transmitted to the visual cortex via the optical nerve, by analyzing their spike train statistics. We compare the maximal entropy models used in the literature of retina spike train analysis to rigorous results establishing the exact form of spike train statistics in conductance-based Integrate-and-Fire neural networks.

This chapter is done in the spirit of the course "Neuronal dynamics", given at the Master of Computational Biology, University of Nice, aiming at showing how a specific problem in neuroscience can be addressed on theoretical grounds, and how it can be related to experimental methods and results. As a consequence, this chapter contains both recent biological results and mathematical developments.

1.1 Introduction

Given a stimulus from the external world (visual scene, sound, smell, ...) biological sensors at the periphery of the nervous system are able to transduce the physical manifestations of this stimulus (light emission, air pressure variations, chemical concentrations) into sequences of action potentials (spike trains), which propagate through the nervous system. Then, the brain is able to *analyze* those spike trains and infer crucial information on the nature of the stimulus. Critical - yet unsolved - questions in neuroscience are How is the physical signal encoded by the nervous system ? How does the brain analyze the spike trains ? What are the underlying computational *coding* principles ? At the current stage of scientific knowledge, answering those questions is still a challenge for biology and computational neuroscience.

Among sensory systems the retina provides functionality such as detection of movement, orientation, temporal and spatial prediction, response to flash omissions and contrast, that were up to recently viewed as the exclusive duty of higher brain centers [22]. The retina is an accessible part of the brain [13] and a prominent system to study the neurobiology and the underlying computational capacity of the neural coding. As a matter of fact, there is currently a wide research activity in understanding how the retina encodes visual information. However, basic questions are still open, such as: Are the ganglion cells (which send spikes from the eyes to the brain via the optical nerve), independent signal-encoders or are neural correlations important for coding a visual scene, and how to interpret them ?

B. Cessac

INRIA NeuroMathComp, 2004 Route des Lucioles, 06902 Sophia-Antipolis, France. e-mail: bruno.cessac@inria.fr

A. G. Palacios

CINV-Centro Interdisciplinario de Neurociencia de Valparaiso, Universidad de Valparaiso, Harrington 287, Valparaiso 2360102, Chile. e-mail: adrian.palacios@uv.cl

A BK (Slo1) channel journey from molecule to physiology

Gustavo F Contreras^{1,2}, Karen Castillo¹, Nicolás Enrique⁴, Willy Carrasquel-Ursulaez^{1,2}, Juan Pablo Castillo^{1,3}, Verónica Milesi⁴, Alan Neely¹, Osvaldo Alvarez³, Gonzalo Ferreira⁵, Carlos Gonzalez^{1,*}, and Ramon Latorre^{1,*}

¹Centro Interdisciplinario de Neurociencia de Valparaíso; Facultad de Ciencias; Universidad de Valparaíso; Valparaíso, Chile; ²Doctorado en Ciencias mención Neurociencia; Universidad de Valparaíso; Valparaíso, Chile; ³Facultad de Ciencias; Universidad de Chile; Santiago, Chile; ⁴Grupo de Investigación en Fisiología Vascular (GINFIV); Universidad Nacional de la Plata; La Plata, Argentina; ⁵Laboratorio de Canales Iónicos; Departamento de Biofísica; Facultad de Medicina; Universidad de la República; Montevideo, Uruguay

Keywords: BK channels, Slo1, auxiliary subunits, voltage sensor, intracellular Ca^{2+} , smooth muscle, diseases

Abbreviations: BK, big conductance voltage and Ca^{2+} -dependent potassium channel; Charybdotoxin, ChTx; Iberotoxin, IbTx; regulator of the conductance of K^+ channels, RCK; voltage sensing domain, VSD; leucine-rich repeat proteins, LRR; nitric oxide, NO; cyclic guanosin mono-phosphate, cGMP

Calcium and voltage-activated potassium (BK) channels are key actors in cell physiology, both in neuronal and non-neuronal cells and tissues. Through negative feedback between intracellular Ca^{2+} and membrane voltage, BK channels provide a damping mechanism for excitatory signals. Molecular modulation of these channels by alternative splicing, auxiliary subunits and post-translational modifications showed that these channels are subjected to many mechanisms that add diversity to the BK channel α subunit gene. This complexity of interactions modulates BK channel gating, modifying the energetic barrier of voltage sensor domain activation and channel opening. Regions for voltage as well as Ca^{2+} sensitivity have been identified, and the crystal structure generated by the 2 RCK domains contained in the C-terminal of the channel has been described. The linkage of these channels to many intracellular metabolites and pathways, as well as their modulation by extracellular natural agents, has been found to be relevant in many physiological processes. This review includes the hallmarks of BK channel biophysics and its physiological impact on specific cells and tissues, highlighting its relationship with auxiliary subunit expression.

Introduction

BK channels are members of a family of Ca^{2+} and voltage-dependent potassium channels. They are constituted by a tetramer of α subunits that form the conducting pore and are encoded by the *slo1* gene. In several tissues, BK channels have been observed to be modulated by auxiliary subunits, which confer important physiological performance to the channels. BK channels are ubiquitously expressed in cell membranes of mammalian tissues, where they couple signals that result from differences in membrane

voltage and intracellular Ca^{2+} concentration, which are both key actors in the physiology of nervous and non-nervous cells. In this review we provide an overview of BK channel function and its relationship with structural cues, the voltage sensor domain and gating properties of the channels, as well as its crosstalk with its auxiliary subunits.

A Short Story about How BK Channels Were Identified

The first evidence of a K^+ permeability induced by increases in intracellular calcium concentration was obtained in red blood cells.¹ Later, a calcium-dependent K^+ current was reported from experiments where Ca^{2+} was injected in motoneurons, resulting in both an increase in membrane conductance and a decrease in cellular excitability.² Moreover, the removal of external Ca^{2+} was found to decrease a voltage-dependent K^+ current in mollusk neurons.^{3,4} A few years later, these currents were defined as being carried by a calcium-dependent potassium current,⁵ after which their critical role in neuronal firing properties and hyperpolarization was soon acknowledged.⁶

The year 1981 was the *annus mirabilis* of BK channel research, since abundant expression of these channels was found in skeletal muscles and chromaffin cells. Single channel recordings from skeletal muscle and chromaffin cells^{7,8} as well as the reconstitution of a calcium-dependent K^+ channel in bilayers⁹ revealed the large conductance of BK channels, which ranges within 200 pS. This magnitude gave rise to the names MaxiK or BK, thus representing the large conductance potassium channel.¹⁰ With the advent of the giga-seal patch clamp technique¹¹ and the possibility of isolating and patching small cells, BK channels were soon found and described in liver, lymphocyte, epithelium, exocrine, and endocrine glands as being linked to excitation-secretion coupling.^{8,12–14} Marty et al. discovered that channels of different conductance give rise to Ca^{2+} -dependent potassium currents in rat lacrimal glands and named the largest conductance type channel *BK*.¹⁵ By that time, another important finding regarding this channel's localization was made, namely that it is abundantly

*Correspondence to: Carlos Gonzalez; Email: carlos.gonzalez@uv.cl; Ramon Latorre; Email: ramon.latorre@uv.cl
Submitted: 07/30/2013; Revised: 08/20/2013; Accepted: 08/22/2013
<http://dx.doi.org/10.4161/chan.26242>

E. Couve^{1,3*}, R. Osorio^{1,3},
and O. Schmachtenberg^{2,3}

¹Departamento de Biología, Laboratorio de Microscopía Electrónica; ²Centro Interdisciplinario de Neurociencia de Valparaíso (CINV); and ³Facultad de Ciencias, Universidad de Valparaíso, Avda. Gran Bretaña 1111, Playa Ancha, Valparaíso, Chile; *corresponding author; eduardo.couve@uv.cl

J Dent Res 92(9):765-772, 2013

ABSTRACT

Odontoblasts are dentin-secreting cells that survive for the whole life of a healthy tooth. Once teeth are completely erupted, odontoblasts transform into a mature stage that allows for their functional conservation for decades, while maintaining the capacity for secondary and reactionary dentin secretion. Odontoblasts are also critically involved in the transmission of sensory stimuli from the dentin-pulp complex and in the cellular defense against pathogens. Their longevity is sustained by an elaborate autophagic-lysosomal system that ensures organelle and protein renewal. However, progressive dysfunction of this system, in part caused by lipofuscin accumulation, reduces the fitness of odontoblasts and eventually impairs their dentin maintenance capacity. Here we review the functional activities assumed by mature odontoblasts throughout life. Understanding the biological basis of age-related changes in human odontoblasts is crucial to improving tooth preservation in the elderly.

KEY WORDS: dentin, human dental pulp, sensory system, lipofuscin.

The Amazing Odontoblast: Activity, Autophagy, and Aging

INTRODUCTION

Odontoblasts derive from cranial neural crest cells that emerge early during vertebrate evolution (Chai *et al.*, 2000; Hall and Gillis, 2013). Their secretory product, dentin, is one of the first manifestations of extracellular matrix based on biomineralized collagen. This hard compound appeared in the Ordovician period in jawless fish with a dermal skeleton, whose tooth-like denticles contained dentinous tissues similar to the orthodentin that forms the bulk of contemporary vertebrate teeth (Smith and Sansom, 2000; Kawasaki and Weiss, 2008). In mammals, odontoblasts are long-lived post-mitotic cells organized in a continuous cellular palisade at the dentin-pulp interface, where they maintain pre-dentin and dentin apposition for the whole life of a tooth. Together with an impressive network of trigeminal nerve fibers, they form a complex sensory organ, detecting and transmitting changes in temperature, mechanical stimuli, and pain (Byers *et al.*, 2003; Magloire *et al.*, 2010; Farahani *et al.*, 2011).

Increasing life expectancy in humans, exceeding 80 years in several developed countries, means that ever more individuals are reaching old age. From an oral health perspective, the increase in human longevity implies the necessity for prolonged tooth preservation (Tsakos, 2011). Understanding the mechanisms and biological foundation of age-related changes affecting odontoblasts and dentin apposition along life may create novel foundations for dental therapy and the application of adequate protective measures, depending on the age of the tooth.

This article reviews the different activities of odontoblasts during their life cycle, with a focus on their secretory, sensory, and defensive functions. In addition, it aims to define the homeostatic mechanisms that maintain mature odontoblasts for several decades, describing their autophagic-lysosomal system as one of the main pathways for organelle turnover in this cell (Couve, 1986; Couve and Schmachtenberg, 2011; Couve *et al.*, 2012). Finally, the consequences of odontoblast aging for tooth preservation are discussed.

DENTIN-SECRETORY ACTIVITY OF ODONTOBLASTS

Odontoblasts differentiate from embryonic ectomesenchymal cells derived from the cranial neural crest during the initial events of tooth development (Koussoulakou *et al.*, 2009). The development of pre-odontoblasts into terminally differentiated odontoblasts results in highly polarized cells with a columnar shape, about 50 μm in height (Couve, 1986). They are connected by junctional complexes forming a densely packed palisade at the dentin-pulp interface, from where each cell projects an odontoblastic process into the pre-dentin/dentin matrix. In erupted permanent teeth with completely formed crowns, coronal odontoblasts are organized in a pseudostratified palisade,

DOI: 10.1177/0022034513495874

Received April 10, 2013; Last revision May 20, 2013;
Accepted June 6, 2013

© International & American Associations for Dental Research



Editorial

Beyond the retina neural coding: On models and neural rehabilitation

In the periphery, known sensory systems are at the boundary between physics and biology. The molecular and cellular designs of such systems allow for the generation and propagation of action potentials (spikes) through an intricate neural-coding network to the brain. Among these sensory systems, the retina stands out for its functionalities, such as motion detection, orientation, anticipatory temporal prediction as well as, response to flash omissions and contrast, which were all viewed in the past as being exclusively processed by higher brain areas (Golisch and Meister, 2008). The retina is an accessible part of the brain (Dowling, 1987) and, thus, has become a unique model system to study phototransduction, shedding light on the first steps into neural coding and its underlying computational capacity. The retina is a tightly packed neural tissue comprised on a rich diversity of neurons and structured into three cellular and two synaptic layers (Masland, 2001a). The anatomical and physiological segregation of different aspects of visual scenes in separate spatial, temporal and chromatic channels start at the retina and relies on local neural networks (Atick, 1992). However, how the precise articulation of this neural network contributes to local solutions and global perception is still largely a mystery. The recent description of a diversity of ganglion cell (GC) neurons (the output of the retina that generate spikes) conveys an important question about the actual early visual capacity of the retina. By using high-density multi-electrode arrays (MEAs) over a small piece of retina, a complete population of GC has been characterized in salamanders (Segev et al., 2004) and monkeys Pillow et al. (2008). Nevertheless, only a reduce fraction (5/20) of the existing GC types, including a diversity of non-standard behaviors (Masland, 2001a; Masland, 2001b; Masland and Martin, 2007; Schwartz et al., 2007) has been studied in detail [e.g. Golisch and Meister, 2008].

How does the brain make sense of the physical world? How is a physical signal encoded by the nervous system? To what extent does our natural environment shape our sensory systems? What are the underlying computational coding principles? Is it possible to characterize neural response statistics under a common framework? What are the underlying mechanisms for color vision? How is sensory information integrated to perform complex computations? Those are some of the question that were reviewed during the II LACONEU summer school (Valparaíso, Chile). Even though the main topic of this summer school was retina oriented, we have included works on neural coding in higher order areas and in visual rehabilitation within this Special Issue of Journal of Physiology – Paris: Neural Coding-Chile 2012.

Sensory systems are endowed with the neural coding capacity to transmit incoming signals from the environment through limited capacity channels all the way to the brain (Attneave, 1954; Barlow, 1961; Shannon, 1948). From the standpoint of biological

adaptation, the adequacy of sensory design (Barlow, 1961) can match an optimal solution, where the biological mechanisms are embedded in an intricate chemical and electrical neural network. Are redundancy and noise reduced prior to signal transmission to visual cortex? Or are they both necessary for sensory encoding? In an influential article, Nirenberg et al. (2001) suggest that rat GCs act as independent encoders. However recently the relevance of correlated (or synchrony) responses, and therefore population code, has been fully reconsidered. Population code refers to the computational capacity of a neural assembly in terms of spike probability distribution, considering, for example, noise removal, relevance for critical behavior and predictability (Averbeck et al., 2006; Pouget et al., 2000) to code for visual signals. Evidence supporting an orchestrated (i.e., correlated) GC spiking response comes from pioneer studies by Rodieck (1967) and Mastronarde (1983a); Mastronarde (1983b); Mastronarde (1983c).

One of the first stages of any neural coding analysis is the identification and classification of retinal GCs present in the studied retina. The receptive fields of retinal GCs integrate information from a certain region of the visual scene. This, has been classically treated as a linear mechanism computing a known feature followed by a nonlinear computation normally associated to spike generation. Golisch (2013), in this issue, reviews the advances in understanding the nonlinear spatial integration performed by some retinal GCs inside their receptive fields. The nonlinear mechanisms here reviewed are not only associated to the spike generation of retinal GCs, but also to the nonlinear mechanism existing between bipolar and GCs. Moreover, these nonlinear interactions have been associated to singular feature extraction performed by some retinal GCs, normally related to relevant natural visual stimuli. Additionally, within the framework of nonlinear computations, one of the features detected by a population of retinal GCs is motion direction. In this issue, the article presented by Escobar et al. (2013) reviews the mathematical, theoretical and biophysical models for motion computation in the retina linking the mathematical constraints needed for motion direction selectivity and the biological mechanisms that can be mapped to accomplish these constraints. This article also reviews the biophysical models existing in the literature, in the effort to explain the underlying mechanisms related to motion direction, as well as to identify the critical actors in this computation.

A critical aspect for neural coding in a population of spiking neurons is to infer the statistical distribution of their response. In fact, the probability distribution associated to sensory spiking neurons has been normally estimated by using the questionable assumption that there is conditional independence between neurons. The article of Cofre and Cessac (2013) reviews the Gibbs distribution, originally used in statistical physics, proposing it as



Review Paper

Mathematical analysis and modeling of motion direction selectivity in the retina



María-José Escobar^a, Danilo Pezo^b, Patricio Orio^{b,c,*}

^a Universidad Técnica Federico Santa María, Department of Electronics Engineering, Avda España 1680, Valparaíso, Chile

^b Centro Interdisciplinario de Neurociencia de Valparaíso, Universidad de Valparaíso, Pje Harrington 287, Valparaíso, Chile

^c Facultad de Ciencias, Universidad de Valparaíso, Avda Gran Bretaña 1111, Valparaíso, Chile

ARTICLE INFO

Article history:

Available online 2 September 2013

Keywords:

Motion direction selectivity
Retina
Direction-selective retinal ganglion cell
Starburst amacrine cell
Computational model
Compartmental model

ABSTRACT

Motion detection is one of the most important and primitive computations performed by our visual system. Specifically in the retina, ganglion cells producing motion direction-selective responses have been addressed by different disciplines, such as mathematics, neurophysiology and computational modeling, since the beginnings of vision science. Although a number of studies have analyzed theoretical and mathematical considerations for such responses, a clear picture of the underlying cellular mechanisms is only recently emerging. In general, motion direction selectivity is based on a non-linear asymmetric computation inside a receptive field differentiating cell responses between *preferred* and *null* direction stimuli. To what extent can biological findings match these considerations? In this review, we outline theoretical and mathematical studies of motion direction selectivity, aiming to map the properties of the models onto the neural circuitry and synaptic connectivity found in the retina. Additionally, we review several compartmental models that have tried to fill this gap. Finally, we discuss the remaining challenges that computational models will have to tackle in order to fully understand the retinal motion direction-selective circuitry.

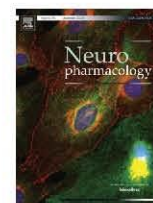
© 2013 Elsevier Ltd. All rights reserved.

Contents

1. Introduction	350
2. The mathematical basis of motion detection and direction selectivity	350
2.1. Theoretical models	350
2.2. Theoretical synaptic models	351
3. Motion mechanisms in the mammalian retina	352
3.1. Neurons computing motion direction in the retina	352
3.2. Mapping theoretical considerations to biological structure	353
3.2.1. Spatial asymmetry	353
3.2.2. Time delay between excitatory and inhibitory inputs	353
3.2.3. Non-linear interactions	353
4. Biophysical models of motion direction selectivity in the retina	354
4.1. Modeling starburst amacrine cells	354
4.2. Models of the SAC network	355
4.3. Modeling direction-selective retinal ganglion cell	355
5. Future challenges	356
Acknowledgements	357
Appendix A. Mathematical analysis of motion detectors	357
A.1. The Reichardt detector	357
A.2. The Gradient detector	358
References	358

* Corresponding author at: Centro Interdisciplinario de Neurociencia de Valparaíso, Universidad de Valparaíso, Pje Harrington 287, Valparaíso, Chile. Tel.: +56 322995538.

E-mail address: patricio.orio@uv.cl (P. Orio).



Diffusion of nitric oxide across cell membranes of the vascular wall requires specific connexin-based channels

Xavier F. Figueroa^{a,*}, Mauricio A. Lillo^a, Pablo S. Gaete^a, Manuel A. Riquelme^a,
Juan C. Sáez^{a,b}

^aDepartamento de Fisiología, Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Alameda 340, Casilla 114-D, Santiago, Chile

^bInstituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Valparaíso, Chile

ARTICLE INFO

Article history:

Received 6 September 2012

Received in revised form

25 February 2013

Accepted 27 February 2013

Keywords:

Vasodilation

Endothelial cells

Myoendothelial gap junctions

Hemichannels

Resistance arteries

ABSTRACT

NO is generated within cells and frequently must be transferred to responsive neighboring cells, as occurs in the endothelium-dependent relaxation of smooth muscle cells observed in blood vessels. It is thought that NO diffuses freely across cell membranes, but it may also permeate through low resistant membrane pathways. Here, we describe the participation of connexin (Cx)-formed channels in the NO transport across cell membranes and between endothelial and smooth muscle cells. We used a water-soluble NO donor of high molecular weight (S-nitrosylated albumin, BSA-NO) that does not permeate through cell membranes or Cx-based channels and the NO-sensitive dye 4,5-diaminofluorescein diacetate to detect changes of intracellular NO concentration. We found that NO generated in the extracellular space was not detected intracellularly in Cx-deficient HeLa cells, suggesting that cell membrane represents a significant diffusion barrier for NO transfer. However, Cx-based channels provide efficient pathways for NO signaling because NO opened and permeated hemichannels expressed in HeLa cells transfected with Cx43, Cx40, or Cx37. In contrast, NO closed hemichannels of HeLa-Cx32 cells, which otherwise are permeable to NO if are opened by a divalent cation-free extracellular solution. Consistent with this, blockade of Cx-based channels abolished the myoendothelial NO transfer and associated NO-dependent vasodilation induced by acetylcholine. These results indicate that Cx-based channels play a key role in the NO-dependent tonic control of vascular function and may direct the NO signal to specific targets, which provides a novel mechanistic basis for the critical role of Cxs in cell–cell communication in the vessel wall.

This article is part of the Special Issue Section entitled ‘Current Pharmacology of Gap Junction Channels and Hemichannels’.

© 2013 Elsevier Ltd. All rights reserved.

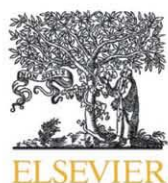
1. Introduction

Nitric oxide (NO) is a critical signaling molecule that plays a central role in the regulation of diverse systems, including immune, nervous and cardiovascular systems (Moncada et al., 1991). In blood vessels, NO is a potent vasodilator involved in the minute-to-minute and long-term control of vascular function (Moncada et al., 1991). Since NO is generated in the endothelium by the endothelial NO synthase (eNOS), it must reach the underlying smooth muscle cells or the lumen of the vessel to exert some of its biological effects (Moncada et al., 1991). The small size and hydrophobicity of NO has led to think that it diffuses freely across cell membranes. However, NO has a very short half-life (~4 s)

(Moncada et al., 1991; Thomas et al., 2001), and its concentration in the cell membrane may decrease its bioavailability and efficacy. Although NO may react with other radicals, as well as transition metal centers and oxygen (O₂), in physiological conditions the lifetime of NO mainly depends on the relative abundance of O₂, which has solubility and diffusion properties similar to that of NO (Moller et al., 2005; Subczynski et al., 2009). Consequently, the chemical reaction between O₂ and NO has been estimated to be 2–2.5 times faster when both gases diffuse and concentrate in the plasma membrane compared to in the aqueous phase (Liu et al., 1998; Subczynski et al., 2009). In addition, the diffusion rate and concentration time of NO in the cell membrane are not consistent with the physiological signaling kinetics of NO. In this context, direct measurements of bradykinin-induced NO production using porphyrinic microsensors showed that the NO concentration in the endothelial cell membrane peaks after ~13 s (Malinski et al., 1993).

* Corresponding author. Tel.: +56 2 686 2356; fax: +56 2 222 5515.

E-mail address: xfigueroa@bio.puc.cl (X.F. Figueroa).



Linoleic acid induces opening of connexin26 hemichannels through a PI3K/Akt/ Ca^{2+} -dependent pathway

Vania Figueroa^{a,b}, Pablo J. Sáez^a, José D. Salas^c, Daniela Salas^a, Oscar Jara^b, Agustín D. Martínez^b, Juan C. Sáez^{a,b,*}, Mauricio A. Retamal^{c,**}

^a Departamento de Fisiología, Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Santiago, Chile

^b Instituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Universidad de Valparaíso, Valparaíso, Chile

^c Departamento de Fisiología, Facultad de Medicina, Clínica Alemana-Universidad del Desarrollo, Santiago, Chile

ARTICLE INFO

Article history:

Received 18 May 2012

Received in revised form 4 December 2012

Accepted 6 December 2012

Available online 19 December 2012

Keywords:

Polyunsaturated fatty acid

Hemichannel

Cx26

Intracellular calcium

Hearing loss

ABSTRACT

Connexin hemichannel (Cx HC) opening is involved in physiological and pathological processes, allowing the cellular release of autocrine/paracrine signaling molecules. Linoleic acid (LA) is known to modulate the functional state of connexin46 (Cx46) HCs. However, the molecular mechanism involved in this effect, or whether LA affects HCs constituted of other connexins, remains unknown. Here, we report the effects of LA on HCs in HeLa cells that express Cx26, one of the main Cxs in the cochlear sensory epithelium. Cx26 HC activity (dye uptake) was increased in a concentration-dependent manner by bath application of LA and inhibited by HC blockers. Moreover, intracellular BAPTA, a Ca^{2+} chelator, and PI3K/AKT inhibitors were found to reduce the LA-induced Cx26 HC opening, suggesting that the LA effect is mediated by an increase of free intracellular Ca^{2+} concentration and activation of the PI3K/Akt-dependent pathway. The LA-induced increase in free intracellular Ca^{2+} concentration was mainly due to Ca^{2+} influx through Cx26 HCs. In addition, the involvement of $-\text{SH}$ groups was ruled out, because dithiothreitol (DTT) did not block the LA-induced dye uptake. LA also increased the membrane current mediated by Cx26 HCs expressed in *Xenopus* oocytes and the dye uptake in HeLa cells expressing Cxs 32, 43 or 45. Since LA is an essential polyunsaturated fatty acid, its effect on HCs might be relevant to cell growth as well as to cellular functions of differentiated cells such as audition.

© 2012 Elsevier B.V. All rights reserved.

1. Introduction

A hemichannel (HC) results from the oligomerization of six protein subunits called connexins (Cxs). Serial docking of two HCs, each one located at the cell membrane of two neighboring cells, forms an intercellular gap-junction channel (GJC). This GJC connects the cytoplasm of two adjacent cells, allowing the exchange of ions and small cytoplasmic molecules, including metabolites and second messengers [1]. Until about a decade ago, HCs were thought to remain closed and open only when they are part of GJCs. However, during the past decade growing evidence has shown that HCs open under physiological as well as pathological conditions, providing a mechanism for communication between the cytoplasm and the external microenvironment

[2,3]. Through controlled release of signaling molecules, HC opening has been implicated in important physiological events such as cell cycle progression and hence in development and also in pathological processes where they promote and/or accelerate cell death [4].

Connexin26 (Cx26) is expressed in a variety of tissues, including skin and cochlear sensory epithelia, where it fulfills important functions [5,6]. The latter have been unveiled by deafness and skin diseases associated with several mutations in gene GJB2, which encodes Cx26 [7,8].

Polyunsaturated fatty acids (PUFAs), omega-6 and omega-3, and their metabolites influence physiological and pathological processes acting as pro-inflammatory or anti-inflammatory agents, respectively [9]. They are structural components of phospholipids in cell membranes, and can affect their fluidity and flexibility [10]. In addition, recent studies suggest that omega-3 PUFAs influence the expression and phosphorylation status of Cx43 [11].

The functional state of Cx26 HCs and GJCs is modulated by fatty acids. For example, oleamide derivatives of omega-9 unsaturated Oleic acid, inhibit Cx26 GJCs in mouse melanoma BL6 cells [12], while arachidonic acid (AA), an omega-6 PUFA, potentiates the cellular release of ATP promoted by a divalent cation-free solution in HeLa-Cx26 cells [13]. Recently, linoleic acid (LA), an essential unsaturated omega-6 fatty acid, was shown to induce a biphasic effect on the functional state of HCs formed by Cx46 in *Xenopus laevis* oocytes, used as an

Abbreviations: Cxs, connexins; GJCs, gap junction channels; HCs, hemichannels; LA, linoleic acid; Etd, ethidium; DTT, dithiothreitol

* Correspondence to: J.C. Sáez, Departamento de Fisiología, Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Santiago, Chile. Tel.: +56 2 6862862; fax: +56 2 2225515.

** Correspondence to: M.A. Retamal, Departamento de Fisiología, Facultad de Medicina, Clínica Alemana-Universidad del Desarrollo, Avenida Las Condes #12438, Santiago, Chile. Tel.: +56 2 3279 407; fax: +56 2 3279306.

E-mail addresses: jsaez@bio.puc.cl (J.C. Sáez), mretamal@udd.cl (M.A. Retamal).

11 Impact of Microglial Activation on Astroglial Connexin Expression and Function in Brain Inflammation

Christian Giaume, Nicolas Froger, Juan Andrés Orellana, Mauricio Retamal, and Juan Carlos Sáez

CONTENTS

11.1 Introduction	219
11.2 Connexin and Pannexin Expression in Microglia and Astrocytes	220
11.2.1 Microglia	220
11.2.2 Astrocytes	221
11.3 Activated Microglia Regulate Cx43 Channels in Astrocytes	222
11.4 Astroglial Cx43 Hemichannels Contribute to Neuronal Death	223
11.5 Alteration of Astroglial Connexin Expression in Brain Pathologies	225
11.6 Cannabinoid System as a Possible Therapeutic Strategy against Connexin Channel-Mediated Neurodegeneration	225
11.7 Conclusions	227
Acknowledgments	227
References	228

11.1 INTRODUCTION

The central nervous system (CNS) is capable of dynamic immune and inflammatory responses mediated by the activation of microglia, the brain resident macrophage population, and astrocyte reactivity. A growing body of evidence shows that the innate immune response exerts a dichotomous role in the brain. Under physiological conditions, microglia exhibit a resting phenotype that is associated with the production of anti-inflammatory and neurotrophic factors. Microglia shift to an activated phenotype in response to a wide range of insults that activate specific signaling pathways, thereby promoting an inflammatory response necessary to further engage the immune system. The sustained inflammation resulting in brain injury and pathology



Connexin and pannexin hemichannels in brain glial cells: properties, pharmacology, and roles

Christian Giaume^{1,2,3*}, Luc Leybaert⁴, Christian C. Naus⁵ and Juan C. Sáez^{6,7}

¹ Collège de France, Center for Interdisciplinary Research in Biology/Centre National de la Recherche Scientifique, Unité Mixte de Recherche 7241/Institut National de la Santé et de la Recherche Médicale U1050, Paris, France

² University Pierre et Marie Curie, Paris, France

³ MEMOLIFE Laboratory of Excellence and Paris Science Lettre Research University, Paris, France

⁴ Physiology Group, Department of Basic Medical Sciences, Faculty of Medicine and Health Sciences, Ghent University, Ghent, Belgium

⁵ Department of Cellular and Physiological Sciences, Life Sciences Institute, Faculty of Medicine, University of British Columbia, Vancouver, BC, Canada

⁶ Departamento de Fisiología, Pontificia Universidad Católica de Chile, Santiago, Chile

⁷ Instituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Valparaíso, Chile

Edited by:

Aida Salameh, Heart Centre
University of Leipzig, Germany

Reviewed by:

Gregory E. Morley, New York
University School of Medicine, USA
Roger J. Thompson, University of
Calgary, Canada
Panagiotis Bargiotas, University of
Heidelberg, Germany

*Correspondence:

Christian Giaume, Collège de France,
Center for Interdisciplinary Research
in Biology/Centre National de la
Recherche Scientifique, Unité Mixte
de Recherche 7241/Institut National
de la Santé et de la Recherche
Médicale U1050, 11 Place Marcelin
Berthelot, 75231 Paris Cedex 05,
France
e-mail: christian.giaume@college-de-
france.fr

Functional interaction between neurons and glia is an exciting field that has expanded tremendously during the past decade. Such partnership has multiple impacts on neuronal activity and survival. Indeed, numerous findings indicate that glial cells interact tightly with neurons in physiological as well as pathological situations. One typical feature of glial cells is their high expression level of gap junction protein subunits, named connexins (Cx), thus the membrane channels they form may contribute to neuroglial interaction that impacts neuronal activity and survival. While the participation of gap junction channels in neuroglial interactions has been regularly reviewed in the past, the other channel function of Cxs, i.e., hemichannels located at the cell surface, has only recently received attention. Gap junction channels provide the basis for a unique direct cell-to-cell communication, whereas Cx hemichannels allow the exchange of ions and signaling molecules between the cytoplasm and the extracellular medium, thus supporting autocrine and paracrine communication through a process referred to as “gliotransmission,” as well as uptake and release of metabolites. More recently, another family of proteins, termed pannexins (Panxs), has been identified. These proteins share similar membrane topology but no sequence homology with Cxs. They form multimeric membrane channels with pharmacology somewhat overlapping with that of Cx hemichannels. Such duality has led to several controversies in the literature concerning the identification of the molecular channel constituents (Cxs versus Panxs) in glia. In the present review, we update and discuss the knowledge of Cx hemichannels and Panx channels in glia, their properties and pharmacology, as well as the understanding of their contribution to neuroglial interactions in brain health and disease.

Keywords: astrocytes, oligodendrocytes, microglia, gap junctions, neuroglial interactions

INTRODUCTION

For a long time, it has been taken as dogma that connexin (Cx) proteins can only function as gap junction channels. Indeed, before the aggregation of Cxs at the junctional plaque and subsequent formation of gap junctions, hexameric rings of Cxs, termed connexons, were initially assumed to remain closed. An obvious reason for this occlusion was that, as gap junction channels are “poorly” selective for ions and permeable to low molecular weight molecules (<1 to 1.2 kDa), if once at the membrane connexons could open, the cell would lose its integrity or at least would have to spend substantial energy to maintain this energetically unfavorable condition. Such statements began to be challenged when evidence for “functional hemichannels,” a term proposed to substitute for plasma membrane connexons, were reported in the early 1990s. In these pioneering studies, Cx hemichannels were opened either by large depolarization in *Xenopus* oocytes (Paul et al., 1991) or by lowering the extracellular calcium ion (Ca^{2+}) concentration in horizontal cells (DeVries and Schwartz, 1992). Later on, the

occurrence of functional hemichannels (i.e., hemichannels that can be turned into the open state) composed of Cx43 was demonstrated in primary cultures of astrocytes in the absence of external Ca^{2+} (Hofer and Dermietzel, 1998). This observation was followed by the demonstration that metabolic inhibition performed in the presence of normal external Ca^{2+} concentrations (1–2 mM) induced cell permeabilization, due to Cx43 hemichannel opening, before loss of membrane integrity (Contreras et al., 2002). More recently, hemichannel opening in astrocytes was triggered either by treatment with pro-inflammatory cytokines or selective lipopolysaccharide (LPS) stimulation of microglia co-cultured with astrocytes, again in the presence of external Ca^{2+} (Retamal et al., 2007a). The opening of Cx43 hemichannels in astrocytes was also demonstrated to occur in experiments designed to decipher the mechanism of intercellular Ca^{2+} wave propagation in astrocytes to which both gap junction channels and hemichannels contribute (Scemes and Giaume, 2006; Leybaert and Sanderson, 2012). In this case, the release of ATP through Cx43 hemichannels

Molecular mechanism of voltage sensing in voltage-gated proton channels

Carlos Gonzalez,^{1,2} Santiago Rebolledo,¹ Marta E. Perez,¹ and H. Peter Larsson¹

¹Department of Physiology and Biophysics, University of Miami, Miami, FL 33136

²Centro Interdisciplinario de Neurociencia de Valparaíso, Universidad de Valparaíso, Valparaíso 2360103, Chile

Voltage-gated proton (Hv) channels play an essential role in phagocytic cells by generating a hyperpolarizing proton current that electrically compensates for the depolarizing current generated by the NADPH oxidase during the respiratory burst, thereby ensuring a sustained production of reactive oxygen species by the NADPH oxidase in phagocytes to neutralize engulfed bacteria. Despite the importance of the voltage-dependent Hv current, it is at present unclear which residues in Hv channels are responsible for the voltage activation. Here we show that individual neutralizations of three charged residues in the fourth transmembrane domain, S4, all reduce the voltage dependence of activation. In addition, we show that the middle S4 charged residue moves from a position accessible from the cytosolic solution to a position accessible from the extracellular solution, suggesting that this residue moves across most of the membrane electric field during voltage activation of Hv channels. Our results show for the first time that the charge movement of these three S4 charges accounts for almost all of the measured gating charge in Hv channels.

INTRODUCTION

Voltage-gated proton (Hv) channels are found in phagocytes, neurons, airway epithelia, muscle, and sperm (Decoursey, 2003; Okochi et al., 2009; Ramsey et al., 2009; Iovannisci et al., 2010; Lishko et al., 2010). In phagocytes, Hv channels play an essential role in bacterial clearing. Depolarization-activated H⁺ currents through Hv channels in the phagosomal membrane compensate for NADPH oxidase currents, activated during a process called respiratory burst, ensuring sustained production of reactive oxygen species in phagocytes to neutralize engulfed bacteria. The importance of Hv channels is supported by the observation that inhibiting Hv channels prevents further production of reactive oxygen species in the phagosomes. In addition, Hv1 knockout mice are unable to efficiently clear bacterial infections, further showing the essential role Hv channels play in bacterial clearing (Ramsey et al., 2009). Because the voltage dependence of Hv channels determines their physiological role, it is important to determine the molecular structure that defines the voltage sensitivity of Hv channels. At present, it is unknown which charged residues underlie the voltage dependence of Hv channels. Here, using the limiting slope method and accessibility experiments, we have identified the charged residues responsible for voltage activation of Hv channels.

Each subunit in an Hv channel has four transmembrane segments, called S1–S4 (Ramsey et al., 2006; Sasaki et al., 2006). These four transmembrane segments of the Hv channels are homologous to the first four transmembrane segments in voltage-gated potassium (Kv) channels (Ramsey et al., 2006; Sasaki et al., 2006). In Kv channels, the first four transmembrane segments form the voltage-sensing domain, whereas the fifth and the sixth transmembrane segments from all four Kv subunits together form the potassium-conducting pore domain (Jiang et al., 2003; Long et al., 2005). In contrast, Hv channels do not have a fifth or sixth transmembrane segment. In addition, unlike the tetrameric structure of Kv channels, Hv channels are composed of two subunits (Koch et al., 2008; Lee et al., 2008; Tombola et al., 2008). Deletion of the cytosolic domains removes the dimerization of the Hv subunits (Koch et al., 2008; Tombola et al., 2008). Interestingly, the monomeric Hv channel is a functional voltage-gated proton channel. Thus, each subunit comprises a voltage-gated H⁺ permeation pathway (Koch et al., 2008; Tombola et al., 2008).

In Kv channels, the fourth transmembrane segment (S4) contains many positively charged residues and has been shown to function as the main voltage sensor in Kv channels (Aggarwal and MacKinnon, 1996; Larsson et al., 1996; Mannuzzu et al., 1996; Seoh et al., 1996; Yang et al., 1996; Tombola et al., 2006). S4 in Hv channels

C. Gonzalez and S. Rebolledo contributed equally to this paper. Correspondence to H. Peter Larsson: Plarsson@med.miami.edu; or Carlos Gonzalez: carlos.gonzalez@uv.cl

Abbreviations used in this paper: FEP, free energy perturbation; MD, molecular dynamics; MTSEA, 2-aminoethyl MTS; MTSET, 2-(trimethylammonium)ethyl MTS; MTSPT, 3-(trimethylammonium)propyl MTS.

© 2013 Gonzalez et al. This article is distributed under the terms of an Attribution–Noncommercial–Share Alike–No Mirror Sites license for the first six months after the publication date (see <http://www.rupress.org/terms>). After six months it is available under a Creative Commons License (Attribution–Noncommercial–Share Alike 3.0 Unported license, as described at <http://creativecommons.org/licenses/by-nc-sa/3.0/>).

Dynamin-2 Regulates Fusion Pore Expansion and Quantal Release through a Mechanism that Involves Actin Dynamics in Neuroendocrine Chromaffin Cells

Arlek M. González-Jamett¹, Fanny Momboisse^{1,2}, María José Guerra¹, Stéphane Ory², Ximena Báez-Matus¹, Natalia Barraza¹, Valerie Calco², Sébastien Houy², Eduardo Couve³, Alan Neely¹, Agustín D. Martínez¹, Stéphane Gasman^{2*}, Ana M. Cárdenas^{1*}

1 Centro Interdisciplinario de Neurociencia de Valparaíso, Facultad de Ciencias, Universidad de Valparaíso, Gran Bretaña, Playa Ancha, Valparaíso, Chile, **2** Institut des Neurosciences Cellulaires et Intégratives, Centre National de la Recherche Scientifique (CNRS UPR 3212), and Université de Strasbourg, Strasbourg, France, **3** Departamento de Biología, Facultad de Ciencias, Universidad de Valparaíso, Gran Bretaña, Playa Ancha, Valparaíso, Chile

Abstract

Over the past years, dynamin has been implicated in tuning the amount and nature of transmitter released during exocytosis. However, the mechanism involved remains poorly understood. Here, using bovine adrenal chromaffin cells, we investigated whether this mechanism rely on dynamin's ability to remodel actin cytoskeleton. According to this idea, inhibition of dynamin GTPase activity suppressed the calcium-dependent *de novo* cortical actin and altered the cortical actin network. Similarly, expression of a small interfering RNA directed against dynamin-2, an isoform highly expressed in chromaffin cells, changed the cortical actin network pattern. Disruption of dynamin-2 function, as well as the pharmacological inhibition of actin polymerization with cytochalasin-D, slowed down fusion pore expansion and increased the quantal size of individual exocytotic events. The effects of cytochalasin-D and dynamin-2 disruption were not additive indicating that dynamin-2 and F-actin regulate the late steps of exocytosis by a common mechanism. Together our data support a model in which dynamin-2 directs actin polymerization at the exocytosis site where both, in concert, adjust the hormone quantal release to efficiently respond to physiological demands.

Citation: González-Jamett AM, Momboisse F, Guerra MJ, Ory S, Báez-Matus X, et al. (2013) Dynamin-2 Regulates Fusion Pore Expansion and Quantal Release through a Mechanism that Involves Actin Dynamics in Neuroendocrine Chromaffin Cells. PLoS ONE 8(8): e70638. doi:10.1371/journal.pone.0070638

Editor: Frederic A. Meunier, The University of Queensland, Australia

Received: April 26, 2013; **Accepted:** June 25, 2013; **Published:** August 5, 2013

Copyright: © 2013 González-Jamett et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work has been supported by the cooperation program between France and Chile ECOS-SUD/Conicyt (#C08B01) to SG and AMC, and by grants FONDECYT 1110552 (to AMC), ANR-07-JCJC-088-01 (to SG), CONICYT AT24100005 and 21080096 (to A G-J), and P09-022-F from ICM-ECONOMIA, Chile. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: Stéphane Gasman, is a PLOS ONE Editorial Board member. This does not alter the authors' adherence to all the PLOS ONE policies on sharing data and materials.

* E-mail: ana.cardenas@uv.cl (AMC); gasman@unistra.fr (SG)

Introduction

Dynamin is a mechano-GTPase encoded by three distinct genes (DNM1, DNM2 and DNM3) that generates membrane deformation and triggers membrane fission [1]. Its best characterized function is the scission of nascent vesicles from the plasma membrane during endocytosis. Of the three dynamin isoforms only dynamin-2 is ubiquitously expressed, while dynamin-1 is exclusively expressed in neuronal tissue, and dynamin-3 is only present in brain, testis, heart and lungs [2], [3]. Studies in knock-out animals show that only dynamin-2 is critical during early embryonic development [4] and that the absence of dynamin-1 or -3 can be compensated by the other isoforms [5]. From these findings arises the idea that the different dynamin isoforms have overlapping roles and their relative contribution to endocytosis in a given tissue is mostly determined by their abundance rather than on structural specialization [5].

Dynamin participates in several cellular processes that are dependent on the actin cytoskeleton dynamics, some of them are actin comet [6], [7] lamellipodia formation [8], T cell activation [9], phagocytosis [10] and different types of endocytosis [11–14].

Furthermore, a functional link between dynamin and actin has been observed during endocytosis, where one regulates the recruitment of the other [14] and both work synergistically to efficiently catalyze membrane scission [12]. The exact mechanism of the crosstalk between dynamin and actin is not completely clear, but some evidences suggest that dynamin binds directly to actin filaments to promote its polymerization by displacing the actin capping protein gelsolin [15]. Additionally, dynamin can control the stability of actin filaments in association with the actin-binding protein cortactin, in a GTP hydrolysis-dependent way [16], [17].

In neuroendocrine cells, both actin and dynamin have been involved in the regulation of the exocytotic process. On one hand, cortical actin network is reorganized during exocytosis [18], wherein it regulates the expansion of the fusion pore [19], an intermediate structure formed during the fusion of the secretory vesicle with the plasma membrane [20]. On the other hand, dynamin appears to regulate both fusion pore expansion [21] and closure [22], [23], and to control the quantal size of release events [24–27]. These actions have been attributed to the neuronal isoform dynamin-1 [21], [23], [25], while dynamin-2 has been proposed to specialize in the regulation of compensatory



Dynamin-2 function and dysfunction along the secretory pathway

Arlek M. González-Jamett¹, Fanny Momboisse¹, Valentina Haro-Acuña¹, Jorge A. Bevilacqua², Pablo Caviedes³ and Ana María Cárdenas^{1*}

¹ Facultad de Ciencias, Centro Interdisciplinario de Neurociencia de Valparaíso, Universidad de Valparaíso, Valparaíso, Chile

² Programa de Anatomía y Biología del Desarrollo, ICBM, Facultad de Medicina, Departamento de Neurología y Neurocirugía, Hospital Clínico Universidad de Chile, Universidad de Chile, Santiago, Chile

³ Programa de Farmacología Molecular y Clínica, ICBM, Facultad de Medicina, Universidad de Chile, Santiago, Chile

Edited by:

Stephane Gasman, Centre national de la Recherche Scientifique, France

Reviewed by:

Jocelyn Laporte, Institut National de la Santé et de la Recherche Médicale, France

Fernando Diego Marengo, Universidad de Buenos Aires, Argentina

*Correspondence:

Ana María Cárdenas, Facultad de Ciencias, Centro Interdisciplinario de Neurociencia de Valparaíso, Universidad de Valparaíso, Gran Bretaña 1111, Playa Ancha 2360102, Valparaíso, Chile
e-mail: ana.cardenas@uv.cl

Dynamin-2 is a ubiquitously expressed mechano-GTPase involved in different stages of the secretory pathway. Its most well-known function relates to the scission of nascent vesicles from the plasma membrane during endocytosis; however, it also participates in the formation of new vesicles from the Golgi network, vesicle trafficking, fusion processes and in the regulation of microtubule, and actin cytoskeleton dynamics. Over the last 8 years, more than 20 mutations in the dynamin-2 gene have been associated to two hereditary neuromuscular disorders: Charcot-Marie-Tooth neuropathy and centronuclear myopathy. Most of these mutations are grouped in the pleckstrin homology domain; however, there are no common mutations associated with both disorders, suggesting that they differently impact on dynamin-2 function in diverse tissues. In this review, we discuss the impact of these disease-related mutations on dynamin-2 function during vesicle trafficking and endocytotic processes.

Keywords: dynamin-2, endocytosis, exocytosis, actin, microtubules, mutations, Charcot-Marie-Tooth neuropathy, centronuclear myopathy

INTRODUCTION

Dynamin was identified for the first time almost 25 years ago as a 100kDa microtubule-associated protein that induced microtubule bundles and promoted microtubule sliding *in vivo* (1). As described by the same authors, the motor activity of dynamin required ATP and other co-purified polypeptides (1). A year later, the same group cloned and sequenced dynamin, and found that it contained a consensus GTP-binding site (2), and subsequently characterized its GTPase activity (3). At present, three dynamin isoforms encoded by three distinct genes (*DNM1*, *DNM2*, and *DNM3*) have been described in mammals (4). These exhibit approximately 80% homology in their sequences, yet they differ in their tissue expression pattern; dynamin-1 is mainly expressed in neuronal tissue, dynamin-2 is ubiquitously expressed, and dynamin-3 is expressed in brain, testis, and lungs (5). Of these three dynamin isoforms, only dynamin-2 appears to play a pleiotropic role during embryonic development (6). In fact, studies in knock-out animals show that deletion of dynamin-1 or -3 can be compensated by the other dynamin isoforms (7), while the deletion of dynamin-2 causes early embryonic lethality (8). Moreover, as discussed below, mutations in *DNM2* result in severe hereditary neuropathies and myopathies in humans, strongly suggesting that dynamin-2 has more susceptible functions in the nervous and skeletal muscle tissues.

All dynamin isoforms exhibit at least four alternatively spliced variants, resulting in different dynamin proteins (5) that share a primary structure comprising: a large amino-terminal GTPase domain (G-domain) that binds and hydrolyzes GTP; a middle and a GTPase effector domains (GED) that form a “stalk”

structurally essential region; a pleckstrin homology domain (PH) that binds inositol phospholipids and a carboxy-terminal proline and arginine rich-domain (PRD) that allows interaction with SH3-domain-containing-proteins (5) (Figure 1).

Dynamin function relies on its ability to form high order oligomers, and its self-assembly is necessary to promote its catalytic activity. Purified dynamin has been shown to spontaneously polymerize in the presence of negatively charged tubular templates such as lipid membranes (11), microtubules (3, 12), or actin bundles (13, 14) as well as after incubation in low ionic strength solutions (15). Over the last years several cryo-electron microscopy (16–18) and X-ray crystallographic studies (19, 20) of dynamin and its domains (21–24) have allowed a better understanding of the mechanisms mediating dynamin oligomerization. It appears that the stable dimers formed by the crossed interaction between the “stalk” regions of monomeric dynamins (20, 25) are the basic unit that allows dynamin polymerization (18), thus promoting the GTPase activation required for membrane remodeling and scission in different cellular processes.

In the present review, we discuss the different roles of dynamin during endocytosis, vesicle trafficking, and exocytosis, specially focusing in dynamin-2, and how disease-linked mutations in dynamin-2 gene alter such cellular processes.

DYNAMIN AS A KEY COMPONENT OF ENDOCYTOSIS AND VESICLE RECYCLING

Dynamin is a GTPase that plays a crucial role in the recycling of secretory vesicle in neuroendocrine cells (26).

Research Article

Boldine Prevents Renal Alterations in Diabetic Rats

Romina Hernández-Salinas,¹ Alejandra Z. Vielma,¹ Marlene N. Arismendi,¹
Mauricio P. Boric,¹ Juan C. Sáez,^{1,2} and Victoria Velarde¹

¹ Departamento de Fisiología, Pontificia Universidad Católica de Chile, Alameda Bernardo O'Higgins, No. 340, 6513677 Santiago, Chile

² Instituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Valparaíso, Chile

Correspondence should be addressed to Juan C. Sáez; jsaez@bio.puc.cl and Victoria Velarde; vvelarde@bio.puc.cl

Received 22 September 2013; Accepted 6 November 2013

Academic Editor: Raffaele Marfella

Copyright © 2013 Romina Hernández-Salinas et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Diabetic nephropathy alters both structure and function of the kidney. These alterations are associated with increased levels of reactive oxygen species, matrix proteins, and proinflammatory molecules. Inflammation decreases gap junctional communication and increases hemichannel activity leading to increased membrane permeability and altering tissue homeostasis. Since current treatments for diabetic nephropathy do not prevent renal damage, we postulated an alternative treatment with boldine, an alkaloid obtained from boldo with antioxidant, anti-inflammatory, and hypoglycemic effects. Streptozotocin-induced diabetic and control rats were treated or not treated with boldine (50 mg/Kg/day) for ten weeks. In addition, mesangial cells were cultured under control conditions or in high glucose concentration plus proinflammatory cytokines, with or without boldine (100 μ mol/L). Boldine treatment in diabetic animals prevented the increase in glycemia, blood pressure, renal thiobarbituric acid reactive substances and the urinary protein/creatinine ratio. Boldine also reduced alterations in matrix proteins and markers of renal damage. In mesangial cells, boldine prevented the increase in oxidative stress, the decrease in gap junctional communication, and the increase in cell permeability due to connexin hemichannel activity induced by high glucose and proinflammatory cytokines but did not block gap junction channels. Thus boldine prevented both renal and cellular alterations and could be useful for preventing tissue damage in diabetic subjects.

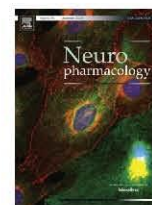
1. Introduction

Diabetic nephropathy (DN) is considered a microvascular complication of diabetes. It is characterized by a chronic injury to renal tissue, mainly glomerular structures. DN is the major complication associated with type I diabetes mellitus, affecting approximately 30–40% of diabetic patients, and is the leading cause of end-stage renal disease [1].

There are several factors involved in the development of diabetic glomerulopathy but the main initiator of this disease is chronic hyperglycemia, which triggers nonenzymatic protein glycosylation and increased production of reactive oxygen species, increasing oxidative stress and favoring coagulation and fibrotic events [2]. Hyperglycemia also sensitizes the vessel wall of efferent arterioles to the action of vasoconstrictors, which, together with an increase in vascular

permeability, lead to hyperfiltration. Also, in the progression of DN persistent proteinuria, glomerular hypertrophy, mesangial expansion, and tubulointerstitial fibrosis occur, which lead to partial or total loss of renal function [3, 4].

In diabetes and in different cell types cultured in high glucose concentration (from now and on named as high glucose), an increase in the inflammatory response is observed. This response is usually initiated by cytokine release and subsequent attraction of macrophages, important actors in the progression of DN [5, 6]. In this and other diseases where inflammation is present, the expression of connexins (Cxs), including Cx43, is altered and increased activity of Cx formed hemichannels is observed. The latter alterations have been linked to a reduction in gap junction activity (essential for intercellular communication) and have been considered a risk factor for certain diseases [7, 8].



Editorial

We've had important advances in the connexin/pannexin field, yet there is still much to do

In recent years, we have seen major advances in the field of connexin (Cx) and pannexin (Panx) proteins. New methods have been developed or adapted to work in this area of research and a number of Cx mutations linked to human disease have been found. Moreover, work on Cx hemichannels (Cx HCs, prior to docking and the formation of gap junction (GJ) channels) has exploded, having implications for a variety of biological processes. Studies of Panx channels have also captivated the interest of investigators and expanded the area of research on cell–cell communication via membrane channels. In addition, several rumors indicate that Panxs also form GJ channels and studies may soon be reported clarifying this gossip. This special issue is composed of original and review articles that correspond to a small sample of progress in specific subareas of the field and undoubtedly underscore the overall advances made thus far.

It is worth emphasizing, however, especially for the students, postdoctoral fellows and scientists from other areas who have turned their attention to this field, that there is yet much to do to answer the fundamental questions of the day. Some of the most attractive questions are: What other diseases are linked to Cx/Panx defects? What are the mutations in Cxs and Panxs telling us about the functional roles of these proteins in living cells? What are we learning as the structural models for Cx/Panx channels are refined further? What may the development of pharmacological approaches to Cx/Panx-based channels reveal about: a) the physiological roles of these channels and b) design therapies for different human diseases? And the list goes on. For relevant progress on the last question, see in this issue: Dahl et al., 2013; Verselis and Srinivas, 2013; Iyyathurai et al., 2013; Liu et al., 2013; Riquelme et al., 2013a).

We would like to highlight another set of topics in the field. In this case, the questions might not be obvious, but we believe they are just as important. We suggest that in all fields of biological research, investigators are dealing with an array of complex questions, ongoing studies and dreams of identifying responsible regulatory mechanisms and functions. Obviously, certain aspects are better understood than others. Embroiled in their research, as they seek to build models, there is a tendency for investigators to simplify some aspects that would benefit from a more detailed analysis. Thus, the current understanding of particular issues is not based on thorough experimental demonstrations, but rather on assumptions that fit well with one's favorite idea. Over time, these simplifications have a way of sending down roots and pervading the thinking in the field – even reaching into academic classrooms, textbooks and presentations at scientific meetings.

Along with many other areas in biology, the field of cell communication, GJs and HCs has been impacted in this way.

An excellent reminder comes from the history of work on HCs over the last ~15 years. This history illustrates how we can be misled when we rely on only theoretical analyses or an incomplete set of experimental data relevant to a fundamental question. For many years, the idea had been that any significant opening of HCs would lead to a major cell catastrophe. And, in fact, the opening of HCs for extended periods of time can be deleterious and in injured cells can accelerate the process of cell death (see in this issue: Orellana et al., 2013; Kozoriz et al., 2013). However, and this was completely unexpected, we have learned that HCs do open to the external milieu with high open probability in healthy cells (Franco et al., 2001; Pearson et al., 2005; Schalper et al., 2008; Huckstepp et al., 2010a; Orellana et al., 2011; Riquelme et al., 2013b), even *in vivo* (Huckstepp et al., 2010b; Li et al., 2011; Riquelme et al., 2013b), where they play essential roles in a variety of cellular processes. Of critical importance, the health of these cells does not appear to be compromised as previously hypothesized. Thus, future theoretical and experimental analysis will be required to explain how cells handle the predicted detrimental conditions caused by HC openings. An even more unexpected finding is that Cx43 HCs have been found to confer resistance to glioblastoma multiform upon treatment with temozolomide used in chemotherapy (Gielen et al., 2013).

We take this opportunity to identify a few ideas in the GJ/HC/Cx/Panx field that are widely held, but not actually supported by a rigorous set of experimental data. Certainly, different investigators describe these ideas in different ways in the literature and at scientific meetings. However, our overall treatment of them and our growing understanding of these topics would benefit from being alert to both the limitations of the existing data and the assumptions incorporated into our models. In the spirit of constructive scientific exchange, we offer the following examples of topics that deserve cautious treatment and further scientific scrutiny.

Common Belief 1: A popular Idea is that a wide variety of small molecules of biological importance pass between cells via GJs and through the cell membrane via HCs.

Problem 1: We obviously do not know at this time what different molecules pass and which ones are excluded by each channel type.

Problem 2: We still do not know the intrinsic permeability properties of GJ channels and high resolution structural features need to be improved; therefore, we cannot predict what passes through and what is excluded by these channels.



Contents lists available at SciVerse ScienceDirect

Neuropharmacology

journal homepage: www.elsevier.com/locate/neuropharm

Cerebral ischemic injury is enhanced in a model of oculodentodigital dysplasia

Michael G. Kozoriz^a, Simon Lai^a, José L. Vega^{b,c}, Juan C. Sáez^{c,d}, Wun Chey Sin^a, John F. Bechberger^a, Christian C. Naus^{a,*}^a Department of Cellular and Physiological Sciences, Life Sciences Institute, University of British Columbia, Vancouver, BC, Canada^b Experimental Physiology Laboratory (EPhyL), Instituto Antofagasta, Antofagasta, Chile^c Departamento de Fisiología, Pontificia Universidad Católica de Chile, Santiago, Chile^d Instituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Valparaíso, Chile

ARTICLE INFO

Article history:

Received 2 January 2013

Received in revised form

1 May 2013

Accepted 4 May 2013

Keywords:

Astrocytes

Stroke

Rare genetic disease

Connexin43

Hemichannels

Gap junctional communication

ABSTRACT

Oculodentodigital dysplasia (ODDD) is a rare autosomal dominant disease that results in visible developmental anomalies of the limbs, face, eyes and teeth. Recently analysis of human connexin43 (Cx43) DNA sequences has revealed a number of different missense, duplication and frame shift mutations resulting in this phenotype. A mouse model of this disorder has been created with a missense point mutation of the glycine amino acid at position 60 to serine (G60S). Heterozygote +/G60S mice exhibit a similar ODDD phenotype as observed in humans. In addition to the malformations listed above, ODDD patients often have neurological findings. In the brain, Cx43 is highly expressed in astrocytes and has been shown to play a role in neuroprotection. We were interested in determining the effect of the +/G60S mutation following stroke. Four days after middle cerebral artery occlusion the volume of infarct was larger in mice with the +/G60S mutation. In astrocyte–neuron co-cultures, exposure to glutamate also resulted in greater cellular death in the +/G60S mutants. Protein levels of Cx43 in the mutant mouse were found to be reduced when compared to the normal tissue. Cx43 protein was observed as a continual line of small punctate aggregates in the plasma membrane with increased intracellular localization, which is distinct from the larger plaques seen in the normal mouse astrocytes. Functionally, primary +/G60S astrocytes exhibited reduced gap junctional coupling and increased hemichannel activity, which may underlie the mechanism of increased damage during stroke.

This article is part of a Special Issue entitled ‘Connexin based channels’.

Crown Copyright © 2013 Published by Elsevier Ltd. All rights reserved.

1. Introduction

Oculodentodigital dysplasia (ODDD) was first described by Meyer-Schwickerath et al. (1957) and further defined by Gorlin

et al. (1963). Common external features in this syndrome include ocular, nasal, dental and digital abnormalities (Paznekas et al., 2009). It is now known that this syndrome is caused by mutation of the gene encoding the gap junction protein connexin43 (Cx43). Currently, 62 different Cx43 mutations have been identified in ODDD patients, and in the majority of cases the mutation is dominant (Paznekas et al., 2009).

Cx43 is expressed in numerous tissues and plays an important role in cellular communication. Cx43 proteins oligomerize into hexamers to form connexons, which are inserted into the plasma membrane of single cells to form hemichannels, or coupled to the connexons in neighboring cells to form gap junction channels that provide cytoplasmic continuity between cells (Sáez et al., 2003). The effect of ODDD Cx43 mutations has been studied by several groups and in general these mutants show reduced gap junction formation and increased hemichannel activity (Dobrowolski et al., 2007; McLachlan et al., 2005). Also, mutated Cx43 often exerts a

Abbreviations: BSA, bovine serum albumin; Cx43, connexin43; DAPI, 4',6-diamidino-2-phenylindole; DMEM, Dulbecco's modified Eagle's medium; EBSS, Earle's balanced salt solution; Etd, ethidium; PI, propidium iodide; La³⁺, lanthanum ion; G60S, missense point mutation of the glycine amino acid at position 60 conversion to serine; GFAP, glial fibrillary acidic protein; HBSS, Hank's buffered salt solution; IBA-1, ionized calcium-binding adaptor molecule 1; LDH, lactate dehydrogenase; MCAO, middle cerebral artery occlusion; ODDD, oculodentodigital dysplasia; RIPA, radioimmune precipitation lysis buffer; TBS-T, Tris-buffered saline with tween.

* Corresponding author. Department of Cellular and Physiological Sciences, Life Sciences Institute, University of British Columbia, 2350 Health Sciences Mall, Rm. 1352, Vancouver, BC V6T 1Z3, Canada. Tel.: +1 604 827 4383; fax: +1 604 827 3922.

E-mail address: cnaus@interchange.ubc.ca (C.C. Naus).

1

C

Ca²⁺ Activation of K⁺ Channels: RCK Domains

Ramon Latorre and David Baez-Nieto
Centro Interdisciplinario de Neurociencia de
Valparaíso, Universidad de Valparaíso, Valparaíso,
Chile

Introduction

Some K⁺ channels can be activated by a rise in cytosolic Ca²⁺ and, in two cases (the K⁺ channels from the archaean *Methanobacterium thermoautotrophicum* (MthK) and the large conductance voltage- and Ca²⁺-activated K⁺ (BK) channel), the Ca²⁺-binding sites are contained within the *regulatory domains for K⁺ conductance* (RCK). The MthK channel is a tetramer formed by subunits containing two transmembrane domains and a C-terminal domain containing a single RCK domain (Jiang et al. 2002). The functional MthK channel, however, contains eight RCK domains, and the green domains in Fig. 1a come from the cytoplasmic milieu.

The structure of the RCK domain of a six transmembrane domain K⁺ channel from *E. coli* (solved at 2.4 Å resolution) has a Rossmann-fold topology, which is a very common structural motif of enzymes and ligand-binding proteins (see ► **Structural Motifs**). Rossmann-fold secondary structures are organized into two linked β-α-β-α-β units (see Fig. 1b) and were first identified in a number of NAD⁺-dependent dehydrogenases. This is the type of structure present in the MthK and in the *Drosophila*, mouse, and human Slo1 (commonly known as BK; see ► **Potassium Channels**

(Kv, Kir, KCa, and K(2P) Channels) channel (Fig. 1b).

The cloning of the BK channel from *Drosophila* showed that it is a member of the S4 superfamily encompassing voltage-dependent K⁺ (Kv), Na⁺, and Ca²⁺ channels. The gene coding for BK was called *Slowpoke* or *Slo* (later renamed *Slo1* after the cloning and expression of *Slo2* and *Slo3* (Salkoff et al. 2006)). In the case of Kv channels, the channel-forming protein possesses six transmembrane domains (S1–S6) containing the pore-forming domain S5–P–S6, and an S4 voltage-sensing element (S1–S4). As Kv channels, the BK channel is a tetramer; however, unlike Kv channels, the BK channel protein consists of seven transmembrane (S0–S6) domains with an exoplasmic N-terminus (Fig. 1c). The intracellular C-terminal domain, comprising two thirds of the protein, contains four hydrophobic segments (S7–S10) and the Ca²⁺- and Mg²⁺-binding sites. The C-terminus of BK channels consists of two tandem RCK domains (only RCK1 is shown in Fig. 1b). The RCK domain in the BK channel was initially unveiled by MacKinnon's group (Jiang et al. 2001) by multiple sequence alignment of the BK channel with prokaryotic K⁺ channels and other proteins known to possess the RCK domain motif. Based on their primary sequence, the C-terminal domains of K⁺ channels *Slo2* and *Slo3* are believed to be structured as two RCK domains in tandem (Salkoff et al. 2006). Unlike *Slo1*, however, *Slo2* is activated by internal Na⁺ and Cl[−], and *Slo3* is activated by protons. The actual structural nature of the C-termini of these two channels is unknown at present and will not be discussed further here.

Keeping you healthy: BK channel activation by omega-3 fatty acids

Ramon Latorre and Gustavo Contreras

Centro Interdisciplinario de Neurociencia de Valparaíso, Facultad de Ciencias, Universidad de Valparaíso, Valparaíso 2366103, Chile

In the exuberant world of K⁺ channels, the Ca²⁺- and voltage-activated K⁺ (BK, MaxiK, Slo1) channel stands alone. It is coded by a single gene (*slo1* or *KCNMA1*), and the pore-forming α subunit has seven transmembrane segments instead of six as found in voltage-dependent K⁺ channels (Atkinson et al., 1991; Adelman et al., 1992; Butler et al., 1993; Wallner et al., 1996). Being activated by depolarizing voltages and cytoplasmic Ca²⁺, the BK channel is the perfect molecular machine to retard or to simply stop excitatory signals. The negative feedback (hyperpolarization) created by the opening of these K⁺ channels is caused by the perfect tuning between Ca²⁺ and voltage sensors. The communication between these two types of sensors is allosterically established, that is, voltage or internal Ca²⁺ alone is able to open the BK channel, but channel opening is increasingly facilitated as more Ca²⁺ and voltage sensors are activated (Horrigan and Aldrich, 2002; Horrigan, 2012) (Fig. 1 A). Another important difference between BK channels and Kv channels, where opening is tightly coupled to voltage-sensor activation (Soler-Llavina et al., 2003) (Fig. 1 B), is that, albeit with a very low probability, BK channels can open in a voltage sensor- and Ca²⁺-independent manner (reaction C \leftrightarrow O defined by the equilibrium constant *L* in Fig. 1 A).

Despite being coded by a single gene, BK channel diversity is large. Alternative splicing, posttranslational modifications, and/or the presence of modulatory β or γ subunits create this diversity (Orío et al., 2002; Salkoff et al., 2006; Yan and Aldrich, 2010, 2012). In particular, modifications induced in BK gating kinetics by the β 1, β 2, and β 4 subunits proved to be of crucial importance in many physiological processes, ranging from shaping neuronal excitability and neurosecretion to smooth muscle tone, and in others not so physiological, such as alcohol tolerance (Brenner et al., 2000; Hu et al., 2001; Gollasch et al., 2002; Grimm and Sansom, 2007; Martin et al., 2008). The expression of β subunits is highly tissue specific; β 1 is the only β subunit expressed in smooth muscle, and β 4 is mainly expressed in the nervous system (Orío et al., 2002; Wu and Marx, 2010). In vascular smooth muscle cells (SMCs), the presence of β 1 plays a vital role in vasoregulation, and its lack leads to hypertension (Brenner et al., 2000; Fernández-Fernández et al., 2004; Nelson and Bonev, 2004). β 1 and β 2 have been observed to dramatically slow down activation and deactivation kinetics as well as

increase the apparent Ca²⁺ sensitivity of the BK channel. Although β 4 also decelerates BK activation and deactivation kinetics, even more so than β 1, Ca²⁺ sensitivity of channels formed by α/β 4 subunits is complex. Channels are less sensitive to Ca²⁺ at low internal Ca²⁺ concentrations (<10 μ M) than channels formed by the α subunit alone. However, Ca²⁺ is more effective in activating α/β 4 channels at higher Ca²⁺ concentrations (Ha et al., 2004; Wang et al., 2006).

In addition to their effects on channel gating, β subunits grant BK channels sensitivity to several physiologically important compounds, thus making these subunits targets for possible pharmacological interventions. For example, β 1-containing BK channels but not channels formed by the α subunit alone appear to be the target of 17 β -estradiol and other compounds such as estrogen analogues, anti-estrogens, and the bile salt component lithocholic acid (Valverde et al., 1999; Dick et al., 2001; Bukiya et al., 2009; Maher et al., 2013). The activation of BK channels by 17 β -estradiol has been proposed as the possible mechanism that mediates the acute relaxation of vascular smooth muscle induced by the hormone (White et al., 1995; Ruehlmann et al., 1998). On the other hand, stress steroids activate channels formed by the α/β 4 complex but not by α/β 2 (King et al., 2006). Polyunsaturated fatty acids such as arachidonic acid (AA) are also able to directly activate BK channels, but in this case, AA enhances BK current in the presence of either β 2 or β 3 (Sun et al., 2007). Findings by Sun et al. (2007) also show that AA is able to remove inactivation, suggesting that this fatty acid is interacting with the β 2-inactivating peptide. Tissue specificity of β subunits and their particular capacity to endow BK channels with different pharmacological profiles have greatly increased the importance of BK channels in maintaining the adequate cellular electrical homeostasis in different tissues.

Docosahexaenoic acid (DHA), an omega-3 fatty acid known to be associated with beneficial cardiovascular effects, was reported to be a potent activator of BK currents in rat coronary artery SMCs and to promote dilation of isolated small coronary arteries (Lai et al., 2009;

© 2013 Latorre and Contreras. This article is distributed under the terms of an Attribution-Noncommercial-Share Alike-No Mirror Sites license for the first six months after the publication date (see <http://www.rupress.org/terms>). After six months it is available under a Creative Commons License (Attribution-Noncommercial-Share Alike 3.0 Unported license, as described at <http://creativecommons.org/licenses/by-nc-sa/3.0/>).

Signal Transduction–Dependent Channels

4

Ramon Latorre, Carlos González, and Patricio Rojas

Brief History

From the moment the first member of this ion channel family, Slo1, was discovered, the scientific world was confronted with a molecular Pandora's box: once opened, its electrical language left scientists bewitched. They were fascinated with this “monster” of a single-channel conductance (250 pS in symmetrical 100 mM K⁺) close to the ceiling imposed by simple diffusion combined with an exquisite K⁺ selectivity. Slo1 channels are essentially impermeable to Na⁺ and conduct K⁺ 10- and 200-fold more effectively than Rb⁺ and Cs⁺, respectively, though it was previously thought that large conductance channels were not supposed to be so selective. At the same time, the channel was activated by voltage and cytoplasmic Ca²⁺. This latter property led Meech in 1978 to hypothesize that this conductance system was perfect link between cell metabolism and electrical activity, and he was right on the mark. Because of its large conductance, this voltage- and calcium-activated K⁺ channel was christened “maxi-K” or “BK” (for big K⁺). This high single-channel

R. Latorre (✉)

Centro de Neurociencia, Universidad de Valparaíso, Valparaíso, Chile

e-mail: ramon.latorre@uv.cl

C. González

Centro Interdisciplinario de Neurociencia de Valparaíso, Universidad de Valparaíso Facultad de Ciencias, Valparaíso, Chile

e-mail: carlos.gonzalez@cinv.cl

P. Rojas

Departamento de Biología, Facultad de Química y Biología Universidad de Santiago de Chile, La Serena, Chile

e-mail: patricio.rojas.m@usach.cl

D.W. Pfaff (ed.), *Neuroscience in the 21st Century*,

DOI 10.1007/978-1-4614-1997-6_5, © Springer Science+Business Media, LLC 2013

81

a0010

Voltage-Dependent K⁺ Channels

Au1,2

R Latorre, F J Morera, and C Zaelzer

© 2012 Elsevier Inc. All rights reserved.

This article is a revision of the previous edition article by Ramón Latorre and Francisco J Morera, volume 1, pp. 399–404. © 2004, Elsevier Inc.

Glossary

dt0010

Delayed rectifier A K⁺ channel that changes the membrane conductance with a delay after a depolarizing voltage step.

dt0015

Gating The opening or closing of a channel in response to some stimulus.

dt0020

Inward rectifier A K⁺ channel that opens when the membrane is hyperpolarized.

Long QT syndrome An inherited cardiac arrhythmia.

dt0025

Missense mutation A mutation in which an incorrect amino acid is incorporated into the protein.

dt0030

Voltage sensor A channel structure able to detect changes in membrane potential. In voltage-dependent ion channels, the voltage-sensing elements are located in the S4 segment.

dt0035

s0010

Structure and Diversity

p0010

Voltage-dependent K⁺ channels are members of the voltage-dependent cation channel family, which includes the voltage-dependent Na⁺, Ca²⁺, and K⁺ channels. The first voltage-dependent K⁺ channel was cloned in 1987 from a fruit fly, *Drosophila*, based on a mutation that causes the *Shaker* phenotype. After that, they have been identified in an immense number of organisms including prokaryotes and eukaryotes.

p0015

The 78 members of the K⁺ channel superfamily can be divided into four structural types based on their mode of activation and the number of their transmembrane segments: (1) voltage-gated six-transmembrane K⁺ channels (K_v), (2) voltage- and Ca²⁺-activated seven-transmembrane K⁺ channels (K_{Ca}), (3) Inward rectifying two-transmembrane K⁺ channels (K_{ir}), and (4) two-pore four-transmembrane K⁺ channels (K_{2P}). Only the K_v and K_{Ca} families have channel-forming proteins containing intrinsic voltage sensors (see below).

p0020

All the voltage-dependent K⁺ channels are tetrameric assemblies. In the K_v channels, each subunit consists of six hydrophobic segments, S1–S6. The primary sequence of these hydrophobic segments shows similarities between all of the K_v channels, including the voltage-sensor domain (VSD, S1–S4) and the ion-conduction pore domain (S5–S6) (**Figure 1(a)**).

Au3

Au4

p0025

All K⁺ channels have a consensus amino acid sequence inside the pore region, TVGYGD, dubbed the 'signature sequence'. The residues, TVGYG, line the selectivity filter (in some K⁺ channels, the tyrosine (Y) residue is replaced by phenylalanine (F), e.g., in the ether-a-go-go (eag) channel discussed later on). In the selectivity filter, carbonyl oxygen atoms are directed toward the pore to coordinate dehydrated K⁺ ions. Hydrophobic chains of valine and tyrosine directed toward the hydrophobic core surrounding the filter stabilize the main chain.

p0030

Gating in the K_v channels is conferred through the attachment of gating domains to the pore. The basic function of these gating domains is to perform mechanical work on the ion-conduction pore to change its conformation between closed and opened states. In voltage-dependent channels, a VSD (S1–S4) is present on each subunit. Thus, a voltage sensor

converts energy stored in the membrane electric field into mechanical work. There is strong evidence that the positive charges contained in S4 are the voltage-sensing elements. Thus, ion-channel gating is essentially an electromechanical coupling between a gating unit and a pore unit.

The crystal structure of a mammalian voltage-dependent K⁺ channel (K_v1.2) had initially been resolved at 2.9 Å and further improved to 2.4 Å using a chimeric K_v1.2–K_v2.1 channel. In the latter case, the channel was crystallized in complex with lipids. These structures showed that the helices of the ion-conduction pore (S5–S6) related to the helices of the VSD (S1–S4) in a special way. The VSD of one subunit is located near the pore domain of an adjacent subunit (**Figure 1(b)**). The connection between the pore and the VSD is made by the S4–S5 linker helix, which runs parallel to the intracellular membrane surface.

Voltage-dependent K⁺ channels are ubiquitously distributed in different cells and tissues, therefore, K⁺ channel diversity is of great importance in determining the variety of electrical responses produced by cells when subjected to stimuli. The possible mechanisms that originate the immense voltage-dependent K⁺ channel diversity are: (1) multiple genes; (2) alternative splicing; (3) formation of heteromultimeric channels; and (4) coexpression with regulatory β-subunits.

Voltage-dependent K⁺ channel diversity seems to have accompanied animal cells through evolution. Some K⁺ channels are the most conserved proteins in eukaryotes. Potassium channels became fundamental to animal cell physiology very early in evolution, and through voltage-dependent K⁺ channel control of membrane potential they couple the cell inner dynamics to the outer environment. A differential expression of voltage-dependent K⁺ channel messenger RNAs (mRNAs) is found to accompany the events of animal development and also occurs in adult animal tissues.

The diversity of voltage-dependent K⁺ channel genes is shown in the dendrogram shown in **Figure 2**. The dendrogram shows the major classes of voltage-dependent K⁺ channels. The ramification of voltage-dependent K⁺ channels was constructed by carrying out sequence alignments. To compare the amino acid sequences of the different voltage-dependent K⁺ channels, the sequences were aligned using at least 300 amino acids from six hydrophobic segment (S1–S6) channels and

p0035

p0040

p0045

p0050

REM SLEEP PHASE PREFERENCE IN THE OCTODON DEGUS

<http://dx.doi.org/10.1002/cnrv.23000>REM Sleep Phase Preference in the Crepuscular *Octodon degus* Assessed by Selective REM Sleep DeprivationAdrián Ocampo-Garcés, MD, PhD¹; Felipe Hernández MSc¹; Adrian G Palacios, PhD²¹Programa de Fisiología y Biofísica, Instituto de Ciencias Biomédicas, Facultad de Medicina, Universidad de Chile, Santiago, Chile; ²Centro Interdisciplinario de Neurociencia de Valparaíso, Facultad de Ciencias, Universidad de Valparaíso, Valparaíso, Chile**Study Objectives:** To determine rapid eye movement (REM) sleep phase preference in a crepuscular mammal (*Octodon degus*) by challenging the specific REM sleep homeostatic response during the diurnal and nocturnal antirepuscular rest phases.**Design:** We have investigated REM sleep rebound, recovery, and documented REM sleep propensity measures during and after diurnal and nocturnal selective REM sleep deprivations.**Subjects:** Nine male wild-captured *O. degus* prepared for polysomnographic recordings**Interventions:** Animals were recorded during four consecutive baseline and two separate diurnal or nocturnal deprivation days, under a 12:12 light-dark schedule. Three-h selective REM sleep deprivations were performed, starting at midday (zeitgeber time 6) or midnight (zeitgeber time 18).**Measurements and Results:** Diurnal and nocturnal REM sleep deprivations provoked equivalent amounts of REM sleep debt, but a consistent REM sleep rebound was found only after nocturnal deprivation. The nocturnal rebound was characterized by a complete recovery of REM sleep associated with an augment in REM/total sleep time ratio and enhancement in REM sleep episode consolidation.**Conclusions:** Our results support the notion that the circadian system actively promotes REM sleep. We propose that the sleep-wake cycle of *O. degus* is modulated by a chorus of circadian oscillators with a bimodal crepuscular modulation of arousal and a unimodal promotion of nocturnal REM sleep.**Keywords:** crepuscular chronotype, *Octodon degus*, REM sleep, REM sleep homeostasis, REM sleep rebound**Citation:** Ocampo-Garcés A; Hernández F; Palacios AG. REM sleep phase preference in the crepuscular *Octodon degus* assessed by selective REM sleep deprivation. *SLEEP* 2013;36(8):XXX-XXX.

INTRODUCTION

Crepuscular mammals exhibit a characteristic bimodal profile of activity, with activity bouts peaking just before dawn (morning peak) and dusk (evening peak). The crepuscular chronotype of rest-activity rhythm is widely distributed among mammals and has been reported in both free-ranging and captive animals.¹⁻³ Physiological and behavioral measures such as body temperature,⁴ wheel-running,^{5,6} cortisol secretion,⁷ and electroencephalographic (EEG) data⁸ have been also found to display bimodal temporal profiles among crepuscular species.

A crepuscular activity pattern has been described in *Octodon degus* (Rodentia: Hystricognatha) living in natural conditions and confinement.^{1,9} Polysomnographic recordings have shown a bimodal crepuscular profile of wakefulness, with phase-opposed modes at zeitgeber time (ZT)11 and ZT23 under a 12:12 light-dark cycle.¹⁰ Wake bouts delimit two sleep-predominant rest plateaus occurring outside the crepuscular periods (antirepuscular rest phases, as defined by McElhinny et al.⁴). No significant diurnal-nocturnal difference has been found for the amount of rapid eye movement (REM) or non-REM (NREM) sleep,^{10,11} casting doubts on the idea of a diurnal or nocturnal phase sleep state preference. The crepuscular bimodal sleep-wake pattern departs

from the typical unimodal profile observed among nocturnal and diurnal mammals. In particular, REM sleep of diurnal and nocturnal mammals is under strong unimodal circadian modulation, with the acrophase coinciding with core body temperature nadir.^{12,13} The opponent process model proposes that circadian modulation of the sleep-wake cycle is explained by an active promotion of arousal by the circadian system that opposes the homeostatic sleep pressure accumulated during wakefulness.¹⁴ If this is true, diurnal and nocturnal antirepuscular sleep-predominant rest intervals observed in *O. degus* occur permissively, mirroring the bimodal crepuscular promotion of arousal.

However, some evidence suggests that REM sleep may be actively promoted by the circadian system, as homeostatic REM sleep recovery after sleep deprivation is facilitated during the corresponding rest phase and impaired during the active phase.^{15,16} To explore phase-dependent REM sleep modulation in the crepuscular *O. degus*, in this report we take advantage of a property of REM sleep: REM sleep quickly compensates for state debts after selective REM sleep deprivation during the rest phase, as has been observed in the rat.^{15,17,18} If REM sleep during the antirepuscular rest phases occurs permissively, without circadian REM sleep promotion, as predicted by the opponent process model, REM sleep deprivation during those intervals would be compensated for by an equivalent homeostatic response. Alternatively, asymmetric diurnal versus nocturnal REM sleep homeostatic responses must be interpreted as the manifestation of phase-specific REM sleep promotion, supporting the notion that sleep states are actively promoted by the circadian system.

METHODS

Animal handling and experimentation was performed according to local institutional animal care guidelines and autho-

Submitted for publication July, 2012

Submitted in final revised form December, 2012

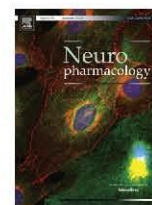
Accepted for publication January, 2013

Address correspondence to: Adrián Ocampo-Garcés, MD, PhD, Programa de Fisiología y Biofísica, Instituto de Ciencias Biomédicas, Facultad de Medicina, Universidad de Chile, Avda. Independencia 1027, Casilla 70005, Postal Code: 8380453, Santiago, Chile; Tel: (56 2) 678 6422; Fax: (56 2) 777 6916; E-mail: aocampo@med.uchile.cl

⁹⁰
SLEEP, Vol. 36, No. 8, 2013

1

REM Sleep in a Crepuscular Mammal—Ocampo-Garcés et al



Invited review

Gap junction channels and hemichannels in the CNS: Regulation by signaling molecules

Juan A. Orellana^{a,*}, Agustín D. Martínez^b, Mauricio A. Retamal^c^aDepartamento de Neurología, Escuela de Medicina, Pontificia Universidad Católica de Chile, Marcoleta 391, Santiago, Chile^bCentro Interdisciplinario de Neurociencias de Valparaíso (CINV), Universidad de Valparaíso, Chile^cDepartamento de Fisiología, Facultad de Medicina, Clínica Alemana-Universidad del Desarrollo, Santiago, Chile

ARTICLE INFO

Article history:

Received 18 September 2012

Received in revised form

8 February 2013

Accepted 26 February 2013

Keywords:

Connexins

Pannexins

Paracrine factors

Brain

Astrocytes

Microglia

Neurons

ABSTRACT

Coordinated interaction among cells is critical to develop the extremely complex and dynamic tasks performed by the central nervous system (CNS). Cell synchronization is in part mediated by connexins and pannexins; two different protein families that form gap junction channels and hemichannels. Whereas gap junction channels connect the cytoplasm of contacting cells and coordinate electric and metabolic activities, hemichannels communicate intra- and extra-cellular compartments and serve as diffusional pathways for ions and small molecules. Cells in the CNS depend on paracrine/autocrine communication via several extracellular signaling molecules, such as, cytokines, growth factors, transmitters and free radical species to sense changes in microenvironment as well as to adapt to them. These signaling molecules modulate crucial processes of the CNS, including, cellular migration and differentiation, synaptic transmission and plasticity, glial activation, cell viability and microvascular blood flow. Gap junction channels and hemichannels are affected by different signaling transduction pathways triggered by these paracrine/autocrine signaling molecules. Most of the modulatory effects induced by these signaling molecules are specific to the cell type and the connexin and pannexin subtype expressed in different brain areas. In this review, we summarized and discussed most of the relevant and recently published information on the effects of signaling molecules on connexin or pannexin based channels and their possible relevance in CNS physiology and pathology.

This article is part of the Special Issue Section entitled 'Current Pharmacology of Gap Junction Channels and Hemichannels'.

© 2013 Elsevier Ltd. All rights reserved.

1. Introduction

Coordinated interaction among cells is critical to perform the extremely complex and dynamic tasks performed by the brain. Cell ability to sense local and neighboring microenvironments has evolved in different ways in more complex organisms. In vertebrates, cell interaction and synchronization is in part mediated by intercellular communication via connexin- and pannexin-based channels. Connexins and pannexins comprise two different gap junction protein families, which in mammals are composed of about 20 and 3 members, respectively (Abascal and Zardoya, 2012). Eumetazoans, with the only exception of echinoderms, express pannexins (called innexins in non-chordates), whereas the connexin family is exclusive to chordates (Abascal and Zardoya, 2012; Phelan and Starich, 2001; Shestopalov and Panchin, 2008). Despite the fact that connexins and pannexins do not share a relevantly

homologous primary structure, they have similar secondary and tertiary structures with four α -helical transmembrane domains, connected by one cytoplasmic and two extracellular loops, where both N- and C-termini are intracellular (Fig. 1). Pannexins and connexins oligomerize into hexamers to constitute single hemichannels, except for pannexin2 (Pannx2), which seems to form octamers (Ambrosi et al., 2010).

After assembly, connexin hemichannels are transported to the non-junctional plasma membrane and diffuse laterally to dock with connexin hemichannels from a neighboring cell to form gap junction channels (Sáez et al., 2003a) (Fig. 1). Gap junctions are aggregates of these intercellular channels and mediate an important form of direct intercellular communication in the animal kingdom. Gap junction channels favor the intercellular exchange of metabolites (e.g., ADP, glucose, glutamate and glutathione), second messengers (e.g., cAMP and IP₃) and ions, allowing the intercellular spread of electrotonic potentials in excitable and non-excitable tissues (Evans et al., 2006; Sáez et al., 2003a; Sohl et al., 2005). For a long time, the main function attributed to connexin

* Corresponding author. Tel.: +56 2 23546951.
E-mail address: jaorella@uc.cl (J.A. Orellana).

Pannexin1 hemichannels are critical for HIV infection of human primary CD4⁺ T lymphocytes

J. A. Orellana,^{*} S. Velasquez,[†] D. W. Williams,[‡] J. C. Sáez,^{§,||} J. W. Berman,^{‡,¶}
and E. A. Eugenin^{†,*,1}

^{*}Departamento de Neurología, Escuela de Medicina, and [§]Departamento de Fisiología, Pontificia Universidad Católica de Chile, Santiago, Chile; [†]Public Health Research Institute and [¶]Department of Microbiology and Molecular Genetics, University of Medicine & Dentistry of New Jersey, Newark, New Jersey, USA; Departments of [‡]Pathology and [¶]Microbiology and Immunology, Albert Einstein College of Medicine, Bronx, New York, USA; and ^{||}Instituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Chile

RECEIVED MAY 24, 2012; REVISED JANUARY 17, 2013; ACCEPTED JANUARY 18, 2013. DOI: 10.1189/jlb.0512249

ABSTRACT

HIV is a major public health issue, and infection of CD4⁺ T lymphocytes is one of its key features. Whereas several cellular proteins have been identified that facilitate viral infection and replication, the role of hemichannels in these processes has not been fully characterized. We now show that the HIV isolates, R5 and X4, induced a transient-early (5–30 min) and a later, persistent (48–120 h) opening of Panx1 hemichannels, which was dependent on the binding of HIV to CD4 and CCR5/CXCR4 receptors. Blocking Panx1 hemichannels by reducing their opening or protein expression inhibited HIV replication in CD4⁺ T lymphocytes. Thus, our findings demonstrate that Panx1 hemichannels play an essential role in HIV infection. *J. Leukoc. Biol.* 94: 399–407; 2013.

Introduction

HIV infects mostly immune cells by binding the viral env protein gp120 to the host cellular proteins, CD4 and CCR5 and/or CXCR4, resulting in fusion of the viral env with the cellular membrane. To date, in addition to CD4, CCR5, and/or CXCR4, no other plasma membrane proteins have been identified to participate directly in the process of viral entry [1]. However, a few studies indicated that upon binding of the virus to its cellular receptors, intracellular-free Ca²⁺ levels rise [2–4], and opening of nonselective cation channels, as well as Ca²⁺-activated K⁺ channels, occurs [5], suggesting that

signaling and activation of other proteins may be required for infection and replication. Recently, studies in cell lines, PBMCs, and human macrophages indicated that ATP release through pannexin-1 (Panx1) hemichannels is required for HIV replication [6, 7], supporting the hypothesis that additional host proteins are required for infection/replication.

Hemichannels are plasma membrane channels that can be opened at the unapposed cell surface, forming aqueous conduits permeable to ions and small molecules (e.g., ATP, glutamate, NAD⁺, and PGE₂). They allow diffusional exchange between the intra- and extracellular compartments, constituting a route for autocrine/paracrine cellular communication [8]. Hemichannels are constituted by the oligomerization of six protein subunits, termed Cxs or Panxs, both highly conserved protein families encoded by 21 or three genes in humans, respectively [9, 10]. Panx1 hemichannels, in concert with purinergic receptors, have been described to be important in different immune functions, including cellular activation [11–13], apoptosis [14], stress signals [15], secretion of inflammatory cytokines [16], and HIV replication [7]. However, how HIV infection changes the opening of these channels in primary human CD4⁺ T lymphocytes—one of the main targets of HIV—remains to be elucidated. Here, we show that depending on the HIV isolate, the interaction of CD4 and CCR5 and/or CXCR4 is crucial for the biphasic Panx1 hemichannel opening induced by HIV in human primary CD4⁺ T lymphocytes. We also found that chemokines that bind CCR5 and CXCR4 increase Panx1 hemichannel activity, but only early, and more transiently, as compared with the biphasic opening of Panx1 hemichannels in response to HIV infection. Down-regulation or pharmacological blockade of Panx1 hemichannels inhibited HIV replication in CD4⁺ T lymphocytes,

Abbreviations: AU=arbitrary unit(s), C_T=comparative threshold, Cx=connexin, env=envelope, Etd=ethidium, F1=fluorescence intensity in each cell, FB=background fluorescence-intensity measurement, La³⁺=lanthanum ion, NIAID=National Institute of Allergy and Infectious Diseases, NIMH=National Institute of Mental Health, Panx1=pannexin1, Prob=probenecid, qRT-PCR=quantitative RT-PCR, scr=scrambled, SDF-1α=stroma cell-derived factor-1α, siRNA=small interfering RNA

The online version of this paper, found at www.jleukbio.org, includes supplemental information.

1. Correspondence: Public Health Research Institute and Dept. of Microbiology and Molecular Genetics, UMDNJ, Newark, NJ 07103, USA. E-mail: eugeniea@umdnj.edu; Twitter: http://phri.org/research/res_pieugenin.asp

Calcium-activated chloride channels do not contribute to the odorant transduction current in the marine teleost *Isacia conceptionis*

R. OSORIO AND O. SCHMACHTENBERG*

Centro Interdisciplinario de Neurociencia de Valparaíso, Facultad de Ciencias, Universidad de Valparaíso, Avda. Gran Bretaña 1111, 2360102, Valparaíso, Chile

(Received 6 April 2013, Accepted 11 September 2013)

This study compared the contribution of the Ca^{2+} -activated Cl^- conductance to the electro-olfactogram (EOG) evoked by different odorant classes between the marine Cabinza grunt *Isacia conceptionis* and rainbow trout *Oncorhynchus mykiss*. The Ca^{2+} -activated Cl^- channel blocker niflumic acid significantly diminished odorant responses in *O. mykiss*, but had no effect on the EOG in *I. conceptionis*, supporting the notion that Ca^{2+} -activated Cl^- channels may not operate as odorant transduction current amplifiers in this marine teleost.

© 2013 The Fisheries Society of the British Isles

Key words: fish; olfaction; olfactory receptor neuron; rainbow trout.

Twenty years ago, two independent publications in the journal *Nature* reported the contribution of a Ca^{2+} -activated Cl^- conductance to the odorant transduction cascade in vertebrate olfactory receptor neurons (Kurahashi & Yau, 1993; Lowe & Gold, 1993). This conductance, which had first been described in olfactory cilia from the northern grass frog *Rana pipiens* (Kleene & Gesteland, 1991), has now been established to contribute a significant portion to the odorant transduction current in all terrestrial and freshwater mammals and amphibians tested (Schild & Restrepo, 1998; Kleene, 2008). Recently, the molecular identity of the olfactory Ca^{2+} -activated Cl^- channel has been tentatively identified as anoctamin2 (Ano2) in mammals, also known as TMEM16B (Stephan *et al.*, 2009; Pifferi *et al.*, 2012). In fishes, the largest group of vertebrates, evidence remains limited to one patch-clamp study in dissociated olfactory receptor neurons from the rainbow trout *Oncorhynchus mykiss* (Walbaum 1792), that demonstrated the presence of a Ca^{2+} -activated Cl^- conductance in these cells and its participation in odorant transduction (Sato & Suzuki, 2000). To date, however, no study has addressed the possible role of Ca^{2+} -activated Cl^- channels in the olfactory transduction cascade of marine or saltwater-adapted euryhaline teleosts. Owing to the elevated chloride concentration in the marine environment, c. 500 mM, the chloride reversal potential is shifted to negative values compared to

*Author to whom correspondence should be addressed. Tel.: +56-32-2995505; email: oliver.schmachtenberg@uv.cl

Protocol

Husbandry and Breeding in the *Octodon degu* (Molina 1782)

Adrian G. Palacios¹ and Theresa M. Lee^{2,3}

¹Centro Interdisciplinario de Neurociencia de Valparaíso, Facultad de Ciencias, Universidad de Valparaíso, 2360102 Valparaíso, Chile; ²Department of Psychology and Neuroscience Program, University of Michigan, Ann Arbor, Michigan 48109

The *Octodon degu* is a native rodent species from South America, which lives in colonies with a well-structured social organization grouping of 5–10 young and 2–5 adult animals sharing a burrow system. They show a temperature-dependent diurnal-crepuscular activity pattern. In nature they rarely survive 2 yr, mostly because of predation. However, in captivity, females reproduce for 4–4.5 yr, and both sexes live for 5–7 yr. Males remain fertile until death. Some care is required to maintain healthy degus, particularly breeding females. Here we describe husbandry and breeding guidelines from the experience of the University of Michigan degu colony. With the husbandry practices described here, 90% of pups born in our colony reach maturity (6 mo of age), and no diarrheal diseases are apparent in our adult population.

MATERIALS

It is essential that you consult the appropriate Material Safety Data Sheets and your institution's Environmental Health and Safety Office for proper handling of equipment and hazardous materials used in this protocol.

Reagents

Alfalfa

Animals

There is no commercial breeder selling degus. Laboratories and zoos that maintain colonies are usually able to provide some pairs to begin a breeding group. Degus can also be found in some pet stores in the United States as well as in Europe.

Reagent for acidification of water

Rodent chow (e.g., LabDiet 5001, PMI International)

Wheat-based PMI diet (ProLab RMH 2000 5P06)

Equipment

Bedding (preferably an inert material such as corn cob or paper chips)

Bathing dust (e.g., LM Chinchilla Dust Bath, purchased from pet stores) can also be used (see Step 1).

Breeding cages (opaque polypropylene; 20 in × 20 in × 8 in)

³Correspondence: terrilee@umich.edu

© 2013 Cold Spring Harbor Laboratory Press

Cite this article as *Cold Spring Harb Protoc*; 2013; doi:10.1101/pdb.prot073577





SVCT2 vitamin C transporter expression in progenitor cells of the postnatal neurogenic niche

Patricia Pastor¹, Pedro Cisternas¹, Katterine Salazar¹, Carmen Silva-Alvarez¹, Karina Oyarce¹, Nery Jara¹, Francisca Espinoza¹, Agustín D. Martínez² and Francisco Nualart^{1*}

¹ Laboratory of Neurobiology and Stem Cells, Department of Cellular Biology, Center for Advanced Microscopy CMA BIOBIO, University of Concepción, Concepción, Chile

² Centro Interdisciplinario de Neurociencia de Valparaíso, Facultad de Ciencias, Universidad de Valparaíso, Valparaíso, Chile

Edited by:

Nelson Osses, Pontificia Universidad Católica de Valparaíso, Chile

Reviewed by:

Flavia C. A. Gomes, Federal University of Rio de Janeiro, Brazil
Nibaldo C. Inestrosa, Pontificia Catholic University of Chile, Chile

*Correspondence:

Francisco Nualart, Departamento de Biología Celular, Facultad de Ciencias Biológicas, Universidad de Concepción, Casilla 160-C, Concepción, Chile
e-mail: fnualart@udec.cl

Known as a critical antioxidant, recent studies suggest that vitamin C plays an important role in stem cell generation, proliferation and differentiation. Vitamin C also enhances neural differentiation during cerebral development, a function that has not been studied in brain precursor cells. We observed that the rat neurogenic niche is structurally organized at day 15 of postnatal development, and proliferation and neural differentiation increase at day 21. In the human brain, a similar subventricular niche was observed at 1-month of postnatal development. Using immunohistochemistry, sodium-vitamin C cotransporter 2 (SVCT2) expression was detected in the subventricular zone (SVZ) and rostral migratory stream (RMS). Low co-distribution of SVCT2 and β III-tubulin in neuroblasts or type-A cells was detected, and minimal co-localization of SVCT2 and GFAP in type-B or precursor cells was observed. Similar results were obtained in the human neurogenic niche. However, BrdU-positive cells also expressed SVCT2, suggesting a role of vitamin C in neural progenitor proliferation. Primary neurospheres prepared from rat brain and the P19 teratocarcinoma cell line, which forms neurospheres *in vitro*, were used to analyze the effect of vitamin C in neural stem cells. Both cell types expressed functional SVCT2 *in vitro*, and ascorbic acid (AA) induced their neural differentiation, increased β III-tubulin and SVCT2 expression, and amplified vitamin C uptake.

Keywords: SVCT2, vitamin C, brain, niche, stem cells, progenitor, ependymal cells

INTRODUCTION

Active neurogenesis occurs within the anterior wall of the lateral ventricle in the adult mammalian brain (Lois and Alvarez-Buylla, 1993; Doetsch et al., 1999). Neurogenic precursors have also been found in the human brain, specifically located in the periventricular region before 18 months of age (Johansson et al., 1999; Nunes et al., 2003; Sanai et al., 2011; Bergmann et al., 2012). The formation of new neurons, which are β III-tubulin-positive, occurs in restricted, organized compartments termed neurogenic niches (Doetsch et al., 1997, 1999; Alvarez-Buylla and Garcia-Verdugo, 2002; Conover and Notti, 2008; Mirzadeh et al., 2008; Nualart et al., 2012). The neuroblasts formed in this region migrate tangentially in chains throughout the rostral migratory stream (RMS), where the presence of neurogenic progenitors and astrocytes has also been described (Doetsch and Alvarez-Buylla, 1996). The neuroblasts present in the RMS reach the olfactory bulb, where they differentiate into interneurons (Lois et al., 1996; Alvarez-Buylla and Garcia-Verdugo, 2002; Lledo et al., 2008). Ultrastructural, immunohistochemistry, and proliferation analyses of the cytoarchitecture of the neurogenic niche (Doetsch et al., 1997) have revealed the presence of four cell types. B-type cells or astrocytes (GFAP- and nestin-positive) are preferentially located in the subventricular zone (SVZ) and are precursor cells. C-type cells are intermediate transient neuronal cells (nIPC) that proliferate rapidly (Eisch and Mandyam, 2007; Ihrle and Alvarez-Buylla, 2008) and differentiate into neuroblasts or type-A

cells (Doetsch et al., 1999; Tramontin et al., 2003; Chojnacki et al., 2009; Kriegstein and Alvarez-Buylla, 2009; Nualart et al., 2012). E-type cells, which are cube-shaped and multiciliated, are ependymocytes. B-type cells are found in the ependymal layer, projecting cilium to the ventricular lumen, similar to what has been described in the radial glia (Tramontin et al., 2003; Spassky et al., 2005); they also have a close relationship with blood vessels (Mirzadeh et al., 2008). Precursor cells reactive to GFAP have been identified in the SVZ of the human brain (Roy et al., 2000; Gibbons and Dragunow, 2010), and some are in direct contact with the cerebrospinal fluid (CSF) (Sanai et al., 2004, 2011; Quinones-Hinojosa et al., 2006).

Vitamin C, which is present in high concentrations in the CSF (Spector and Lorenzo, 1974; Kratzting et al., 1985), may be important in postnatal neural differentiation. An important role for vitamin C in embryonic cerebral development and in the differentiation of dopaminergic and serotonergic neurons has been described (Lee et al., 2000; Yan et al., 2001). Embryonic precursors supplemented with vitamin C show an increase in neural and glial markers (Lee et al., 2003). Recently, Esteban et al. (2010) found that vitamin C favored the generation of induced pluripotent stem cells (iPS) (Esteban et al., 2010). Furthermore, cells grown *in vitro* in the presence of vitamin C expressed two histone demethylases, Jhdmla and Jhdmlb (Wang et al., 2011), which are required for iPS cell production. Together, these results suggest that vitamin C is able to positively regulate stem cell generation and proliferation.

Subunit Interactions during Cooperative Opening of Voltage-Gated Proton Channels

Feng Qiu,¹ Santiago Rebolledo,¹ Carlos Gonzalez,² and H. Peter Larsson^{1,*}

¹Department of Physiology and Biophysics, University of Miami, 1600 NW 10th Avenue, RMSB 5123, Miami, FL 33136, USA

²Centro Interdisciplinario de Neurociencias de Valparaíso, Universidad de Valparaíso, Valparaíso 2366103, Chile

*Correspondence: plarsson@med.miami.edu

<http://dx.doi.org/10.1016/j.neuron.2012.12.021>

SUMMARY

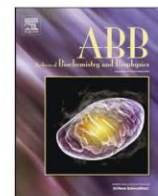
Voltage-gated proton (Hv1) channels are dimers, where each subunit has a separate permeation pathway. However, opening of the two pathways is highly cooperative. It is unclear how Hv1 channels open their permeation pathways, because Hv1 channels lack a classic pore domain. Using voltage-clamp fluorometry, we here detect two conformational changes reported by a fluorophore attached to the voltage sensor S4 in Hv1 channels. The first is voltage dependent and precedes channel opening, with properties consistent with reporting on independent S4 charge movements in the two subunits. The second is less voltage dependent and closely correlates with channel opening. Mutations that reduce dimerization or alter the intersubunit interface affect both the second conformational change and channel opening. These observations suggest that, following an initial S4 charge movement in the two subunits, there is a second, cooperative conformational change, involving interactions between subunits, that opens both pathways in Hv1 channels.

INTRODUCTION

Voltage-gated proton (Hv1) channels were first discovered in snail neurons (Thomas and Meech, 1982). Subsequently, Hv1 have been found in a variety of cell types and have been implicated in a wide variety of biological functions, such as the respiratory burst in phagocytic cells and the capacitation of the sperm (DeCoursey, 2010; El Chemaly et al., 2010; Iovannisci et al., 2010; Lishko et al., 2010; Morgan et al., 2009). In neurons, Hv1 channels are thought to play a role in the control of intracellular pH by extruding protons in response to excess intracellular acidification (DeCoursey, 2003) and have been recently implicated in enhancement of brain damage in ischemic stroke (Wu et al., 2012). Hv1 channels belong to the superfamily of voltage-gated cation channels (Ramsey et al., 2006; Sasaki et al., 2006). However, in contrast to other members of this superfamily, such as Kv and Nav channels, Hv1 channel subunits have just four transmembrane segments (S1–S4; Figure 1A), which correspond to the S1–S4 voltage sensor domain in Kv channels (Ramsey et al., 2006; Sasaki et al., 2006). Hv1 channels lack

the characteristic pore domain that is made up of transmembrane segments S5 and S6 in other voltage-gated cation channels (Jiang et al., 2003; Long et al., 2005). In other voltage-gated cation channels, the intracellular end of S6 constitutes the activation gate that opens and closes the pore (Liu et al., 1997). The lack of S6 in Hv1 channels makes it unclear what constitutes the gate in Hv1 channels. We here characterize conformational changes that are coupled to opening and closing (i.e., gating) of Hv1 channels, in order to elucidate how Hv1 channels are gated.

Hv1 channel subunits oligomerize as dimers (Koch et al., 2008; Lee et al., 2008; Tombola et al., 2008), not tetramers as other members of the superfamily of voltage-gated cation channels (Jiang et al., 2003; Long et al., 2005). This dimerization is most likely through the formation of a coiled-coil structure of the two C-termini from the two subunits in the dimer (Fujiwara et al., 2012; Koch et al., 2008; Lee et al., 2008; Tombola et al., 2008). Each subunit of the dimer has its own permeation pathway (Koch et al., 2008; Tombola et al., 2008), but there is a strong cooperativity between the two subunits during channel activation (Fujiwara et al., 2012; Gonzalez et al., 2010; Tombola et al., 2010). Using voltage-clamp fluorometry and cysteine accessibility to thiol reagents on Hv1 channels, we previously showed that the fourth transmembrane segment S4 with its three positively charged residues moves outward in response to depolarizations, as if S4 functions as the voltage sensor (Gonzalez et al., 2010). Cysteines introduced in the external end of S4 are exposed to the extracellular solution at depolarized membrane potentials, but not at hyperpolarized potentials. Conversely, cysteines introduced in the internal end of S4 are exposed to the intracellular solution at hyperpolarized membrane potentials, but not at depolarized potentials (Gonzalez et al., 2010). In addition, in response to depolarizations, a fluorophore attached close to S4 displays a fluorescence decrease that precedes the proton current (Gonzalez et al., 2010), suggesting that the fluorescence decrease reports on S4 charge movement preceding channel opening. The time course of the fluorescence decrease raised to a power of two overlaps well the time course of the proton currents for moderate depolarizations (Gonzalez et al., 2010). Using a Hodgkin-Huxley type analysis, we previously interpreted the time courses of the fluorescence and currents to denote that S4s in both subunits need to activate before either of the two proton permeation pathways in the two subunits is activated. This suggests that there is a high degree of cooperativity between the two subunits in the Hv1 dimer during channel opening. Using linked Hv1 subunits with different



Penicillium purpurogenum produces two GH family 43 enzymes with β -xylosidase activity, one monofunctional and the other bifunctional: Biochemical and structural analyses explain the difference



María Cristina Ravanal^a, Melissa Alegría-Arcos^b, Fernando Danilo Gonzalez-Nilo^{b,*}, Jaime Eyzaguirre^{a,*}

^a Universidad Andres Bello, Facultad de Ciencias Biológicas, Santiago, Chile

^b Universidad Andres Bello, Center for Bioinformatics and Integrative Biology, Facultad de Ciencias Biológicas, Santiago, Chile

ARTICLE INFO

Article history:

Received 28 August 2013

and in revised form 8 October 2013

Available online 31 October 2013

Keywords:

β -D-Xylosidase

Penicillium purpurogenum

Heterologous expression

Glycosyl hydrolase family 43

Homology modeling

Molecular docking simulations

ABSTRACT

β -Xylosidases participate in xylan biodegradation, liberating xylose from the non-reducing end of xylooligosaccharides. The fungus *Penicillium purpurogenum* secretes two enzymes with β -D-xylosidase activity belonging to family 43 of the glycosyl hydrolases. One of these enzymes, arabinofuranosidase 3 (ABF3), is a bifunctional α -L-arabinofuranosidase/xylobiohydrolase active on p-nitrophenyl- α -L-arabinofuranoside (pNPArA) and p-nitrophenyl- β -D-xylopyranoside (pNPXyl) with a K_M of 0.65 and 12 mM, respectively. The other, β -D-xylosidase 1 (XYL1), is only active on pNPXyl with a K_M of 0.55 mM. The *xy1* gene was expressed in *Pichia pastoris*, purified and characterized. The properties of both enzymes were compared in order to explain their difference in substrate specificity. Structural models for each protein were built using homology modeling tools. Molecular docking simulations were used to analyze the interactions defining the affinity of the proteins to both ligands. The structural analysis shows that active complexes (ABF3-pNPXyl, ABF3-pNPArA and XYL1-pNPXyl) possess specific interactions between substrates and catalytic residues, which are absent in the inactive complex (XYL1-pNPArA), while other interactions with non-catalytic residues are found in all complexes. pNPArA is a competitive inhibitor for XYL1 ($K_i = 2.5$ mM), confirming that pNPArA does bind to the active site but not to the catalytic residues.

© 2013 Elsevier Inc. All rights reserved.

Introduction

Lignocellulose is a major component of plants and it represents the main source of renewable organic matter. There is considerable interest in the exploitation of lignocellulosic materials as a source of feed, fuels and chemical feedstocks. Both the abundance and renewable nature of lignocellulosic materials have provided input for intensive research over recent years. It can be converted to value-added products through saccharification by lignocellulolytic enzymes [1]. Lignocellulose is composed of lignin, pectin, cellulose and hemicelluloses. Xylan is the main hemicellulose of annual plants and hardwoods, and it is composed of a linear chain of xylopyranose residues joined by β (1 \rightarrow 4) glycosidic linkages. The main chain is substituted by a variety of compounds, such as arabinofuranose, methyl glucuronate and acetate, and the arabinoses may be linked to hydroxycinnamic acids [2].

The biodegradation of xylan is a complex process. The main chain is hydrolyzed by the action of endoxylanases (EC 3.2.1.8),

which liberate xylooligosaccharides of different length and are eventually hydrolyzed to xylose by β -xylosidases (EC 3.2.1.37). These enzymes are produced by fungi and bacteria and are mainly extracellular [3]. β -Xylosidases are active against artificial substrates such as p-nitrophenyl glycosides; most of them are very specific for p-nitrophenyl- β -D-xylopyranoside (pNPXyl)¹ [4–6]. Others are able to cleave p-nitrophenyl- α -L-arabinofuranoside (pNPArA) [7–10], p-nitrophenyl- β -D-galactopyranoside [9] or p-nitrophenyl- α -D-glucopyranoside [5,11,12]. β -Xylosidases are grouped in families, according to their amino acid sequence similarities, in the Carbohydrate Active Enzymes database (CAZy; <http://www.cazy.org>); they are classified in eight families of glycoside hydrolases (GH): 3, 30, 39, 43, 52, 54, 116 and 120 [13–16]. Filamentous fungal β -xylosidases have been described only for families 3, 43 and 54 [5,12,17]. Members of GH families 3 and 54 operate with retention of the anomeric configuration, while family GH43 contains inverting glycoside hydrolases. These last enzymes possess a proton donor acting as general acid and a nucleophile as general base; in

* Corresponding authors. Fax: +56 2 6980414.

E-mail addresses: fernando.gonzalez@unab.cl (F.D. Gonzalez-Nilo), jeyzaguirre@unab.cl (J. Eyzaguirre).

¹ Abbreviations used: ABF3, arabinofuranosidase 3; XYL1, β -xylosidase; pNPXyl, p-nitrophenyl- β -D-xylopyranoside; pNPArA, p-nitrophenyl- α -L-arabinofuranoside; GH, glycoside hydrolases.



The ATP required for potentiation of skeletal muscle contraction is released via pannexin hemichannels[☆]



Manuel A. Riquelme^a, Luis A. Cea^{a,f}, José L. Vega^{a,b,c}, Mauricio P. Boric^a, Hannah Monyer^{b,c}, Michael V.L. Bennett^d, Marina Frank^e, Klaus Willecke^e, Juan C. Sáez^{a,f,*}

^aDepartamento de Fisiología, Pontificia Universidad Católica de Chile, Santiago 8, Chile

^bLaboratorio de Fisiología Experimental (EPhyL), Instituto Antofagasta, Universidad de Antofagasta, Antofagasta, Chile

^cDepartment of Clinical Neurobiology, University of Heidelberg, 6012 Heidelberg, Germany

^dDepartment of Neuroscience, Albert Einstein College of Medicine, Bronx, NY 10461, USA

^eLife and Medical Sciences Institute, Molecular Genetics, University of Bonn, 53115 Bonn, Germany

^fInstituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Valparaíso, Chile

ARTICLE INFO

Article history:

Received 8 January 2013

Received in revised form

7 March 2013

Accepted 11 March 2013

Keywords:

Pannexin

Purinergic receptors

Contractile force

ABSTRACT

During repetitive stimulation of skeletal muscle, extracellular ATP levels raise, activating purinergic receptors, increasing Ca^{2+} influx, and enhancing contractile force, a response called potentiation. We found that ATP appears to be released through pannexin1 hemichannels (Pannx1 HCs). Immunocytochemical analyses and function were consistent with pannexin1 localization to T-tubules intercalated with dihydropyridine and ryanodine receptors in slow (*soleus*) and fast (*extensor digitorum longus*, *EDL*) muscles. Isolated myofibers took up ethidium (Etd^+) and released small molecules (as ATP) during electrical stimulation. Consistent with two glucose uptake pathways, induced uptake of 2-NBDG, a fluorescent glucose derivative, was decreased by inhibition of HCs or glucose transporter (GLUT4), and blocked by dual blockade. Adult skeletal muscles apparently do not express connexins, making it unlikely that connexin hemichannels contribute to the uptake and release of small molecules. ATP release, Etd^+ uptake, and potentiation induced by repetitive electrical stimulation were blocked by HC blockers and did not occur in muscles of pannexin1 knockout mice. MRS2179, a $\text{P2Y}_1\text{R}$ blocker, prevented potentiation in *EDL*, but not *soleus* muscles, suggesting that in fast muscles ATP activates P2Y_1 but not P2X receptors. Phosphorylation on Ser and Thr residues of pannexin1 was increased during potentiation, possibly mediating HC opening. Opening of Pannx1 HCs during repetitive activation allows efflux of ATP, influx of glucose and possibly Ca^{2+} too, which are required for potentiation of contraction.

This article is part of the Special Issue Section entitled 'Current Pharmacology of Gap Junction Channels and Hemichannels'.

© 2013 Elsevier Ltd. All rights reserved.

1. Introduction

A single twitch of a vertebrate skeletal muscle can occur in the absence of extracellular Ca^{2+} , and the required rise in intracellular free Ca^{2+} , $[\text{Ca}^{2+}]_i$, is released from intracellular stores (Araya et al., 2003). Repetitive twitches lead to potentiation of the contraction through accumulation of free Ca^{2+} in the cytoplasm (Sandona et al., 2005; Zhi et al., 2005). ATP signaling is involved in both, fast and slow muscle potentiation. In slow skeletal muscles, potentiation depends on activation of purinergic P2X_4 receptors present in the T-tubule membrane and entry of extracellular Ca^{2+} (Sandona et al., 2005). However, P2X_4 receptors are not found in fast skeletal muscle (Sandona et al., 2005), and potentiation in these muscles occurs without of extracellular Ca^{2+} (Louboutin

Abbreviations: Pannx1 HCs, pannexin1 hemichannels; Etd^+ , ethidium.

[☆] The data of this work was presented by Dr. Manuel A. Riquelme as partial fulfillment of the requirements to obtain the degree of Ph.D. in Physiological Sciences at the Pontificia Universidad Católica de Chile. This work was partially supported by FONDECYT grants (1111033 to J.C.S. and 3120006 to J.L.V.) Anillo ACT71 (to J.C.S.) and a CONICYT Doctoral thesis grant (AT-23070155 to M.A.R.), an NIH grant (NS045287 to M.V.L.B.) and DAAD-CONICYT grant (to J.C.S. and K.W.).

* Corresponding author. Departamento de Fisiología, Pontificia Universidad Católica de Chile, casilla 114-D, Santiago 8, Chile. Tel.: +56 2 6862862; fax: +56 2 2225515.

E-mail addresses: jsaez@bio.puc.cl, jcsaez@gmail.com (J.C. Sáez).

Disruption in Connexin-Based Communication Is Associated with Intracellular Ca^{2+} Signal Alterations in Astrocytes from Niemann-Pick Type C Mice

Pablo J. Sáez^{1*}, Juan A. Orellana², Natalia Vega-Riveros¹, Vania A. Figueroa^{1,3}, Diego E. Hernández¹, Juan F. Castro⁴, Andrés D. Klein⁴, Jean X. Jiang⁵, Silvana Zanlungo^{4,6*}, Juan C. Sáez^{1,3*}

1 Departamento de Fisiología, Pontificia Universidad Católica de Chile, Santiago, Chile, **2** Departamento de Neurología, Facultad de Medicina, Pontificia Universidad Católica de Chile, Santiago, Chile, **3** Instituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Valparaíso, Chile, **4** Departamento de Gastroenterología, Facultad de Medicina, Pontificia Universidad Católica de Chile, Santiago, Chile, **5** Department of Biochemistry, University of Texas Health Science Center, San Antonio, Texas, United States of America, **6** FONDAF Center for Genome Regulation (CGR), Santiago, Chile

Abstract

Reduced astrocytic gap junctional communication and enhanced hemichannel activity were recently shown to increase astroglial and neuronal vulnerability to neuroinflammation. Moreover, increasing evidence suggests that neuroinflammation plays a pivotal role in the development of Niemann-Pick type C (NPC) disease, an autosomal lethal neurodegenerative disorder that is mainly caused by mutations in the *NPC1* gene. Therefore, we investigated whether the lack of NPC1 expression in murine astrocytes affects the functional state of gap junction channels and hemichannels. Cultured cortical astrocytes of NPC1 knock-out mice (*Npc1*^{−/−}) showed reduced intercellular communication via gap junctions and increased hemichannel activity. Similarly, astrocytes of newborn *Npc1*^{−/−} hippocampal slices presented high hemichannel activity, which was completely abrogated by connexin 43 hemichannel blockers and was resistant to inhibitors of pannexin 1 hemichannels. *Npc1*^{−/−} astrocytes also showed more intracellular Ca^{2+} signal oscillations mediated by functional connexin 43 hemichannels and P2Y₁ receptors. Therefore, *Npc1*^{−/−} astrocytes present features of connexin based channels compatible with those of reactive astrocytes and hemichannels might be a novel therapeutic target to reduce neuroinflammation in NPC disease.

Citation: Sáez PJ, Orellana JA, Vega-Riveros N, Figueroa VA, Hernández DE, et al. (2013) Disruption in Connexin-Based Communication Is Associated with Intracellular Ca^{2+} Signal Alterations in Astrocytes from Niemann-Pick Type C Mice. PLoS ONE 8(8): e71361. doi:10.1371/journal.pone.0071361

Editor: Eliana Scemes, Albert Einstein College of Medicine, United States of America

Received: April 3, 2013; **Accepted:** June 27, 2013; **Published:** August 15, 2013

Copyright: © 2013 Sáez et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was partially supported by the CONICYT 24100062 (to PJS); Welch Foundation grant AQ-1507 and National Institutes of Health (NIH) EY012085 (to JXJ); FONDECYT 1110310 and Fondo Nacional de Desarrollo de Áreas Prioritarias (FONDAF) 15090007, Center for Genome Regulation (CGR) (to SZ); FONDECYT 1111033, FONDEF D071086 and ANILLO ACT-71 (to JCS). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: pjsaez@uc.cl (PJS); jsaez@bio.puc.cl (JCS); silvana@med.puc.cl, (SZ)

Introduction

Niemann-Pick type C (NPC) disease is an autosomal recessive neurodegenerative disorder that is caused by mutations in the *NPC1* or *NPC2* genes [1]. Most cases of NPC disease are caused by mutations in the *NPC1* gene that yields a dysfunctional protein [1,2]. NPC1 and NPC2 proteins are required for the trafficking of cholesterol; hence, a loss of function in these proteins results in the intracellular accumulation of free cholesterol and other lipids in late endosomes/lysosomes [3]. Progressive neurodegeneration, hepatosplenomegaly, and dysfunction of other organs are observed in patients affected with NPC disease [2]. These symptoms are also observed in a murine model of NPC disease [2,4].

Npc1^{−/−} mice show hippocampal and cortical neuronal dysfunction [5–7], apoptosis of Purkinje neurons of the cerebellum and neuronal death in different brain regions [8–10]. Astrocytes express NPC1; and in the *Npc1*^{−/−} mouse brain, *Npc1*^{−/−} astrocytes exhibit morphological changes and become activated [11,12]. The global neuronal deletion of NPC1, but not astrocyte-specific NPC1 deficiency, leads to the complete development of NPC neuropathology [13], which suggests that neuronal NPC1

deficiency is sufficient to mediate neurodegeneration. However, rescuing NPC1 expression in astrocytes delays neuronal loss and prolongs the life span in *Npc1*^{−/−} mice [14], suggesting that astrocytes may play an important role in the neuroinflammatory state of NPC disease. Neuroinflammation is present in *Npc1*^{−/−} mouse brain at an early post-natal age and is characterized by an enhanced number of microglia, increased levels of interleukin-1 β and the presence of activated astrocytes [15]. Because astrocytes form extensive communicating networks [16], it is conceivable that NPC-induced neurodegeneration could depend on intercellular signaling and coordination among astrocytes. Such intercellular communication between astrocytes is partially attained by sharing cytoplasmic content through gap junction channels (GJCs); these intercellular channels allow direct but selective cytoplasmic communication between contacting cells, thereby promoting the exchange of metabolites and second messengers [17]. Each GJC is formed by the serial docking of two hemichannels (HCs), each contributed by one of two adjacent cells. HCs are composed of six protein subunits termed connexins (Cx) [18]. Under defined conditions HCs mediate the uptake or release of ions and small molecules such as Ca^{2+} and ATP, respectively [19]. *In vivo*,

Research Article

ATP Is Required and Advances Cytokine-Induced Gap Junction Formation in Microglia In Vitro

Pablo J. Sáez,^{1,2} Kenji F. Shoji,^{1,2} Mauricio A. Retamal,³ Paloma A. Harcha,^{1,2} Gigliola Ramírez,⁴ Jean X. Jiang,⁵ Rommy von Bernhardt,⁴ and Juan C. Sáez^{1,2}

¹ Departamento de Fisiología, Pontificia Universidad Católica de Chile, Alameda 340, 6513677 Santiago, Chile

² Instituto Milenio, Centro Interdisciplinario de Neurociencias de Valparaíso, Pasaje Harrington 287, Playa Ancha, 2360103 Valparaíso, Chile

³ Departamento de Fisiología, Facultad de Medicina, Clínica Alemana-Universidad del Desarrollo, Las Condes 12438, 7710162 Santiago, Chile

⁴ Departamento de Neurología, Escuela de Medicina, Pontificia Universidad Católica de Chile, Marcoleta 392, 8330024 Santiago, Chile

⁵ Department of Biochemistry, University of Texas Health Science Center, 7703 Floyd Curl Drive, San Antonio, TX 78229, USA

Correspondence should be addressed to Pablo J. Sáez; pjsaez@uc.cl

Received 4 February 2013; Revised 21 March 2013; Accepted 22 March 2013

Academic Editor: Diego Gomez-Nicola

Copyright © 2013 Pablo J. Sáez et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Microglia are the immune cells in the central nervous system. After injury microglia release bioactive molecules, including cytokines and ATP, which modify the functional state of hemichannels (HCs) and gap junction channels (GJCs), affecting the intercellular communication via extracellular and intracellular compartments, respectively. Here, we studied the role of extracellular ATP and several cytokines as modulators of the functional state of microglial HCs and GJCs using dye uptake and dye coupling techniques, respectively. In microglia and the microglia cell line EOC20, ATP advanced the TNF- α /IFN- γ -induced dye coupling, probably through the induction of IL-1 β release. Moreover, TNF- α /IFN- γ , but not TNF- α plus ATP, increased dye uptake in EOC20 cells. Blockade of Cx43 and Panx1 HCs prevented dye coupling induced by TNF- α /IFN- γ , but not TNF- α plus ATP. In addition, IL-6 prevented the induction of dye coupling and HC activity induced by TNF- α /IFN- γ in EOC20 cells. Our data support the notion that extracellular ATP affects the cellular communication between microglia through autocrine and paracrine mechanisms, which might affect the timing of immune response under neuroinflammatory conditions.

1. Introduction

Microglia are the major immune effectors in the central nervous system (CNS). Under resting conditions, surveillance microglia have a ramified morphology and monitor their local microenvironment [1, 2]. However, microglia can rapidly become activated in response to diverse stimuli and danger signals, such as ATP or bacterial lipopolysaccharide (LPS) [1–3]. Consistently, microglia are activated in neuroinflammatory conditions and are a common hallmark in many neurodegenerative diseases [1, 2, 4].

Microglial cell activation includes morphological changes, proliferation, recruitment to the site of injury, and expression of specific proteins including MHC II molecules and cell

adhesion molecules [1, 2]. Activated microglia also release cytokines, including TNF- α , IL-1 β , IL-6, IFN- γ , and other soluble molecules, such as glutamate and ATP [5–9]. Many of these pro-inflammatory molecules act in an autocrine manner and show synergism, increasing the activation of microglia [10–12].

Many studies have focused on ATP release mechanisms and the subsequent receptors activation at the CNS, because they promote the release of other pro-inflammatory molecules, such as TNF- α and IL-1 β [13]. These cytokines mediate cell communication and Ca²⁺ signaling among microglia, as well as among microglia and astrocytes [14–16]. Microglia sense extracellular ATP through P2Y and P2X receptors [1]. Under control conditions, microglia express

4 Gap Junctions in Antigen-Presenting Cells

Pablo J. Sáez, Kenji F. Shoji, and Juan Carlos Sáez

CONTENTS

4.1	Introduction	61
4.2	Expression of Connexins by Dendritic Cells and T Cells	64
4.2.1	Connexins in Dendritic Cells	64
4.2.2	Connexins in T Cells	67
4.3	Why Cell Contacts between Dendritic Cells and T-Cells Might Present Gap Junctions?	69
4.3.1	Importance of Cell–Cell Contacts between Dendritic Cells	69
4.3.2	Gap Junctions between Dendritic Cells	70
4.4	Are Gap Junctions Sufficient to Explain the Need for Direct Physical Contact between Dendritic Cells So to Be Good Antigen-Presenting Cells?....	72
4.4.1	Gap Junctions between Dendritic Cells and T Cells as Part of Immune Synapse	73
4.4.2	Intercellular Communication Pathways during Antigen Presentation and Cross-Presentation	73
4.5	Connexin Gap Junctions in Other Antigen-Presenting Cells	75
4.6	Concluding Remarks	80
	Acknowledgments.....	81
	References.....	81

4.1 INTRODUCTION

Dendritic cells (DCs) constitute a heterogeneous cell population that emerges in the bone marrow (BM) from a macrophage and DC precursor, generating the following: (1) a common DC precursor and (2) a monocyte precursor (Liu and Nussenzweig 2010). In addition, DCs can also emerge from monocytes under inflammatory conditions (Shortman and Naik 2007). Two major categories of DCs have been established as follows: (1) conventional (cDCs) and (2) inflammatory DCs. DC differentiation is reached by the former during resting steady-state conditions, while the latter do so during inflammatory conditions (Merad and Manz 2009). In addition, cDCs can be classified into several subtypes that share the ability to pick up, process, and present antigens to T cells. DC subtypes can be recognized through the expression of different cell surface markers, pattern cytokine secretions, migration pathways, locations, and functions (Shortman and Naik 2007). Although plasmacytoid DCs (pDCs) are

In Silico Analysis of Putative Paralytic Shellfish Poisoning Toxins Export Proteins in Cyanobacteria

Katia Soto-Liebe¹, Xaviera A. López-Cortés², Juan José Fuentes-Valdes¹, Karina Stucken³, Fernando Gonzalez-Nilo⁴, Mónica Vásquez^{1*}

1 Pontificia Universidad Católica de Chile, Santiago, Chile, **2** Nanobiotechnology Division at University of Talca, Fraunhofer Chile Research Foundation - Center for Systems Biotechnology, Talca, Chile, **3** Institute of Molecular Evolution Heinrich-Heine, Universität Düsseldorf, Düsseldorf, Germany, **4** Universidad Andres Bello, Center for Bioinformatics and Integrative Biology, Santiago, Chile

Abstract

Paralytic shellfish poisoning toxins (PSTs) are a family of more than 30 natural alkaloids synthesized by dinoflagellates and cyanobacteria whose toxicity in animals is mediated by voltage-gated Na⁺ channel blocking. The export of PST analogues may be through SxtF and SxtM, two putative MATE (multidrug and toxic compound extrusion) family transporters encoded in PSTs biosynthetic gene cluster (*sxt*). *sxtM* is present in every *sxt* cluster analyzed; however, *sxtF* is only present in the *Cylindrospermopsis-Raphidiopsis* clade. These transporters are energetically coupled with an electrochemical gradient of proton (H⁺) or sodium (Na⁺) ions across membranes. Because the functional role of PSTs remains unknown and methods for genetic manipulation in PST-producing organisms have not yet been developed, protein structure analyses will allow us to understand their function. By analyzing the *sxt* cluster of eight PST-producing cyanobacteria, we found no correlation between the presence of *sxtF* or *sxtM* and a specific PSTs profile. Phylogenetic analyses of SxtF/M showed a high conservation of SxtF in the *Cylindrospermopsis-Raphidiopsis* clade, suggesting conserved substrate affinity. Two domains involved in Na⁺ and drug recognition from NorM proteins (MATE family) of *Vibrio parahaemolyticus* and *V. cholerae* are present in SxtF/M. The Na⁺ recognition domain was conserved in both SxtF/M, indicating that Na⁺ can maintain the role as a cation anti-transporter. Consensus motifs for toxin binding differed between SxtF and SxtM implying differential substrate binding. Through protein modeling and docking analysis, we found that there is no marked affinity between the recognition domain and a specific PST analogue. This agrees with our previous results of PST export in *R. brookii* D9, where we observed that the response to Na⁺ incubation was similar to different analogues. These results reassert the hypothesis regarding the involvement of Na⁺ in toxin export, as well as the motifs L³⁹⁸XGLQD⁴⁰³ (SxtM) and L³⁹⁰VGLRD³⁹⁵ (SxtF) in toxin recognition.

Citation: Soto-Liebe K, López-Cortés XA, Fuentes-Valdes JJ, Stucken K, Gonzalez-Nilo F, et al. (2013) In Silico Analysis of Putative Paralytic Shellfish Poisoning Toxins Export Proteins in Cyanobacteria. PLoS ONE 8(2): e55664. doi:10.1371/journal.pone.0055664

Editor: Tamir Tuller, Tel Aviv University, Israel

Received: September 21, 2012; **Accepted:** December 29, 2012; **Published:** February 15, 2013

Copyright: © 2013 Soto-Liebe et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: The first author KSL is a Comisión Nacional de Investigación Científica y Tecnológica (CONICYT) PhD fellow. This work was financed and supported by Fondo Nacional de Desarrollo Científico y Tecnológico (FONDECYT) Grant 1050433 and 1080075. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: mvasquez@bio.puc.cl

Introduction

Cyanobacteria are a biochemically and morphologically diverse clade of photosynthetic bacteria, with major environmental and economic roles. They are main primary producers in marine and freshwater ecosystems, key biocatalysts in the N₂ cycle [1] and some are capable of producing toxins. Toxin production can present waterborne health hazards for humans and animals. This has become of particular interest lately, due to the proliferation and dominance of harmful blooms of cyanobacteria; a result of the increase of eutrophication and climate change among others [2]. In addition, toxic strains of *Anabaena*, *Anabaenopsis*, *Microcystis* and *Nodularia* are salt tolerant, and increasing halotolerant HABs have been observed more frequently [2].

Major cyanobacterial toxins include microcystins, cylindrospermopsins, nodularin, anatoxins and saxitoxins (STX) [3]. STX and its analogues (more than 30) have been detected in filter-feeding bivalve mollusks. In humans, these toxins cause paralytic shellfish poisoning (PSP). PSP-toxins (PSTs) produce symptoms that vary

from a slight tingling sensation or numbness around the lips to a fatal respiratory paralysis. The long-established molecular target of PSTs is the voltage-gated sodium channel in nerve and muscle cells, to which PSTs bind with analogue-dependent affinity, blocking Na⁺ channels in nanomolar concentrations [4]. STX has also been shown to target voltage-gated potassium [5] and calcium [6] ion channels.

STX is a trialkyltetrahydropurine molecule (Figure 1), with two pK_a's of 8.22 and 11.28 in aqueous solution, which belong to the 7,8,9 and 1,2,3 guanidinium groups, respectively. At physiological pH, the 1,2,3-guanidine carry a positive charge, whereas the 7,8,9-guanidine group is partially deprotonated. STX and its analogs can be structurally classified into several classes such as non-sulfated (STX, neoSTX), mono-sulfated (gonyautoxins-GTX1 6), di-sulfated (C1 4), and decarbamoylated, each with varying levels of toxicity. Of the decarbamoyl variants of these analogs, there are decarbamoyl-saxitoxins (dcSTX, dcneoSTX) and decarbamoyl-gonyautoxins (dcGTXs 1 4).

Emerging Role of Calcium-Activated Potassium Channel in the Regulation of Cell Viability Following Potassium Ions Challenge in HEK293 Cells and Pharmacological Modulation

Domenico Tricarico^{1,2*}, Antonietta Mele^{1,2}, Sara Calzolaro^{1,2}, Gianluigi Cannone^{1,2}, Giulia Maria Camerino^{1,2}, Maria Maddalena Dinardo^{1,2}, Ramon Latorre^{1,2}, Diana Conte Camerino^{1,2}

1 Department of Pharmacy-Drug-Science, University of Bari, Bari, Italy, **2** Centro de Neurociencias de Valparaíso, Facultad de Ciencias, Universidad de Valparaíso, Valparaíso, Chile

Abstract

Emerging evidences suggest that Ca^{2+} -activated- K^{+} -(BK) channel is involved in the regulation of cell viability. The changes of the cell viability observed under hyperkalemia (15 mEq/L) or hypokalemia (0.55 mEq/L) conditions were investigated in HEK293 cells expressing the hsl α subunit (hsl α -HEK293) in the presence or absence of BK channel modulators. The BK channel openers (10^{-11} - 10^{-3} M) were: acetazolamide (ACTZ), Dichlorophenamide (DCP), methazolamide (MTZ), bendroflumethiazide (BFT), ethoxzolamide (ETX), hydrochlorothiazide (HCT), quercetin (QUERC), resveratrol (RESV) and NS1619; and the BK channel blockers (2×10^{-7} M- 5×10^{-3} M) were: tetraethylammonium (TEA), iberiotoxin (IbTx) and charybdotoxin (ChTX). Experiments on cell viability and channel currents were performed using cell counting kit-8 and patch-clamp techniques, respectively. hsl α whole-cell current was potentiated by BK channel openers with different potency and efficacy in hsl α -HEK293. The efficacy ranking of the openers at -60 mV (Vm) was BFT > ACTZ > DCP > RESV > ETX > NS1619 > MTZ > QUERC; HCT was not effective. Cell viability after 24 h of incubation under hyperkalemia was enhanced by 82±6% and 33±7% in hsl α -HEK293 cells and HEK293 cells, respectively. IbTx, ChTX and TEA enhanced cell viability in hsl α -HEK293. BK openers prevented the enhancement of the cell viability induced by hyperkalemia or IbTx in hsl α -HEK293 showing an efficacy which was comparable with that observed as BK openers. BK channel modulators failed to affect cell currents and viability under hyperkalemia conditions in the absence of hsl α subunit. In contrast, under hypokalemia cell viability was reduced by -22±4% and -23±6% in hsl α -HEK293 and HEK293 cells, respectively; the BK channel modulators failed to affect this parameter in these cells. In conclusion, BK channel regulates cell viability under hyperkalemia but not hypokalemia conditions. BFT and ACTZ were the most potent drugs either in activating the BK current and in preventing the cell proliferation induced by hyperkalemia. These findings may have relevance in disorders associated with abnormal K^{+} ion homeostasis including periodic paralysis and myotonia.

Citation: Tricarico D, Mele A, Calzolaro S, Cannone G, Camerino GM, et al. (2013) Emerging Role of Calcium-Activated Potassium Channel in the Regulation of Cell Viability Following Potassium Ions Challenge in HEK293 Cells and Pharmacological Modulation. PLoS ONE 8(7): e69551. doi:10.1371/journal.pone.0069551

Editor: Stuart E Dryer, University of Houston, United States of America

Received: May 2, 2013; **Accepted:** June 10, 2013; **Published:** July 16, 2013

Copyright: © 2013 Tricarico et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work is supported by Telethon Grants. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

* E-mail: domenico.tricarico@uniba.it

Introduction

Potassium ions regulate inflammation, oxidative stress, vascular biology and blood pressure, the excitability of the cells, exerting beneficial effects on different tissues [1–3]. Abnormalities in the serum potassium ion levels are associated with acquired and congenital diseases affecting several apparatus including skeletal muscle [4].

Severe hyperkalemia characterizes the hyperkalemic renal tubular Acidosis (type IV), mineralocorticoid deficiency (hypoaldosteronism states) as well as tumor lysis syndrome, rhabdomyolysis, marked leucocytosis and thrombocytosis, trauma and burns [5]. Disease progression and increased hearth mortality are observed in chronic kidney disease under hypokalemia or hyperkalemia conditions and these effects are gender and race dependent [6]. Severe nephropathy with renal

Review Article

Role of Gap Junctions and Hemichannels in Parasitic Infections

José Luis Vega,^{1,2} Mario Subiabre,² Felipe Figueroa,³ Kurt Alex Schalper,⁴ Luis Osorio,⁵ Jorge González,⁵ and Juan Carlos Sáez¹

¹ Departamento de Fisiología, Pontificia Universidad Católica de Chile, 8330025 Santiago, Chile

² Laboratorio de Fisiología Experimental (EPhyL), Instituto Antofagasta (IA), Universidad de Antofagasta, 1270300 Antofagasta, Chile

³ Inmunología, Departamento de Tecnología Médica, Universidad de Antofagasta, 1270300 Antofagasta, Chile

⁴ Department of Pathology, Yale School of Medicine, New Haven, CT 06520-8023, USA

⁵ Unidad de Parasitología Molecular, Facultad Ciencias de la Salud, Universidad de Antofagasta, 1270300 Antofagasta, Chile

Correspondence should be addressed to José Luis Vega; juvega@uc.cl

Received 21 May 2013; Revised 7 August 2013; Accepted 26 August 2013

Academic Editor: Christophe Duranton

Copyright © 2013 José Luis Vega et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

In vertebrates, connexins (Cx) and pannexins (Panx) are proteins that form gap junction channels and/or hemichannels located at cell-cell interfaces and cell surface, respectively. Similar channel types are formed by innexins in invertebrate cells. These channels serve as pathways for cellular communication that coordinate diverse physiologic processes. However, it is known that many acquired and inherited diseases deregulate Cx and/or Panx channels, condition that frequently worsens the pathological state of vertebrates. Recent evidences suggest that Cx and/or Panx hemichannels play a relevant role in bacterial and viral infections. Nonetheless, little is known about the role of Cx- and Panx-based channels in parasitic infections of vertebrates. In this review, available data on changes in Cx and gap junction channel changes induced by parasitic infections are summarized. Additionally, we describe recent findings that suggest possible roles of hemichannels in parasitic infections. Finally, the possibility of new therapeutic designs based on hemichannel blockers is presented.

1. Introduction

Members of gap junction (GJ) family proteins form intercellular communication channels, which connect the cytoplasm of neighboring cells and hemichannels, which connect the intra- and extracellular milieu [1]. Both intercellular channels and hemichannels participate in physiologic and pathologic processes including electrical conduction [2], inflammation [3], immune system activation [4], tissue repair/remodeling [5], and response to bacterial [6, 7] and viral infections [8]. However, little is known about the role of GJ channels in parasite infection and studies on the possible role of hemichannels are not available. Herein, we summarize the available data on the role of GJ channels in parasitic diseases and we also present new data suggesting that hemichannels might serve as key paracrine communication pathway during parasitic infections.

2. Gap Junction Channels and Hemichannels

Connexins (Cx) and pannexins (Panx) are members of two different GJ protein families in vertebrates [1]. Both protein subtypes can form channels that serve as pathways of cellular communication [1, 9]. Cxs and Panxs show similar membrane topology but only modest sequence homology [1]. In rodents and humans Cxs are encoded by 20 and 21 genes, respectively [10], whereas Panxs include only three members [11]. Moreover, innexins (Inxs) are members of a GJ family expressed only in invertebrates (Figure 1) [12]. They show similar membrane topology with Cxs and Panxs and can also form intercellular channels and hemichannels [13]. Inxs were originally identified in *Drosophila melanogaster* and *Caenorhabditis elegans*; however, Inx genes have been cloned recently from several other invertebrates (reviewed in [13]).

Does Nocturnality Drive Binocular Vision? Octodontine Rodents as a Case Study

Tomas Vega-Zuniga¹, Felipe S. Medina¹, Felipe Fredes¹, Claudio Zuniga¹, Daniel Severín¹, Adrián G. Palacios², Harvey J. Karten³, Jorge Mpodozis^{1*}

1 Departamento de Biología, Facultad de Ciencias, Universidad de Chile, Santiago, Chile, **2** Centro Interdisciplinario de Neurociencia de Valparaíso, Facultad de Ciencias, Universidad de Valparaíso, Valparaíso, Chile, **3** Department of Neurosciences, School of Medicine, University of California San Diego, La Jolla, California, United States of America

Abstract

Binocular vision is a visual property that allows fine discrimination of in-depth distance (stereopsis), as well as enhanced light and contrast sensitivity. In mammals enhanced binocular vision is structurally associated with a large degree of frontal binocular overlap, the presence of a corresponding retinal specialization containing a fovea or an area centralis, and well-developed ipsilateral retinal projections to the lateral thalamus (GLd). We compared these visual traits in two visually active species of the genus *Octodon* that exhibit contrasting visual habits: the diurnal *Octodon degus*, and the nocturnal *Octodon lunatus*. The *O. lunatus* visual field has a prominent 100° frontal binocular overlap, much larger than the 50° of overlap found in *O. degus*. Cells in the retinal ganglion cell layer were 40% fewer in *O. lunatus* (180,000) than in *O. degus* (300,000). *O. lunatus* has a poorly developed visual streak, but a well developed area centralis, located centrally near the optic disk (peak density of 4,352 cells/mm²). *O. degus* has a highly developed visual streak, and an area centralis located more temporally (peak density of 6,384 cells/mm²). The volumes of the contralateral GLd and superior colliculus (SC) are 15% larger in *O. degus* compared to *O. lunatus*. However, the ipsilateral projections to GLd and SC are 500% larger in *O. lunatus* than in *O. degus*. Other retinorecipient structures related to ocular movements and circadian activity showed no statistical differences between species. Our findings strongly suggest that nocturnal visual behavior leads to an enhancement of the structures associated with binocular vision, at least in the case of these rodents. Expansion of the binocular visual field in nocturnal species may have a beneficial effect in light and contrast sensitivity, but not necessarily in stereopsis. We discuss whether these conclusions can be extended to other mammalian and non-mammalian amniotes.

Citation: Vega-Zuniga T, Medina FS, Fredes F, Zuniga C, Severín D, et al. (2013) Does Nocturnality Drive Binocular Vision? Octodontine Rodents as a Case Study. PLoS ONE 8(12): e84199. doi:10.1371/journal.pone.0084199

Editor: Andrew Iwaniuk, University of Lethbridge, Canada

Received: August 5, 2013; **Accepted:** November 12, 2013; **Published:** December 31, 2013

Copyright: © 2013 Vega-Zuniga et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by Fondecyt 1080094, Fondecyt 1120124 to Jorge Mpodozis; Fondecyt 1090794, Iniciativa Científica Milenio ICM-P05-002, Proyecto Fondo Basal PFB-23-CONICYT to Rodrigo Vásquez. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: epistemo@uchile.cl

Introduction

Among vertebrates, the size, shape and position of the eyes and their orbits exhibit a high degree of variability, ranging from the small, unilaterally placed eyes of the flatfishes (*Achirus lineatus*), to the large, highly convergent frontal eyes of humans and other apes [1,2]. Peculiar (from a human perspective) placement of the eyes is not exceptional among fishes, perhaps reflecting the high diversity of morphological features present in these taxa. For instance, the eccentric lateral placement of the eyes of the hammerhead sharks (Sphyrnidae) endows them with a 360° span of the visual field in the horizontal and vertical dimensions, while retaining at the same time a significant amount of binocular overlap [3].

Among amniotes it is a well-established fact that convergent, more frontally oriented eyes, grant higher degrees of binocular overlap, while lateralized placement of eyes result in narrow binocular visual fields [1,4,5]. The degree of ocular convergence, in turn, is associated with differences in other functional aspects of the visual system, such as the presence and position of retinal specializations and the relative emphasis of the different retinal projections. In mammals, the position of the area centralis (the

retinal specialization that subserves the binocular area of the visual field) varies from temporal in species with lateralized eyes to pericentral in species with frontal eyes [6–8]. Furthermore, in birds and mammals, the relative size of the visual thalamofugal projection is larger in species with frontally-oriented eyes [4,9,10]. In mammals the retinal ipsilateral projections to the thalamic nucleus geniculatus lateralis pars dorsalis (GLd), the visual pathway that mediates functional binocular vision, is positively correlated with the degree of binocular overlap [11–14].

Various observations show that among amniotes feeding behavior influences eye orientation. Aerial and terrestrial predators (e.g. Felidae, Strigiformes and some Caprimulgiformes) have a larger degree of binocular overlap than their prey (ranging from large herbivores to small ground feeding birds) [4,15–17]. In addition, comparative studies indicate that irrespective of their feeding habits, nocturnal animals also exhibit a high degree of binocularity. Different taxa of nocturnal amniotes such as the grey-headed flying fox (Megachiroptera), the Galago (Galagidae), and even the nocturnal parrot Kakapo (Strigopidae), can be cited as representative examples of this tendency [18–20].

Computationally Efficient Methodology for Atomic-Level Characterization of Dendrimer–Drug Complexes: A Comparison of Amine- and Acetyl-Terminated PAMAM

Ariela Vergara-Jaque,[†] Jeffrey Comer,^{‡,§} Luis Monsalve,[†] Fernando D. González-Nilo,^{§,||,‡} and Claudia Sandoval^{*,§}

[†]Center for Bioinformatics and Molecular Simulation, Universidad de Talca, 2 norte 685, Talca-Chile

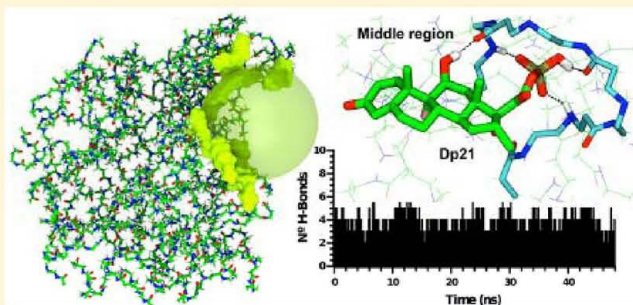
[‡]Fraunhofer Chile Research, Mariano Sánchez Fontecilla 310 piso 14, Las Condes, Santiago-Chile

[§]Universidad Andres Bello, Center for Bioinformatics and Integrative Biology, Facultad de Ciencias Biológicas, Av. Republica 239, Santiago-Chile

^{||}Centro Interdisciplinario de Neurociencia de Valparaíso, Universidad de Valparaíso, Gran Bretaña 287, Valparaíso-Chile

Supporting Information

ABSTRACT: PAMAM dendrimers have been widely studied as a novel means for controlled drug delivery; however, computational study of dendrimer–drug complexation is made difficult by the conformational flexibility of dendrimers and the nonspecific nature of the dendrimer–drug interactions. Conventional protocols for studying drug binding have been designed primarily for protein substrates, and, therefore, there is a need to establish new protocols to deal with the unique aspects of dendrimers. In this work, we generate cavities in generation-5 polyamidoamine (PAMAM) dendrimers at selected distances from the center of mass of the dendrimer for the insertion of the model drug: dexamethasone 21-phosphate or Dp21. The complexes are then allowed to equilibrate with distance between centers of mass of the drug and dendrimers confined to selected ranges; the free energy of complexation is estimated by the MM-GBSA (MM, molecular mechanics; GB, generalized Born; SA, surface area) method. For both amine- and modified acetyl-terminated PAMAM at both low and neutral pH, the most favorable free energy of complexation is associated with Dp21 at distance of 15–20 Å from the center of mass of the dendrimer and that smaller or larger distances yield considerably weaker affinity. In agreement with experimental results, we find acetyl-terminated PAMAM at neutral pH to form the least stable complex with Dp21. The greatest affinity is seen in the case of acetyl-terminated PAMAM at low pH, which appears to be due a complex balance of different contributions, which cannot be attributed to electrostatics, van der Waals interactions, hydrogen bonds, or charge–charge interactions alone.



■ INTRODUCTION

A recent trend in the pharmaceutical industry has been the focus on improving the properties of drugs that are currently used in various treatments, rather than focus on the creation of new drugs. Side effects in most drugs are considerable, in some cases more severe than the malady the drug is chosen to treat. To provide a solution to this problem, diverse nanocarriers have been proposed, such as micelles,¹ polymersomes,^{2–4} carbon nanotubes,^{5,6} nanospheres,^{7,8} and many others. These nanocarriers can reduce the side effects while improving targeting and controlled release all without changing the chemical structure of the drug.⁹

Dendritic nanoparticles are actually one of the most studied for use as nanocarriers,^{10,11} because these can enhance the transport and release of drugs due to their particular multifunctional structure. Dendrimers are synthetic hyper-

branched structures with a globular architecture. They may have hydrophobic or hydrophilic internal cavities, and their terminal groups can be modified with different molecular moieties in order to decrease cytotoxicity, improve bioavailability, and optimize transport of particular drugs.^{12–14} Dendrimers may encapsulate a drug through intermolecular interactions or, alternatively, form covalent bonds with a drug, and in this way enhance the uptake, cellular trafficking,¹⁵ and controlled release of the drugs. Polyamidoamine (PAMAM)^{10,11} is one of the most studied classes of dendrimers,^{16,17} and has widely been investigated for delivery of drugs targeting a diverse set of diseases.¹⁸ Studies have

Received: January 2, 2013

Revised: April 10, 2013

Published: May 3, 2013



Paclitaxel-PHBV nanoparticles and their toxicity to endometrial and primary ovarian cancer cells

Cristian Vilos^{a,c}, Francisco A. Morales^a, Paula A. Solar^a, Natalia S. Herrera^a, Fernando D. Gonzalez-Nilo^b, Daniel A. Aguayo^b, Hegaly L. Mendoza^b, Jeffrey Comer^{b,h}, Maria L. Bravo^d, Pamela A. Gonzalez^d, Sumie Kato^e, Mauricio A. Cuello^e, Catalina Alonso^f, Erasmo J. Bravo^f, Eva I. Bustamante^g, Gareth I. Owen^d, Luis A. Velasquez^{a,c,*}

^a Universidad Andres Bello, Facultad de Medicina, Center for Integrative Medicine and Innovative Science (CIMIS), Echaurren 183, Santiago, Chile

^b Universidad Andres Bello, Facultad de Biología, Center for Bioinformatics and Integrative Biology (CBIB), Republica 239, Santiago, Chile

^c Centro para el Desarrollo de la Nanociencia y Nanotecnología, Universidad de Santiago de Chile, Ecuador 3493, Santiago, Chile

^d Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Alameda 340, Santiago, Chile

^e Facultad de Medicina, Pontificia Universidad Católica de Chile, Alameda 340, Santiago, Chile

^f Hospital Gustavo Frick, Alvarez 1532, Viña del Mar, Chile

^g Fundación Arturo López Pérez, Rancagua 878, Santiago, Chile

^h Fundación Fraunhofer Chile Research, M. Sánchez Fontecilla 310 piso 14, Las Condes, Chile

ARTICLE INFO

Article history:

Received 28 November 2012

Accepted 12 February 2013

Available online 5 March 2013

Keywords:

Paclitaxel

PHBV

Nanoparticle

Ovarian cancer

Drug delivery

Molecular dynamics simulations

ABSTRACT

This report is an integrated study to include the molecular simulation, physicochemical characterization and biological analysis of a paclitaxel-loaded PHBV nanoparticle that demonstrates uptake, release and cytotoxicity in cancer cell lines. Taking this nanoparticle one step closer to its use in a clinical setting, we demonstrate that it causes significant cell death in primary cultures of stage IIIc serous ovarian cancer cells isolated from six patients. Molecular simulations revealed a high affinity of paclitaxel for the water–polymer interface, thus the drug is delivered only when the polymer near it is degraded. The Fourier transform infrared spectroscopy suggests the formation of a short-lived crystalline phase, also observed in the CG simulations, and transmission electron microscopy revealed branched structures on the surface of particles, which disappeared after 4 days. Biological analyses indicated that these particles have a 48-h window of toxicity protection, allowing for the endocytosis of the particle by the cells; this finding was corroborated by confocal microscopy and flow cytometry. The low cost to synthesize PHBV using microorganisms and the potential chemical modifications of the polymer make it attractive for inexpensive, large-scale pharmaceutical production.

© 2013 Elsevier Ltd. All rights reserved.

1. Introduction

The worldwide cancer burden continues to grow and is becoming a major economic expenditure for all developed countries. Although the 5-year survival for ovarian cancer patients has improved to nearly 30%, this disease remains the fifth deadliest cancer among American women and is the leading cause of death from gynecologic malignancy. In 2010, 21,880 new cases and 13,850 new deaths were recorded in the United States [1,2].

The major obstacles in the field of oncology include the tremendous effort that is required for early detection, the development of many new drugs and drug delivery systems and the existence of effective chemotherapeutic agents [3,4]. One additional challenge is the development of nanoparticles that meet the clinical demands of therapeutic agents, such as biocompatibility, biodegradability, a clinically relevant circulating half-life, a low rate of intravascular aggregation, and a long-term storage capacity [5]. During the last decade, drug delivery systems based on polymeric nanoparticles have been central to many significant advances in nanomedicine [6]. Accordingly, these particles have generated interest related to their potential use in preclinical and clinical cancer drug development [5].

Polymeric nanoparticles have numerous uses in drug delivery, including the specific and targeted delivery of therapeutic agents,

* Corresponding author. Center for Integrative Medicine and Innovative Science (CIMIS), Universidad Andres Bello, Facultad de Medicina, Echaurren 183, 8370071, Santiago, Chile.

E-mail address: luis.velasquez@unab.cl (L.A. Velasquez).

Annex 4

Organization of Scientific Events



CINV

II MEETING MILLENNIUM INSTITUTE CINV

"The science of magic, when attention meets illusion"

Organized by the PhD students of the Millennium Institute
"Centro Interdisciplinario de Neurociencia de Valparaíso"

August 1, 2013
Valparaíso, Chile



INSTITUTO DE
Sistemas Complejos
DE VALPARAÍSO



CENTRO INTERDISCIPLINARIO DE
Neurociencia de
Valparaíso



September 23rd - 30th
CINV - Universidad de Valparaíso

More information:
www.cinv.cl

For funding and scholarships contact:
Alejandra Pinto (alejandra.pinto@cinv.cl)

Deadline: August 1st, 2013



Introduction,
types of ion channels,
convergence, essential
techniques for studying
ion channels and
Transporters

Function
Electrophysiology -
Voltage clamp techni-
ques



Structure
Biochemistry - Membrane Proteins Biochemistry
Molecular Biology of ion channels

Fluorescence - Voltage clamp fluorimetry, LRET,
patch clamp fluorimetry, real time fluorescence
spectroscopy

Electronic paramagnetic resonance
Crystallography

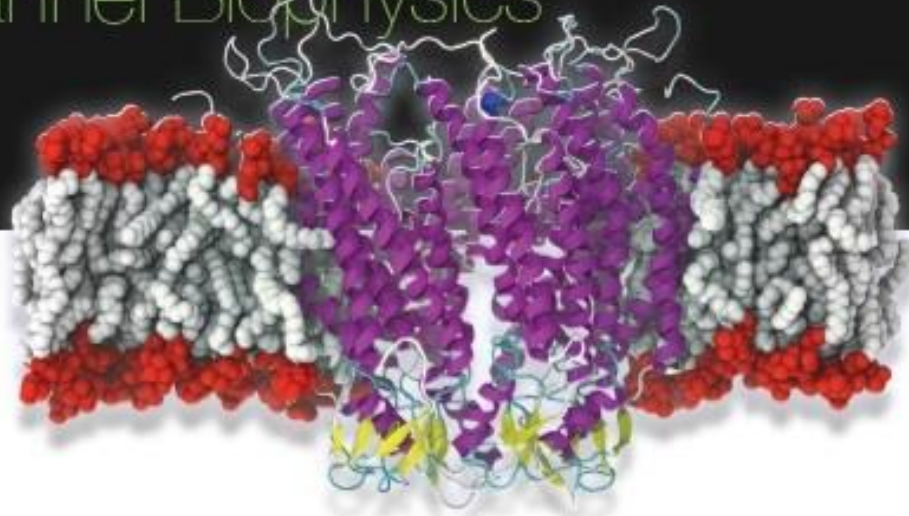
Channels in native systems
Smooth Muscle
Nanotechnology - Remote control of ion channels

Channels and diseases
Potassium channels
Hemichannels, connexins and pannexins



Course / Workshop - Ion Channels: Structure, function and disease

Latin american school on Ion Channel Biophysics



Design: Juan Carlos Gendel, CINV

Organizers:

Ramón Latorre (Chile)
Carlos González (Chile)
Teresa Giráldez (Spain)

Teaching staff:

Héctor Barajas (USA) James Hall (USA)
Francisco Bezanilla (USA) Miguel Holmgren (USA)
Sebastián Brauchi (Chile) Peter Larsson (USA)
Jorge Contreras (USA) Alan Neely (Chile)
Luis Cuello (USA) Verónica Milesi (Argentina)
Gonzalo Ferreira (Uruguay) Pablo Miranda (Spain)
Ehud Isacoff (USA) Matthias Salathe (USA)

Teaching assistants:

David Báez (Chile)
Juan Pablo Castillo (Chile)
Gustavo Contreras (Chile)

Organizers:



CENTRO INTERDISCIPLINARIO DE
Neurociencia de
Valparaíso



Support:





Centro Interdisciplinario de
Neurociencia de Valparaíso



Robert Brooks Phillips
Professor at the Department
of Applied Physics
at the California Institute
of Technology (CalTech)



Jane Kondev
Professor at the Department
of Physics, Quantitative
Biology at Brandeis
University



Hernán Gustavo García
Postdoctoral fellow at
Princeton University

January 8th, 2013
10:00 Hrs

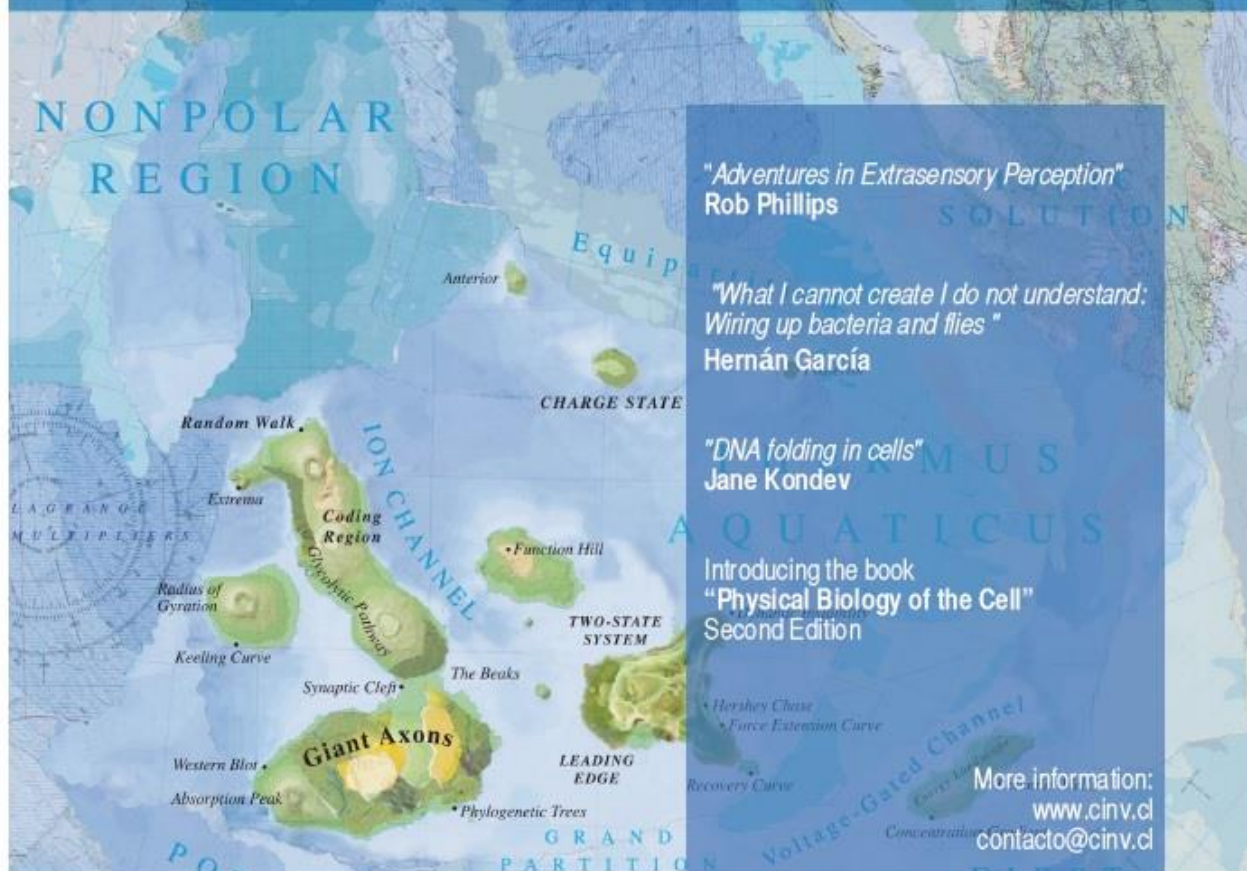
Auditorio Juan Araya
Facultad de Arquitectura
Universidad de Valparaíso

Symposium

Rob Phillips
Jane Kondev
Julie Theriot
Hernán G. García

Illustrated by Nigel Orme

PHYSICAL BIOLOGY OF THE CELL SECOND EDITION





VIII Congreso Iberoamericano de Biofísica

IX Reunión Anual Sociedad Chilena de Neurociencia

1º - 4 de Octubre, 2013
Parque Cultural de Valparaíso
CHILE

CONFERENCIAS

Ramón Latorre - CINV Universidad de Valparaíso, Chile
Emanuel C. Mora - Universidad de La Habana, Cuba
Enrico Nasi - Universidad Nacional de Colombia
Gail Mandel - HHMI, Vollum Institute, Oregon USA
Paul Brehm - Vollum Institute, Oregon H & S University USA
Fernando Torrealba - Pontificia Universidad Católica de Chile
Marcelo Morales - Instituto de Biofísica Carlos Chagas Filho, Brasil

SIMPOSIOS

Membranes - N. C. Santos, L. Bagatolli
New insights on calcium signaling - A. Escobar, E. Ríos
Diversity of chemical senses - J. Badgalupo, J. Alcayaga
Exocytosis, neurotransmission and synapse structure - A.M. Cárdenas, B. van Zundert
Synaptic plasticity - P. Muñoz, C. Hidalgo
Protein structure - E. Perozo
Role & regulation of channels and hemichannels formed by connexins or panexins in the nervous system - J.C. Sáez, A. Martínez
Calcium channels - E. Jaimovich, D. Varela
Lightning the structure and function of ion channels - T. Giráldez, C. González

Más información en: www.socneurociencia.cl | Inscripciones hasta el 31 de agosto 2013

Respaldado por

CENTRO INTERDISCIPLINARIO DE
Neurociencia de
Valparaíso



PCdV

[PARQUE CULTURAL DE VALPARAÍSO]



Auspiciado por



Annex 7.3

Articles and Interviews

Title: "In Valparaíso was launched TV series about neuroscience and life mysteries"

Media: www.envalparaiso.cl Informative web page

Date: June 21st, 2013

envalparaiso gastronomía hospedaje artesanía salas y museos

Lanzan serie científica acerca la neurociencia y los misterios de la vida



Lanzamiento de serie científica acerca la neurociencia y los misterios de la vida

viernes 21 de junio de 2013

Abordar de forma interactiva temas de interés universal como la belleza, la ilusión, las emociones, entre otros, desde las perspectivas de la neurociencia y de la innovación es el objetivo del Centro Interdisciplinario de Neurociencia (CINV) de la Universidad de Valparaíso en conjunto con Novasur, Televisión Educativa del Consejo Nacional de Televisión (CNTV) con el lanzamiento de la serie "Neuromantes. Exploradores de la vida" que se realizará el próximo jueves 27 de junio.

El proyecto televisivo, el cual fue financiado gracias al Primer Concurso de Divulgación y Valoración de la Ciencia y la Tecnología del Programa EXPLORA CONICYT, se complementa con la visión de personalidades relevantes de otros ámbitos de la cultura y de las artes de la región (ver información anexa en dossier).

Valparaíso es otro de los protagonistas, ya que en cada capítulo se hace referencia a la ciudad patrimonial, la cual sirve como locación o telón de fondo para cada capítulo.

Además, la serie, a través de Novasur, tiene como fin proporcionar material audiovisual y complementario en apoyo al currículum escolar en materias como neurociencias, innovación, arte, literatura e historia, además de transmitir y proyectar a nivel nacional e internacional el conocimiento científico generado por el CINV.

La actividad contará con la presencia de autoridades de las diferentes instituciones involucradas, así como también del gobierno regional y en especial, la del alcalde de la Ilustre Municipalidad de Valparaíso, Jorge Castro, quien firmará un convenio de emisión de la serie en las diferentes oficinas municipales con atención de público y que cuentan con televisor.

El evento se llevará a cabo en la Biblioteca Santiago Severín de Valparaíso a las 19 horas y es abierta a todo público.

Title: "Scientific TV Series set in Valparaíso tries to resolve life mysteries"

Media: www.elfmartutino.cl Online Newspaper

Date: June 24th, 2013

Serie científica hecha en Valparaíso intenta resolver los misterios de la vida

El proyecto televisivo es realizado por el Centro Interdisciplinario de Neurociencia (CINV) de la Universidad de Valparaíso en conjunto con Novasur, Televisión Educativa del Consejo Nacional de Televisión (CNTV)



Por El Martutino
523 Lecturas

24 de Junio, 2013 00:06

[Comentar](#)

Abordar de forma interactiva temas de interés universal como la belleza, la ilusión, las emociones, entre otros, desde las perspectivas de la neurociencia y de la innovación es el objetivo del Centro Interdisciplinario de Neurociencia (CINV) de la Universidad de Valparaíso en conjunto con Novasur, Televisión Educativa del Consejo Nacional de Televisión (CNTV) con el lanzamiento de la serie "Neuromantes. Exploradores de la vida" que se realizará el próximo jueves 27 de junio.



El proyecto televisivo, el cual fue financiado gracias al Primer Concurso de Divulgación y Valoración de la Ciencia y la Tecnología del Programa EXPLORA CONICYT, se complementa con la visión de personalidades relevantes de otros ámbitos de la cultura y de las artes de la región (ver información anexa en dossier).

Title: "CINV and Novasur launched Neuromantes"

Media: www.elfmartutino.cl Online Newspaper

Date: June 24th, 2013

CINV y Novasur lanzan Neuromantes

Como programa de divulgación científica, *Neuromantes* tiene como objetivo dar a conocer a la región como un polo de la generación de conocimiento de nivel mundial.



Por Novasur Chile
369 Lecturas

26 de Junio, 2013 09:06

[Comentar](#)

El año 2012 Novasur la televisión educativa del Consejo Nacional de Televisión, CNTV, convocó en cada región del país a actores significativos de la comunidad para co-producir audiovisuales educativos, que fueran representativos de los temas e intereses propios de las región y de las comunidades locales.

En la quinta región, gracias a una alianza con el Centro Interdisciplinario de Neurociencias de Valparaíso, CINV de la Universidad de Valparaíso, surgió *Neuromantes. Exploradores de la Vida*, serie de divulgación científica que trata temas universales como la Belleza, Memoria, Percepción y otros desde la mirada de la neurociencia, la innovación y la cultura. El nombre del programa viene de las palabras *neuro*:sistema nervioso o mente y *mantes*: magia.

Este próximo 27 de junio se realizará el lanzamiento de la serie, que consta de seis capítulos y que fue financiada por Explora-CONICYT -a través del Primer Concurso de Productos de Apropiación y Divulgación de la Ciencia y la Tecnología- y por el CNTV. La producción estuvo a cargo de Cábala Producciones, reconocidos realizadores con amplia experiencia, creadores de programas como *Cuaderno de Ciencias* y *Cambio Global*.

Title: "Launch: Neuromantes, Exploradores de la vida "

Media: www.sientevalpo.cl Informative web page

Date: June 26th, 2013

SIENTE VALPO
CULTURA DESDE LA V REGION

TAGS MÚSICA
TEATRO
f e

LANZAMIENTO: NEUROMANTES, EXPLORADORES DE LA VIDA

26 junio, 2013 11:17 pm Publicado por Viví Gutiérrez B. | Deje sus pensamientos

Participa este jueves 27 de junio en el lanzamiento de la serie "Neuromantes. Exploradores de la vida", que se realizará en la **biblioteca Santiago Severín de Valparaíso** a las 19 horas.

Esta nueva serie científica toca de forma interactiva temas de interés universal como la belleza, la ilusión, las emociones, entre otros. El proyecto está bajo la perspectiva del **Centro Interdisciplinario de Neurociencia de la U. de Valparaíso** junto con **Novasur**, Televisión Educativa del Consejo Nacional de Televisión.

Entrada liberada

¡Están todos invitados a explorar nuestra ciudad patrimonial de una manera distinta y nueva!

En estudio:



Title: "Launch of scientific series that tries resolving life mysteries"

Media: www.ucv.cl

Date: June 27th, 2013



103.5 FM

Valparaíso



[Inicio](#) | [UCV Radio Noticias](#) | [Avenida 80](#) | [Golazo](#) | [Dimensión Latinoamericana](#) | [UCV Radio](#)

[RSS](#) | [Subscribe via E-mail](#) | [Síguenos en Twitter](#) | [Agréganos en Facebook](#)

Lanzamiento de serie científica intenta resolver los misterios de la vida

27 de junio de 2013

Abordar de forma interactiva temas de interés universal como la belleza, la ilusión, las emociones, entre otros, desde las perspectivas de la neurociencia y de la innovación es el objetivo del Centro Interdisciplinario de Neurociencia (CINV) de la Universidad de Valparaíso en conjunto con Novasur, Televisión Educativa del Consejo Nacional de Televisión (CNTV) con el lanzamiento de la serie "Neuromantes. Exploradores de la vida" que se realizará el próximo jueves 27 de junio.

El proyecto televisivo, el cual fue financiado gracias al Primer Concurso de Divulgación y Valoración de la Ciencia y la Tecnología del Programa EXPLORA CONICYT, se complementa con la visión de personalidades relevantes de otros ámbitos de la cultura y de las artes de la región.



Valparaíso es otro de los protagonistas, ya que en cada capítulo se hace referencia a la ciudad patrimonial, la cual sirve como locación o telón de fondo para cada capítulo.

Además, la serie, a través de Novasur, tiene como fin proporcionar material audiovisual y complementario en apoyo al currículum escolar en materias como neurociencias, innovación, arte, literatura e historia, además de transmitir y proyectar a nivel nacional e internacional el conocimiento científico generado por el CINV.

La actividad contará con la presencia de autoridades de las diferentes instituciones involucradas, así como también del gobierno regional y en especial, la del alcalde de la Ilustre Municipalidad de Valparaíso, Jorge Castro, quien firmará un convenio de emisión de la serie en las diferentes oficinas municipales con atención de público y que cuentan con televisor.

El evento se llevará a cabo en la Biblioteca Santiago Severín de Valparaíso a las 19 horas y es abierta a todo público.

Title: “Universidad de Valparaíso and Novasur will broadcast scientific serie “Neuromantes: Exploradores de la vida”

Media: www.elepicentro.cl, online newspaper

Date: June 28th, 2013

Universidad de Valparaíso y Novasur emitirá serie científica “Neuromantes: exploradores de la vida”



“Neuromantes: exploradores de la vida” es el nombre de la nueva serie científica que a partir de julio se emitirá a través de las redes asociadas a Novasur, canal educativo del Consejo Nacional de Televisión. La serie la protagonizan científicos del Centro Interdisciplinario de Neurociencia de la Universidad de Valparaíso (CINV), junto a personajes del mundo del arte y la cultura.

“El objetivo es mostrar la ciencia como parte de nuestra cultura”, explicó el doctor Ramón Latorre, director del CINV, en el lanzamiento de la serie, realizado en la tarde del jueves en la Biblioteca Severín.

“Esto es un empujón que nos da, por un lado, el Instituto Milenium, nuestro Centro Interdisciplinario de Neurociencia de Valparaíso (CINV), y por otro la Universidad de Valparaíso, para llevar la ciencia a la calle, la ciencia al ciudadano, la ciencia a los niños. Nuestro deber como miembros de esta sociedad, porque recibimos un financiamiento del Estado, es comunicar a la gente lo que hacemos, y tratamos de transmitirlo de una manera cariñosa, de una manera que llegue naturalmente y que no confunda la ciencia con la tecnología, sino que la ciencia se perciba más bien como hermana del arte, de las humanidades y de la cosa social como un todo, porque la ciencia es parte de la cultura”, afirmó Latorre.

Title: “Squid is used to study the epilepsy and muscular paralysis”

Media: www.soyvalparaiso.cl Online Newspaper

Date: October 07th, 2013

Usan jibias para estudiar el impulso eléctrico de las neuronas

07.10.2013

Centro de investigación de Montemar desarrolla investigación sobre este calmar desde hace 40 años.



Seguir a @soyvalparaiso

La jibia no sólo es un molusco que busca imponerse en la cocina de numerosos países. Es además, el objeto de estudio de científicos nacionales que desde Montemar en el camino costero Viña-Concón, investigan las propiedades eléctricas de las neuronas y la importancia de este impulso para múltiples funciones del organismo, entre ellas, el movimiento muscular.

Uno de los científicos que se encuentra estudiando al calamar es el chileno Miguel Holmgren, quien está radicado en Estados Unidos. Según comentó, junto con estar “lleno de electricidad”, la jibia posee los axones más grandes de la naturaleza -de 1 mm aproximadamente-, los que pueden ser percibidos por el ojo humano.

“Una vez generada la señal eléctrica, las neuronas tienen la propiedad de poder conducirla a través de estas prolongaciones llamadas axones las que, a modo de graficar, serían los cables del circuito eléctrico”, comenta Holmgren, quien además es presidente de la Sociedad de Biofísicos Latinoamericanos (Sobla).

Title: "Open green building in house of Playa Ancha"

Media: La Estrella de Valparaíso, Newspaper

Date: October 08th, 2013

Actualidad



DAVIDE LA JORNADA DE AÑOS, EL LOCAL FUNCIONÓ SIN LA RESPECTIVA MAQUINARIA

Auto loco provocó caos en un McDonald's y dejó heridos a dos brasileños

El hecho ocurrió en Placilla y fue provocado por un choque en cadena.

Gratificación Matus O.

A las 9.20 horas, una tarde de domingo. Una mujer, de iniciales U.F.S.F., llega con su Suzuki Grand Nismo hasta el servicio al cliente, ubicado en el kilómetro 163 de la ruta 68, en Placilla. Al mismo tiempo, un Volkswagen Golf blanco se encuentra estacionado frente a un local de McDonald's, que está a metros de la bomba, con tres personas en su interior.

Hasta aquí todo está en perfecto orden, pero... cuando la mujer intenta salir de la bomba el vehículo que conduce, inesperadamente pierde el control y se lanza rápidamente hacia el auto estacionado, impactándolo por atrás. Como resultado, el Volkswagen traspana la rampa del local, arrojando a una joven turista brasileña, que como en una mesa junto a su pequeño hijo de tres años.

INFORMANTES

según la información entregada por Carabineros, las personas que se encontraban en el vehículo blanco



EL MOMENTO EN QUE EL VEHÍCULO TRASPASÓ EL VENTANAL.

AMUCHO FRIJO

«Durante la mañana de ayer, afuera del local de comida rápida aún quedaban restos de vidrios sobre evidencia del siniestro accidental. De la mañana ya no quedaba absolutamente nada, por lo que algunos trabajadores comentaban que se sentían felices, ya que a esa hora las temperaturas eran bajas. También señalan que quedaron asustados, porque les pareció 'de película' lo que sucedió».

trahen en el vehículo blanco estaban esperando a tres menores que realizaban un picnic, quienes, por fortuna, salieron ilesos.

Los turistas, que fueron atendidos por personal del hospital Carlos Van Buren de Valparaíso, no corrieron la misma suerte, ya que I.C.E. de 3 años, quedó con contusión y erosión de rodilla derecha, de carácter

leve, mientras que D.E.F.S. de 26 años, resultó con una herida con una cortante de tobillo derecho, de carácter leve.

Respecto a la madre del conductor, la autoridad policial indicó que conlleva a "terminación de temerancia", pero sin la documentación del vehículo, por lo cual el asunto quedó a disposición del Tercer Juzgado de Policía Local, el que adoptará el procedimiento por cuasidelito de lesiones leves en amparo y datos.

Respecto al local de McDonald's, éste resultó con daños estructurales en una de las mamparas y la base de ésta, cuyos daños también serán evaluados en el tribunal. Cabe señalar que tanto a la mujer como al conductor del Volkswagen que tenía todos los documentos al día se les realizaron las alcoholometrías pertinentes.



AGRICULTOR AGRIERO

Emergencia agrícola en 21 comunas

«Las inusuales heladas que cayeron en las últimas semanas y que han complicado la producción de frutas, hortalizas y hasta flores, motivaron que el Intendente de Agricultura decretara emergencia agrícola para 21 comunas de la V Región. Estas son: Calle Larga, Los Andes, Riconada, San Esteban, Limache, Olmué, Zaldívar, La Laguna, Penco, Zapallar, Hijaes, La Cruz, Nogales, Quilón, Cartagena, San Antonio, Catemu, Paiguano, San Felipe, Santa María y Cabañeros».

La mujer que cultiva el "alimento sagrado"

«Cristina Pizarro no es una agricultora convencional. Después de haber vivido en Santiago, Brasil y Estados Unidos, viajó un día la necesidad de volver a su tierra. Cabildo, e iniciar un viaje espiritual, donde se encontró con lo que hoy es su fuente de trabajo: el amaranto, alimento sagrado de los incas, que hoy compete con la quinoa en cuanto a poderes medicinales y de pasto, es el emblema de los ecologistas, pues la planta crece en terrenos de cultivos transgénicos en Estados Unidos. Una noche, instalada en el valle de Fubión, en la V Región interior, Cristina tuvo un sueño revelador: un hombre le habló del "alimento del futuro", se trataba de la brecha inca, alimento que posee varias



LA AMARANTO EN LA QUINUA

propiedades medicinales que actúan sobre el cuerpo y, dicen, también el espíritu. Su primera siembra de amaranto dio como resultado 250 kilos de semillas. Hoy, Cristina tiene un invernadero y su emprendimiento, levantado con apoyo de Indap, crece. «Siento que el amaranto va a ser de mucha utilidad a la humanidad al ser una alternativa eficaz de alimentación que nutre y sana el alma», asegura la cabildana.

Inauguran edificio verde en casona de Playa Ancha

«Mañana será inaugurado oficialmente el edificio verde de "Ciencia Al Tiro", un nuevo espacio tecnológico y sustentable, dirigido a la comunidad escolar y de estudiantes, ubicado en calle Pedro León Gallo, en Playa Ancha. La edificación, una casa patrimonial del sector, permitirá la realización de charlas y talleres que promuevan el intercambio de ideas a nivel local, nacional e internacional».

Recolector de basura traficaba droga en Placeres

«Carabineros del retiro Cabo I° Juan Silva Toro, de corno Los Placeres, arrestó a un recolector de basura municipal por microtráfico. La policía sorprendió a R.A.D.Z., de 52 años, alias "Conejo", en la población Los Placeres. Andaba trabajando en el caso, pero a la vez vendía droga a adictos. Portaba 22 envoltorios de marihuana prensada y \$71.600 de las ganancias. En su hogar de Rodellón se hallaron otros 280 gramos de la yerba a granel. Pasó al juzgado de garantía».



EL VEHÍCULO QUE SE ACCIDENTÓ AL PERCUTIR A VEHÍCULO

Motorista de Carabineros se accidentó en patrullaje

«Con lesiones de consideración resultó ayer un efectivo policial en un accidente de tránsito. El siniestro se registró pasada las 17.30 horas en calle Norwaga N°190, en dirección al centro, donde se vieron involucrados un motorista de Carabineros, un taxi colectivo y una furgoneta. El jefe de la Tercera Comisaría Norte, capitán Pablo Fischel, expresó que el caso 2° de una patrulla se encontraba en el segundo turno de patrullaje preventivo cuando ocurrió el incidente. Al parecer, el taxi colectivo se detuvo en un paradero a dejar pasajeros, generando que el vehículo particular que

lo antecedía frenara en forma intempestiva. Este fue impactado por el motorista que cayó al suelo y quedó con contusiones en la cervical. Fue trasladado al hospital Van Buren a constatar lesiones. Los chóferes fueron al mismo centro de salud para los exámenes de alcoholometría. Personal de la Sst efectuó peritajes».



EL VEHÍCULO QUE SE ACCIDENTÓ AL PERCUTIR A VEHÍCULO

Inauguran edificio verde en casona de Playa Ancha

● Mañana será inaugurado oficialmente el edificio verde de "Ciencia Al Tiro", un nuevo espacio tecnológico y sustentable, dirigido a la comunidad escolar y de estudiantes, ubicado en calle Pedro León Gallo, en Playa Ancha. La edificación, una casa patrimonial del sector, permitirá la realización de charlas y talleres que promuevan el intercambio de ideas a nivel local, nacional e internacional.

Title: "Provide house for children learn funny science "

Media: El Mercurio de Valparaíso, Newspaper

Date: October 10th, 2013



RECICLAJE.— Con materiales nobles rescatados de demoliciones, una vieja caserna del barrio de Playa Ancha fue recuperada y dotada de tecnologías eficientes y sustentables para fomentar la ciencia entre los niños.

Edificio Verde, en Valparaíso:

Equipan casa para que niños aprendan ciencia entretenida

Programa "Ciencia al tiro" fomenta el interés científico bajo el alero del Centro Interdisciplinario de Neurociencia.

HERNÁN CISTERNA ARELLANO

Cientos de alumnos de 7° y 8° año básico que estudian en escuelas municipales de sectores vulnerables de Valparaíso protagonizan una experiencia científica única en Chile y que, desde ayer, cuenta con un espacio propio: el Edificio Verde.

Se trata de una tradicional caserna del barrio de Playa Ancha que fue reciclada y equipada con tecnologías eficientes y sustentables.

Allí los monitores, que son candidatos a doctorados y magísteres del Centro Interdisciplinario de Neurociencia de Valparaíso (CINV) —que dirige el biofísico Ramón Latorre—, realizarán talleres que buscan interesar a los niños en

materias científicas.

Estos estudiantes ya participan en forma periódica de las iniciativas del proyecto "Ciencia al tiro" que creó la neurobióloga norteamericana y profesora del CINV Kathleen Witlock.

La experiencia partió el 2008 en la escuela de la población Montedónico, uno de los sectores de mayor vulnerabilidad del puerto.

En estos talleres, los alumnos aprendieron que a través de un horno solar podían calentar agua, cocinar galletas o preparar huevos duros sin necesidad de otro tipo de energía. También generaron biogás a partir de los desechos orgánicos.

Incluso avanzaron en conocimientos sobre genética, el desarrollo del sistema nervioso y su

funcionamiento, y la acuaponía (sistema sustentable de producción de plantas y peces) a través de su trabajo con moscas del vino y peces cebra.

"Aquí los niños pueden utilizar los equipos, microscopios y lupas, lo que es 'bacán'. En otras partes les prohíben tocar todo. Descubren que la ciencia es superinteresante y divertida", subraya Kathleen Witlock.


Ramón Latorre, Premio Nacional de Ciencias 2002, expresó al respecto que "un país sin ciencia es un país cojo. Chile tiene poca ciencia. Lo que está haciendo Kate es un proyecto que llega a los niños. Es ahí donde uno tiene que empezar a enseñar la ciencia, a despertar la curiosidad".

Title: “Green Building of Playa Ancha turns schoolchild into small Scientists”

Media: El Mercurio de Valparaíso, Newspaper

Date: November 06th, 2013

Fotografía: Media Training Consultores
pemoreno@mercuriovalpo.cl

Con la presencia del alcalde de Valparaíso, Jorge Castro; el director regional del Consejo Nacional de la Cultura y las Artes, Rafael Torres; el rector de la Universidad de Valparaíso, Aldo Valle, y el director del Centro Interdisciplinario de Neurociencia de Valparaíso, Ramón Latorre, se realizó la inauguración del primer edificio verde en Chile, dedicado a la educación de la ciencia. El espacio ubicado en el Cerro Playa Ancha, pertenece a la iniciativa Ciencia Al Tiro, que dirige la Dra. Kathleen Whitlock, quien además es integrante del CINV. 



Ramón Latorre, director Centro Interdisciplinario Neurociencia de Valparaíso; Aldo Valle, rector Universidad de Valparaíso; Kathleen Whitlock, directora Ciencia Al Tiro, y Jorge Castro, alcalde de Valparaíso.



Kathleen Whitlock, directora Ciencia Al Tiro, junto a alumnos de la escuela básica Pacífico de Playa Ancha, Valparaíso.



Moisés Acevedo, Centro Interdisciplinario Neurociencia de Valparaíso; Pablo Muñoz, Universidad de Valparaíso; Alejandra Pinto, Universidad de Valparaíso, y David Carrillo, director de Extensión y Comunicaciones Universidad de Valparaíso.



Carlos Briceño, consejero regional; Víctor Fuentes, director regional de Corfo, y Agustín Martínez, Centro Interdisciplinario Neurociencia de Valparaíso.



Juan Carlos García, Centro Interdisciplinario Neurociencia de Valparaíso; Rodrigo Sepúlveda, seremi de Energía; Patricia Arancibia, directora edificio verde Ciencia Al Tiro, y John Ewer, Centro Interdisciplinario Neurociencia de Valparaíso.

Title: "TV Series set in Valparaíso talks about beauty from the science"

Media: www.biobiochile.cl

Date: December 08th, 2013



MUJER

Domingo 8 diciembre 2013 | 20:47 · Actualizado: 21:48

Serie televisiva ambientada en Valparaíso aborda el concepto de belleza desde la ciencia



277



www.cinv.uv.cl

Title: "TV series Neuromantes"

Media: www.24horas.cl

Date: December 11th, 2013



Valparaíso: Neuromantes, exploradores de la vida

Es la serie científica que produce el Centro de Neurociencia de la Universidad de Valparaíso y que busca acercar la ciencia a las personas. ¿Quiere saber de qué se trata? Entonces ponga atención a la siguiente nota.

POR **NATALIA GUTIÉRREZ**

Email [Twitter](#) 3 [Facebook](#) 0 [Compartir](#) 0

11

diciembre
2013

Title: “ Iberoamerican Congress of Biophysics in Valparaíso”

Media: www.soyvalparaiso.cl

Date: September 30th, 2013

soyvalparaiso.cl

Ciudades

Diarios

Blogs

« Regresar a soychile.cl

DEPORTES

ESPECTÁCULOS

POLICIAL

SOCIEDAD

POLÍTICA

TECNOLOGÍA

CULTURA

Todas

Arica

Iquique

Calama

Antofagasta

Copiapó

Valparaíso

Quillota

San Antonio

Chillán

San Carlos

Tomé

Talcahuano

Concepción

Coronel

Arauco

Temuco

Valdivia

Osorno

Puerto Montt

Chiloé

2

f Compartir

10

Twitter

g+ +1

Comentar

Imprimir

Comienza en Valparaíso el Congreso Iberoamericano de Biofísicos

Destacado científicos del área se darán cita hasta el jueves en la ciudad Puerto. Mañana el doctor Miguel Holmgren dictará conferencia sobre la electricidad como motor de los seres vivos.

30.09.2013

#BIOFÍSICOS

f

Twitter

g+

Comentar

Imprimir

Seguir a @soyvalparaiso

En el Parque Cultural de Valparaíso (ex Cárcel) se desarrollará entre mañana y el viernes el VIII Congreso Iberoamericano de Biofísicos, que en esta ocasión estará dirigido a unos 300 escolares de la Región. Para mañana destaca la conferencia sobre la importancia de la electricidad en seres vivos, que dictará el científico chileno que trabaja en Estados Unidos, Miguel Holmgren, quien además es Presidente de la Sociedad de Biofísicos Latinoamericanos.

La jornada inaugural también será presidida por Emanuel Mora, científico cubano experto en audición y bioacústica de humanos, y diversos animales tales como: murciélagos, mariposas, ranas y delfines.

El VIII Congreso Iberoamericano de Biofísicos se realizará de forma conjunta con la IX Reunión Anual de la Sociedad Chilena de Neurociencias. Se trata de la primera vez que esta cita se desarrollará en Valparaíso. El evento contará, además, con la participación de 80 científicos de Chile y el extranjero.

Entre los principales expositores destacan: Paul Brehm y Gail Mandel, deVollum Institute, Portland, Estados Unidos; Enrico Nasi, de la Universidad Nacional de Colombia; Marcelo Morales, de la Universidad Federal de Espírito Santo, Brasil; y Ramón Latorre, Director del Centro Interdisciplinario de Neurociencia de la Universidad de Valparaíso, CINV, y Premio Nacional de Ciencias.

Alan Neely, subdirector del Centro Interdisciplinario de Neurociencia de Valparaíso (CINV), sostuvo que las expectativas respecto de la realización del encuentro científico se elevadas y se orientan a "generar intercambio de ideas, fortalecer redes, y sobre todo, lograr una actualización de conocimientos".

soyvalparaiso.cl

f Me gusta

REGIÓN. El proyecto piloto consta de la cobrerización de los cinco tipos de pasamanos en dos coches. Así se eliminan virus y bacterias.

18 millones de pasajeros transportó Metro Vía Rapido durante el año 2012 en sus 27 tramos.

Polio Bermúdez, su par de Miriam, Hernán de Solís fue el presidente ejecutivo de Codelco, Thomas Butler, el presidente del Directorio Víctor Valparaíso, José Luis Domínguez y el vicepresidente de Abasco S.A., Julio Friedmann.



Duoc UC
UNIVERSIDAD DE CHILE

**CENTRO DE EXTENSIÓN
ESPACIO COLEGIO**

Cartelera Cultural Octubre 2013
Español y Comunicación



Con la COLA de febrero 2010, miércoles 2 de octubre 10:00 hrs.

El teatro es una forma de arte que surge en el siglo XVIII cuando los salones de la nobleza se transformaron en escenarios para la representación de obras teatrales.

Conferencia: "YOU CITT" Editorial Vozes, 14 de octubre 10:00 hrs. 10:00 hrs.

El teatro es una forma de arte que surge en el siglo XVIII cuando los salones de la nobleza se transformaron en escenarios para la representación de obras teatrales.

Conferencia: "YOU CITT" Editorial Vozes, 14 de octubre 10:00 hrs. 10:00 hrs.

El teatro es una forma de arte que surge en el siglo XVIII cuando los salones de la nobleza se transformaron en escenarios para la representación de obras teatrales.

Conferencia: "YOU CITT" Editorial Vozes, 14 de octubre 10:00 hrs. 10:00 hrs.

El teatro es una forma de arte que surge en el siglo XVIII cuando los salones de la nobleza se transformaron en escenarios para la representación de obras teatrales.

Desarrolla el rol ministerial del mero, Gloria Huallata, quien ayer entregó en la Intendencia las primeras credenciales a los examinadores que se capacitarán en las cuartas del Gran Valgarzura.

La actividad fue organizada por el "Examen de la Vida en la montaña, que duraba 5 minutos y se decidía el resultado o no". La diferencia con el desarrollo de exámenes se dividirá en dos etapas de conducción: una libre y una guiada. La prueba será complementada con un taller de motivación de estudio que estará disponible para los profesores. El objetivo de conducir clase a través de la página www.comunad.com.

Como en videos que muestran manifestaciones que se conocen al portante, es

Destacan rol porteno en desarrollo de la ciencia

Allamamos de último a primer medio abstinente a la conferencia inaugural del VII Congreso de Bioética, que organizamos en conjunto el Centro de Bioética y el

El congreso se inició con la charla "Electricidad, luz y vida", que dio cuenta sobre la importancia de la electricidad en los seres vivos.

El sector de la Universidad de Valparaíso, Aldo Valle, destacó el rol de la ciencia y tecnología en la transformación de la universidad a través de sus actividades científicas que como modelo (dijó) los avances, asociados al conocimiento a la comunidad. "Es un orgullo contar con los académicos que se dedican a la neurociencia en la Universidad de Valparaíso".

La conferencia estuvo a

El Presidente

El rector de la Universidad de Valparaíso, Aldo Valle, destacó el rol de la ciencia y cómo la universidad a través de sus científicos tiene como misión difundir los avances, acercando el conocimiento a la comunidad. "Es un orgullo contar con los académicos que se dedican a la neurociencia en la Universidad de Valparaíso", expuso.

Title: “With a magic session ended the last Tertulias Porteñas of this 2013”

Media: www.uv.cl

Date: December 20th, 2013

Ciclo de charlas del CINV de la Universidad de Valparaíso

Con mágica jornada terminó ciclo 2013 de Tertulias Porteñas

Última charla convocó a cerca de 200 personas en el Parque Cultural de Valparaíso.

20

Me gusta

Twitter 2



El ilusionismo complementó las visiones de la ciencia y el arte en la última jornada del ciclo 2013 del ya tradicional encuentro Tertulias Porteñas, que organiza el Centro Interdisciplinario de Neurociencia de la Universidad de Valparaíso y que en esta ocasión abordó, desde las distintas miradas, el tema de la percepción sensorial.

En una inédita jornada Tertulias Porteñas reunió al mago Juan Esteban Varela, quien ha destacado a nivel mundial por su innovador acto de magia para ciegos; al científico Pedro Maldonado, quien se ha concentrado en estudiar la percepción visual a través de mecanismos de la percepción sensorial, y al reconocido escultor Francisco Gazitúa, autor de numerosas obras que ornamentan modernas edificaciones, parques y plazas de Chile y el extranjero.



IMÁGENES

Reinventándose

La última jornada de Tertulias Porteñas del ciclo 2013, moderada por el editor Ernesto Pfeiffer, quien en esta versión reemplazó a Cristián Warnken, reveló el espíritu de sus organizadores por reinventarse e innovar en la puesta en escena, esta vez incorporando apoyo audiovisual, para contextualizar el trabajo desarrollado por el neurobiólogo Pedro Maldonado y las esculturas de Francisco Gazitúa, cuyo sello es la utilización de materiales autóctonos, como témpanos de hielo de la Antártica y rocas de la precordillera, que se sumaron a la imagen en vivo, en pantalla gigante, de los sorprendentes juegos de magia de Juan Esteban Varela.

Este último dijo estar maravillado con la experiencia: “¿Sabes lo que más me gusta? Es la fusión de puntos de vistas. Cuando uno piensa en el aprendizaje, en la palabra universidad, es justamente esa universalidad, una manera de enriquecer el conocimiento muy profunda y maravillosa. El hecho de que la magia, que yo hago con locura desde los seis años, esté presente en esta integración de puntos de vista para mí es un honor y un privilegio. Y además que la gente ha sido muy cariñosa”, afirmó.

Con respecto a cómo influiría esta experiencia en el arte, afirmó que “es una fuente constante de inspiración”. Además, fundamentó: “A veces es simplemente un disparo inspirador, otras veces el conocimiento duro. Está demostrado científicamente que tenemos mala capacidad para enfocar el movimiento; es una conclusión importante para un mago, qué bueno que esté demostrado científicamente, ya lo sospechábamos, pero de alguna manera son cosas que son más a nivel inspiracional y otras de nivel más técnico. Pero sobre todo la mayor riqueza viene a nivel humano, a nivel de las conexiones que estás haciendo, las conversaciones que vas teniendo, eso te va enriqueciendo. Si te enriquece como persona te va a enriquecer como profesional. Sin ninguna duda”.

Title: “Squid is used to study the epilepsy and muscular paralysis”

Media: www.24horas.cl

Date: December 20th, 2013

24 HORAS > TENDENCIAS > Ciencia

Imagen



Con jibias chilenas estudian la epilepsia y parálisis muscular

El doctor Miguel Holmgren, chileno radicado en EE.UU, explicó que estos calamares poseen los axones más grandes entre todos los animales.

Mediante el estudio de **jibias** que habitan en la **región de Valparaíso**, científicos **chilenos** indagan en las propiedades eléctricas de las neuronas y la importancia de este impulso para múltiples funciones del organismo, entre ellas, el **movimiento muscular**.

“La electricidad es generadora de vida”, señala Miguel Holmgren, científico chileno radicado en Estados Unidos y quien participó en el Octavo Congreso de Biofísicos Latinoamericanos realizado en la ciudad porteña. Esta cualidad es tan vital en el organismo de los seres vivos, que cuando existen fallas en la transmisión eléctrica, es posible que ocurran algunos tipos de epilepsia, parálisis musculares y migraña. Por esta razón, el científico explica que conocer los mecanismos de la bioelectricidad y especies como la jibia, puede contribuir al mejor entendimiento de patologías como éstas, y su manejo terapéutico”.

La exploración de este calamar gigante constituye un hito en la biofísica de Chile y el mundo. Esto, debido a que el animal, junto con estar “lleno de electricidad”, posee los axones más grandes de la naturaleza -de 1 mm aproximadamente-, los que pueden ser percibidos por el ojo humano.

•Dicha característica, el interés de diversos investigadores cómo se transmite el impulso nervioso . “Una vez generada la señal eléctrica, las neuronas tienen la propiedad de poder conducirla a través de prolongaciones llamadas axones las que, modo de , serían los cables del circuito eléctrico”, comenta el Dr. Holmgren, quien además es Presidente de la Sociedad de Biofísicos Latinoamericanos –

Media: www.aricahoy.cl
Date: October 16th, 2013

ESTUDIO CIENTÍFICO A JIBIAS

Escrito Por Administrador En Octubre 16th, 2013 05:51 PM | Destacado, Nacional, Región



Investigan propiedades eléctricas de las neuronas y la importancia de este impulso para múltiples funciones del organismo, entre ellas, el movimiento muscular.



Mediante el estudio de jibias, calamar que habita en nuestras costas, científicos chilenos, investigan las propiedades eléctricas de las neuronas y la importancia de este impulso para múltiples funciones del organismo, entre ellas, el movimiento muscular. Este trabajo está siendo desarrollado en la zona de Montemar, comuna de Concón (Valparaíso), por especialistas del Centro Interdisciplinario de la Neurociencia, CINV, con apoyo del científico chileno radicado en Estados Unidos, Dr. Miguel Holmgren. "La electricidad es generadora de vida. Esta cualidad es tan vital en el organismo de los seres vivos, que cuando existen fallas en la transmisión eléctrica, es posible que ocurran algunos tipos de epilepsia, parálisis musculares y migraña" señaló el especialista.

Por esta razón, el científico explica que conocer los mecanismos de la bioelectricidad y especies como la jibia, puede contribuir al mejor entendimiento de patologías como éstas, y su manejo terapéutico.

La exploración de este calamar gigante constituye un hito en la biofísica de Chile y el mundo. Esto, debido a que el animal, junto con estar "lleno de electricidad", posee los axones más grandes de la naturaleza -de 1 mm aproximadamente-, los que pueden ser percibidos por el ojo humano.

Title: "Squid will allows study the epilepsy and muscular paralysis"

Media: www.lanacion.cl

Date: October 19th, 2013



Inicio » Tecnología » Ciencia

19/10/2013 | ENVIAR | IMPRIMIR

JIBIAS "ELÉCTRICAS" PERMITIRÁN ESTUDIAR EPILEPSIA Y PARÁLISIS MUSCULAR



La exploración de este calamar gigante constituye un hito en la biofísica de Chile y el mundo. Esto, debido a que el animal, junto con estar "lleno de electricidad", posee los axones más grandes de la naturaleza -de 1 mm aproximadamente-, los que pueden ser percibidos por el ojo humano. La investigación es llevada a cabo por el Centro Interdisciplinario de Neurociencia de Valparaíso.

Sábado 19 de octubre de 2013 | por Nacion.cl + Sigue a Nación.cl en Facebook y Twitter

Mediante el estudio de jibias que habitan en la Región de Valparaíso, científicos chilenos indagan las propiedades eléctricas de las neuronas y la importancia de este impulso para múltiples funciones del organismo, entre ellas, el movimiento muscular.

"La electricidad es generadora de vida", señala Miguel Holmgren, científico chileno radicado en Estados Unidos - y quien participó en el Octavo Congreso de Biofísicos Latinoamericanos, realizado en la ciudad porteña-.

Esta cualidad es tan vital en el organismo de los seres vivos, que cuando existen fallas en la transmisión eléctrica, es posible que ocurran algunos tipos de epilepsia, parálisis musculares y migraña. Por esta razón, el científico explica que conocer los mecanismos de la bioelectricidad y especies como la jibia, puede contribuir al mejor entendimiento de patologías como éstas, y su manejo terapéutico.



La exploración de este calamar gigante **constituye un hito en la biofísica de Chile y el mundo**. Esto, debido a que el animal, junto con estar “lleno de electricidad”, **posee los axones más grandes de la naturaleza -de 1 mm aproximadamente-**, los que pueden ser percibidos por el ojo humano.

Dicha característica captó el interés de diversos investigadores tanto en Europa como en EEUU y fueron estos axones los que permitieron dilucidar **cómo se transmite el impulso nervioso**, estudios que le dieron el Premio Nobel a los Ingleses, Alan Hodgkin y Andrew Huxley en Medicina y Fisiología el año 1962.

“Una vez generada la señal eléctrica, las neuronas tienen la propiedad de poder conducirla a través de sus prolongaciones llamadas axones, las que de un modo de gráfico serían los cables del circuito eléctrico”, comenta el doctor Holmgren, quien además es Presidente de la Sociedad de Biofísicos Latinoamericanos (SOBLA).

Respecto a cómo sucede este proceso, los expertos determinaron que se debe a la **permeabilidad de la membrana celular**, la cual contiene unas proteínas que permiten el flujo constante de cargas eléctricas, desde el interior y hacia el exterior de la célula.

ELECTRICIDAD, MOTOR DE VIDA

Respecto al poder de la electricidad, el biofísico explica que **cada célula del cuerpo es una especie de batería eléctrica de aproximadamente 0.1 voltios** y cuyo polo negativo se ubica al interior de la célula.

Gracias a esto, se generan una serie de eventos, desde la ocurrencia de una idea, hasta el movimiento. “Además, la mayoría de las células usan la electricidad para alimentarse y descartar toxinas, entre otras funciones. Asimismo, las plantas también utilizan electricidad para poder sobrevivir”.

Title: “Green Building turns schoolchild into small Scientists ”

Media: www.lanacion.cl

Date: October 20th, 2013



Inicio » Tecnología » Ciencia

20/10/2013 | ENMAR | IMPRIMIR

EDIFICIO VERDE CONVIERTE A ESCOLARES EN PEQUEÑOS CIENTÍFICOS



Alumnos de escuelas públicas de Valparaíso podrán experimentar en laboratorios y aprender sobre neurociencia, genética y tecnologías sustentables, con estudiantes de Doctorado en Neurociencia de la Universidad de Valparaíso.

Domingo 20 de octubre de 2013 | por Nación.cl + Sigue a [Nación.cl](https://www.facebook.com/Nacion.cl) en Facebook y Twitter

Una casa especialmente equipada con tecnología sustentable y laboratorios, es el nuevo proyecto que tendrá por protagonistas a niños de escuelas vulnerables en Valparaíso.

Se trata del Edificio Verde de [Ciencia Al Tiro](#), iniciativa dirigida por la doctora Kathleen Whitlock, del Centro Interdisciplinario de Neurociencia de Valparaíso, que busca enseñar y acercar la ciencia de forma cercana y entretenida.

Este lugar, una antigua casona del Cerro Playa Ancha que fue refaccionada, abrió sus puertas para que alumnos, principalmente de séptimo y octavo básico, puedan experimentar y conocer aspectos de la neurociencia, la genética, la energía inteligente y la acuaponía: un sistema sustentable de producción de plantas acuáticas y peces.

CIENCIA PARA TODOS

Según explica Kathleen Whitlock, neurobióloga norteamericana, el proyecto responde a un “fuerte compromiso social que busca apoyar a niños de escasos recursos”, incentivándoles a aprender ciencia de manera didáctica, y generando oportunidades para desarrollar sus capacidades.

“Nos interesa transmitir la idea de que la ciencia está en todas partes y es entretenida. Además, creemos que ésta puede ser útil en sus vidas”, comenta.

Al interior de este edificio también, se permitirá la realización de charlas y talleres que promuevan el intercambio de ideas a nivel local, nacional e internacional. Esto, a fin de continuar y profundizar la tarea ya iniciada a partir del 2008 por [Ciencia Al Tiro](#), cuando se dio curso a las actividades con la Escuela Básica Pacífico y la Escuela Árabe República de Siria.

El programa [Ciencia al Tiro](#) es reconocido por la Universidad de Valparaíso como parte de la formación de los estudiantes de doctorado, incluyendo créditos de la malla curricular.

Title: "Chilean Squid is used to study the epilepsy and muscular paralysis"

Media: El Día de la Serena, Newspaper

Date: October 18th, 2013



INVESTIGACIÓN DEL CENTRO INTERDISCIPLINARIO DE LA NEUROCIENCIA

Electricidad de jibia chilena permitirá estudiar epilepsia y parálisis muscular



✿ **Exploración de este calamar gigante, que también está en la Región de Coquimbo, constituye un hito en biofísica de Chile y el mundo**

La Serena

Mediante el estudio de jibias, calamar que habita en nuestras costas, científicos chilenos, se encuentran indagando en las propiedades eléctricas de las

neuronas y la importancia de este impulso para múltiples funciones del organismo, entre ellas, el movimiento muscular.

Este trabajo está siendo desarrollado en la zona de Montemar, comuna de Concón (Valparaíso), por especialistas del Centro Interdisciplinario de la Neurociencia, con apoyo del científico chileno radicado en Estados Unidos, Dr. Miguel Holmgren.

"La electricidad es generadora de vida. Esta cualidad es tan vital en el organismo de los seres vivos, que cuando existen fallas en la transmisión

eléctrica es posible que ocurran algunos tipos de epilepsia, parálisis musculares y migraña" señaló el especialista.

Por esta razón, el científico explica que conocer los mecanismos de la bioelectricidad y especies como la jibia puede contribuir al mejor entendimiento de patologías como éstas, y su manejo terapéutico.

La exploración de este calamar gigante constituye un hito en la biofísica de Chile y el mundo. Esto, debido a que el animal, junto con estar "lleno de electricidad", posee los axones más grandes de la naturaleza de 1 mm aproximadamente, los que pueden ser percibidos por el ojo humano.

"Generada la señal eléctrica, las neuronas tienen la propiedad de poder conducirla a través de sus prolongaciones llamadas axones las que, de un

modo de gráfico, serían los cables del circuito eléctrico", comenta el Dr. Holmgren, quien, además, es presidente de la Sociedad de Biofísicos Latinoamericanos.

Respecto a cómo sucede este proceso, los expertos determinaron que se debe a la permeabilidad de la membrana celular, la cual contiene unas proteínas que permiten el flujo constante de cargas eléctricas, desde el interior y hacia el exterior de la célula.

Dicha característica, captó el interés de diversos investigadores tanto en Europa como en los EE.UU. y fueron estos axones los que permitieron dilucidar cómo se transmite el impulso nervioso, estudios que le dieron el Premio Nobel a dos ingleses, Alan Hodgkin y Andrew Huxley en Medicina y Fisiología el año 1962. ¹⁰⁰⁴

FOTO: CARLOS VALLANZANO
Científicos buscan conocer los mecanismos de la bioelectricidad. Especies como la jibia son claves.

Title: "Squid whet the appetite of Scientifics: It is used to study the epilepsy "

Media: La Cuarta, Newspaper

Date: October 25th, 2013

18 **IPATS**

Viélenos en www.lacuarta.com

Viénes 25 de octubre de 2013 **La Cuarta**



La jibia les abrió el apetito a los científicos: sirve para la epilepsia

Por **Nicole Salvatierra**

Chilenos cacharon que, al estudiarla, se entienden enfermedades nerviosas

Aunque cuando la cocinan deja la casa *pa'rd* a mariscos, la jibia es una delicia marina que no sólo sirve al paladar. El tremendo calamar es tan pulento que incluso llama la atención de científicos.

Justo ahora hay unos capos chilenos estudiando los nervios de la jibia para entender y tratar enfermedades cuáticas que afectan el sistema nervioso, como la epilepsia. Todo un hito en la biofísica chilensis y del mundo.

"La electricidad es generadora de vida. Esta cualidad es tan vital en el organismo de los seres vivos que cuando existen fallas en la transmisión eléctrica, es posible que ocurran algunos tipos de epilepsia, parálisis musculares y migraña", soltó el doc Miguel Holmgren.

El especialista, radicado en Estados Unidos pero más chileno que los porotos, apoya la investigación que realizan los científicos del Centro Interdisciplinario de la Neurociencia, CINV, en la zona de Montemar, ubicado en las costas de Concón.

Ocurre que el cefalópodo está dotado con los nervios más grandes del reino animal, perceptibles al ojo humano. Por eso, la jibia sirve pa' cachar las propiedades eléctricas de las neuronas y la importancia de este impulso para múltiples funciones del organismo, entre ellas, el movimiento muscular.

Para hacerla más clarita, el biofísico explica que cada célula del cuerpo es una especie de batería eléctrica de aproximadamente 0.1 voltios, cuyo polo negativo se ubica al interior de la célula. Gracias a esto, el ser humano puede pensar y actuar. O sea, se aviva con una idea y la lleva al movimiento.

"Además, la mayoría de las células usan la electricidad para alimentarse y descartar toxinas, entre otras funciones. Asimismo, las plantas también utilizan electricidad para poder sobrevivir", versea Holmgren.

El hombre de ciencia, en todo caso, también disfruta de la jibia con mayo. ¡Obvio!

Son buenas pa'l buche

Además de ser muy ricas con mayo, las jibias sirven para entender y tratar enfermedades nerviosas como la epilepsia.

NICOLAS KUBRONOVICH

Title: “Chilean scientist discover that organism’s Protein Causes Genetics Deafness”

Media: www.24horas.cl

Date: November 15th, 2013

24
HORAS.CL

VIERNES
15 DE NOVIEMBRE

NACIONAL POLITICA ECONOMIA INTERNACIONAL DEPORTES TENDENCIAS REGIONES VIDEOS

Imagen



Chileno descubre que proteína produciría sordera

El doctor Agustín Martínez, del Centro Interdisciplinario de Neurociencia de la Universidad de Valparaíso, espera que sus hallazgos contribuyan a la creación de terapias.

POR 24HORAS.CL

  [Twitter](#) 26  [Recomendar](#) 17  [Compartir](#) 1

Title: With the Severin Building try to recover patrimonial sector of Valparaíso.

Media: www.biobiochile.cl

Date: December 12th, 2013

NACIONAL

Lunes 2 diciembre 2013 | 13:04 - Actualizado: 13:50

Buscan recuperar sector patrimonial del puerto con obras en Edificio Severín de Valparaíso

3 7 0 28
Recomendar Twittear +1 Visitas



Recuperar el sector patrimonial del barrio puerto, donde fue asaltado el experto argentino de la Unesco, es el principal objetivo de los trabajos que se realizan en el edificio Severín, que queda atrás de la iglesia La Matriz, lugar en donde funcionaría el Centro Interdisciplinario de Neurociencia de la Universidad de Valparaíso.

Este proyecto ya está terminado y fue presentado a la comisión de ICOMOS de la Unesco, compuesta por el experto argentino y brasilero, a quienes un día antes del robo se les informó la peligrosidad del sector y su necesidad de recuperarlo. Así lo dio a conocer a La Radio Ramón Latorre, director y gerente del Centro Interdisciplinario de Neurociencia de la Universidad de Valparaíso.



La latente peligrosidad en el sector, que es donde se funda Valparaíso, gatilló que los científicos presentaran esta nueva iniciativa que ya cuenta con una inversión de mil quinientos millones de pesos, por lo que se espera solicitar fondos al Gobierno Regional, puesto que la infraestructura tiene un costo total de 5 millones de pesos aproximadamente.

El edificio Severín fue elegido porque este lugar- según explicó Latorre, fue sede del primer Congreso Bicameral de Chile en 1828, inscribiéndose la Constitución de 1925, por lo cual presentaría las ventajas para generar la recuperación.

En tanto, Ramón Latorre, enfatizó que la construcción del Centro no solo es un beneficio patrimonial y cultural sino que también social porque las dependencias pueden ser utilizadas por la comunidad, entregando incluso wi-fi a un perímetro del barrio puerto.



De obtener los recursos para la ejecución, se espera que la construcción comience a mediados del 2014 para estar terminada a más tardar el 2015, donde sería un espacio para la comunidad, la ciencia y la cultura.

